

Anomia-associated bryozoans from the upper Pliocene (Piacenzian) lower Tamiami Formation of Florida, USA

Emanuela Di Martino, Paul D. Taylor, and Roger W. Portell

ABSTRACT

Commercial mining northeast of Sarasota, SW Florida (USA), since the 1960s has exposed Plio-Pleistocene marine shell beds of the Tamiami Formation, a complex sequence of mixed carbonate and siliciclastic beds containing a malacofauna that is a mixture of subtropical and temperate species. Material used for the current study consists of shells of the bivalve *Anomia simplex* d'Orbigny, 1853 encrusted by bryozoans. The shells were collected from Units 10/11 of the lower Tamiami Formation, estimated as being late Pliocene (Piacenzian). This paper describes the bryozoan fauna associated with these 'jingle shells', which is of relatively low diversity, totaling 29 species, and comprises one cyclostome and 28 cheilostomes. Six cheilostome species are new: *Micropora stellata* sp. nov., *Microporella sarasotaensis* sp. nov., *Microporella tamiamiensis* sp. nov., *Pourtalesella chiarae* sp. nov., *Spiniflabellum laurae* sp. nov., and *Trypostega composita* sp. nov. Of the previously described species, six are extant and have western Atlantic distributions, while 12 species are known only from the fossil record. The bulk of the assemblage comprises a limited number of species represented by hundreds of colonies, while the remaining species are rare and represented by a single or a few colonies. Colonization of the shells is likely to have happened post-mortem, considering the high percentage of valves encrusted on both surfaces. A large number of overgrowth interactions have been observed among the bryozoan colonies, thus this taxonomic revision is the necessary baseline for ecological analyses aiming to establish a ranking of species in a competitive hierarchy.

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Di Martino, Emanuela, Taylor, Paul D., and Portell, Roger W. 2019. *Anomia*-associated bryozoans from the upper Pliocene (Piacenzian) lower Tamiami Formation of Florida, USA. *Palaeontologia Electronica* 22.1.11A 1-65. <https://doi.org/10.26879/920> palaeo-electronica.org/content/2019/2404-tamiami-formation-bryozoans

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Keywords: Bryozoa; Cyclostomata; Cheilostomata; new species; Piacenzian; Tamiami Formation

Submission: 28 August 2018 Acceptance: 14 January 2019

INTRODUCTION

The monograph published by Canu and Bassler in 1923 is still the most comprehensive work yet published on North American Neogene and Quaternary bryozoans. It includes bryozoan faunas from Miocene to Pleistocene formations in several North American states (California, Florida, Maryland, North Carolina, South Carolina, Virginia) as well as the Caribbean (Costa Rica, Cuba, Dominican Republic, Jamaica), totaling approximately 250 species.

Among formations exposed in Florida, Canu and Bassler (1923) reported bryozoans from the lower Miocene Chipola Formation (only five species, cf. 47 in the subsequent, more comprehensive work of Sclaro (1968), and 60 in the most recent revision of Di Martino et al. (2017)), the upper Miocene Choctawhatchee Formation (now assigned to the upper Pliocene Jackson Bluff Formation; 22 species), and the lower Pleistocene Caloosahatchee Formation (23 species).

No information was available on the bryozoan fauna from the Plio-Pleistocene Tamiami Formation until the unpublished MSc dissertation of Echols (1960) that aimed to describe this previously unknown bryozoan fauna and to analyze its ecological and stratigraphical significance. Echols (1960) reported 26 species, including 25 cheilostomes and one cyclostome, from units exposed on the west side of State Road 29 at Sunniland in Collier County, corresponding to the lower Tamiami Formation of Zullo and Harris (1992). Unfortunately, Echols (1960) provided no images of the species described, precluding any significant comparisons with the assemblage studied here.

After Echols' work, Knowles (2008) reported bryozoans from the Tamiami Formation (upper units) in a paper revising three species of the genus *Floridina* from Neogene Coastal Plain deposits of North America. In addition, Darrell and Taylor (1989) reported bryozoans associated with coral symbionts of hermit crabs from the so-called 'Pinecrest Beds', which represent the upper Tamiami Formation in modern terminology.

This present paper aims to describe the taxonomy of the bryozoans associated with newly collected *Anomia*-shells from the late Pliocene lower Tamiami Formation (units 10/11). We add new

bryozoan records for this formation, introduce six new species, and provide improved descriptions, illustrations, and taxonomy for existing species. The taxonomic baseline necessary for an ecological study in preparation of competition for substrate space is thus established.

MATERIALS AND METHODS

Material used for this study was collected in July 2017 by two of us (EDM and RWP) from Phase 10 excavations (now water-filled) of the SMR Aggregates quarries, situated northeast of Sarasota, Florida (27.373907°, -82.376859°; WGS84) (Figure 1). Excavations exposed the marine shell-rich Plio-Pleistocene beds of the Tamiami Formation (Figure 2), a complex sequence containing mixed carbonate and siliclastic beds. Based on Zullo and Harris (1992), the Tamiami Formation is divided into upper and lower parts. The upper Tamiami Formation is subdivided into upper Pinecrest beds and lower Pinecrest beds with predominantly tropical to temperate marine mollusks and corals in unconsolidated sediments deposited at water depths of less than 30 m (Allmon et al., 1993). The lower Tamiami Formation is characterized by a mixture of subtropical and temperate species, but the fauna appears to be leached of most of its aragonitic shells.

We picked 867 shells (left valves) of the bivalve *Anomia simplex* d'Orbigny, 1853, commonly known as 'jingle shells' (Figure 3), visibly encrusted by bryozoans, from Units 10/11 of the

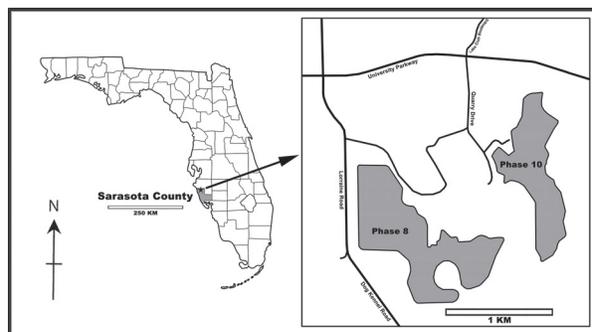


FIGURE 1. Location of the collecting locality (Phase 10 excavations of the SMR Aggregates quarries), northeast of Sarasota, Florida.

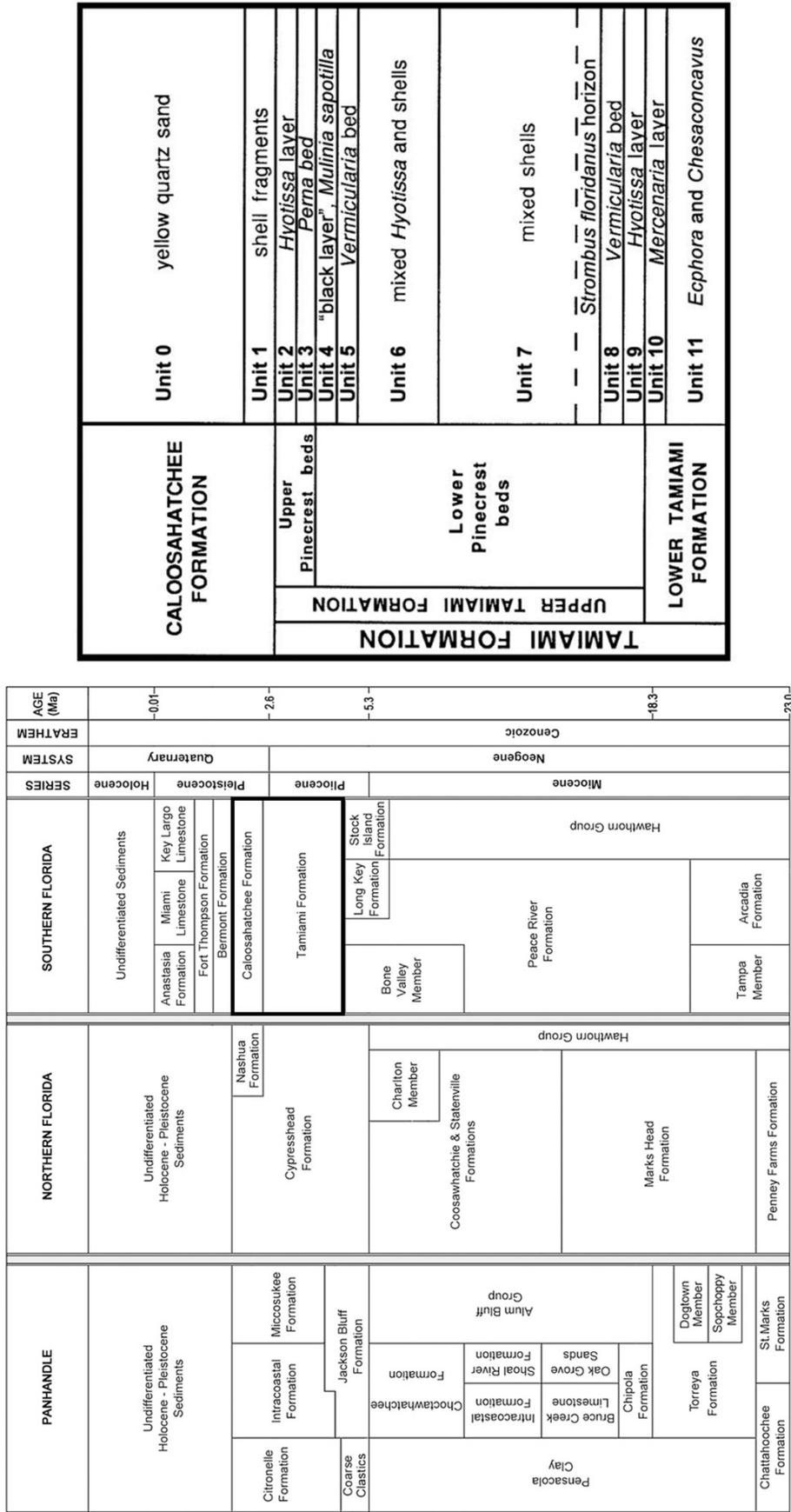


FIGURE 2. Overview of the Neogene and Quaternary stratigraphic units in Florida (modified slightly from Bryan and Means, 2014, figure 4), and detailed stratigraphic nomenclature of the Tamiami Formation units used for the APAC and SMR Aggregates quarries in Sarasota and Manatee Counties taken from Zullo and Harris (1992) incorporating and slightly modifying the APAC stratigraphic units of Petuch (1982).



FIGURE 3. *Anomia simplex* d'Orbigny, 1853

lower Tamiami Formation (Petuch, 1982; Zullo and Harris, 1992), estimated as being 3.0 (± 0.5) Ma and corresponding to a late Pliocene (Piacenzian) age, based on $^{87}\text{Sr}/^{86}\text{Sr}$ isotope dating of bivalves, paleomagnetism, and invertebrate and vertebrate biochronology (Jones et al., 1991). Before examining the encrusting bryozoan colonies, the shell substrates were gently cleaned under running water, scrubbed with a soft toothbrush, washed in an ultrasonic bath to remove sediment, and air-dried. Scanning electron microscopy (SEM) was conducted on the best-preserved specimens. SEM observations were made on uncoated specimens using a low-vacuum scanning electron microscope (LEO VP-1455) at the Natural History Museum in London (NHMUK). Zooidal measurements were taken from SEM images using the image processing program ImageJ (available at <https://imagej.nih.gov/>). Each measurement is given in the text as the mean value in $\mu\text{m} \pm$ the standard deviation, observed range, number of specimens used, and total number of measurements (the latter two values enclosed in parentheses). Abbreviations for the measurements are: ApL, aperture length; ApW, aperture width; AvL, avicularium length; AvW, avicularium width; AvOpL, avicularium opesia length; AvOpW, avicularium opesia width; FWL, frontal wall length; FWW, frontal wall width; OL, orifice length; OW, orifice width; OWmin, minimum orifice width; OWmax, maximum orifice width; OvL, ovicell length; OvW, ovicell width; ZcL, zoeciule length; ZcW, zoeciule width; ZL, autozooid length; and ZW, autozooid width.

For taxonomic comparisons, the holotypes of *Micropora robusta* Cook, 1985 (NHMUK 1972.3.3.1) and *Cellepora umbilicata* Lonsdale,

1845 (NHMUK D53195) were studied. Through the kindness of JoAnn Sanner (USNM), we were able to obtain new SEM images of the holotypes of *Amphiblestrum constrictum* Ulrich and Bassler, 1904 (USNM 68459), *Amphiblestrum tenuiparietis* Canu and Bassler, 1923 (USNM 68460); syntypes of *Cyclocolposa perforata* Canu and Bassler, 1923 (USNM 68617, two specimens), *Cycloperiella rubra* Canu and Bassler, 1923 (USNM 68620, two specimens), *Smittina maleposita* Canu and Bassler, 1923 (USNM 68641), *Stephanollina vorax* (Canu and Bassler, 1923) (USNM 651310 and USNM 651312); hypotype of *Micropora coriacea* (Esper, 1791) *sensu* Canu and Bassler (1923) (USNM 68480); plesiotypes of *Metroperiella reversa* (Ulrich and Bassler, 1904) (USNM 68651) and *Stephanosella biaperta* Michelin, 1842 *sensu* Canu and Bassler (1923) (USNM 68544 and USNM 68546). All figured and type specimens from the Tamiami Formation are registered in the paleontological collections of the Invertebrate Paleontology Division, Florida Museum of Natural History, University of Florida (acronym UF).

SYSTEMATIC PALEONTOLOGY

Order Cyclostomata Busk, 1852
 Suborder Tubuliporina Milne Edwards, 1838
 Family Oncousoeciidae Canu, 1918
 Genus *Oncousoecia* Canu, 1918
Oncousoecia sp.

Figure 4

Figured material. UF 305758 (Shell 111). Pliocene, lower Tamiami Formation.

Description. Colony encrusting, low, flat, oligoserial, ribbon-like (Figure 4.1–2). Autozooids distinct, bordered by faint furrows, sinuous, elongate, with isolated, transversely elliptical apertures and short peristomes (Figure 4.3). Frontal wall with thick, transverse, undulose wrinkles and circular pseudopores less than 5 μm in diameter (Figure 4.4). Gonozooid transversely elongate, 340 μm long by 430 μm wide, not penetrated by autozooidal peristomes (Figure 4.3, 5); ooeciostome terminal, short; ooeciopore transversely elliptical, 30 μm long by 70 μm wide (Figure 4.6). Distal fringe of basal lamina including 2–4 rows of kenozooids (Figure 4.3, 5). Terminal diaphragms present, closing the apertures of some of the oldest autozooids (Figure 4.6). Protooecium and ancestrula not observed.

Measurements (μm). FWL 310 \pm 42, 258–378 (1, 10); FWW 138 \pm 20, 118–176 (1, 10); ApL 112 \pm 11, 94–128 (1, 8); ApW 77 \pm 8, 62–88 (1, 8).

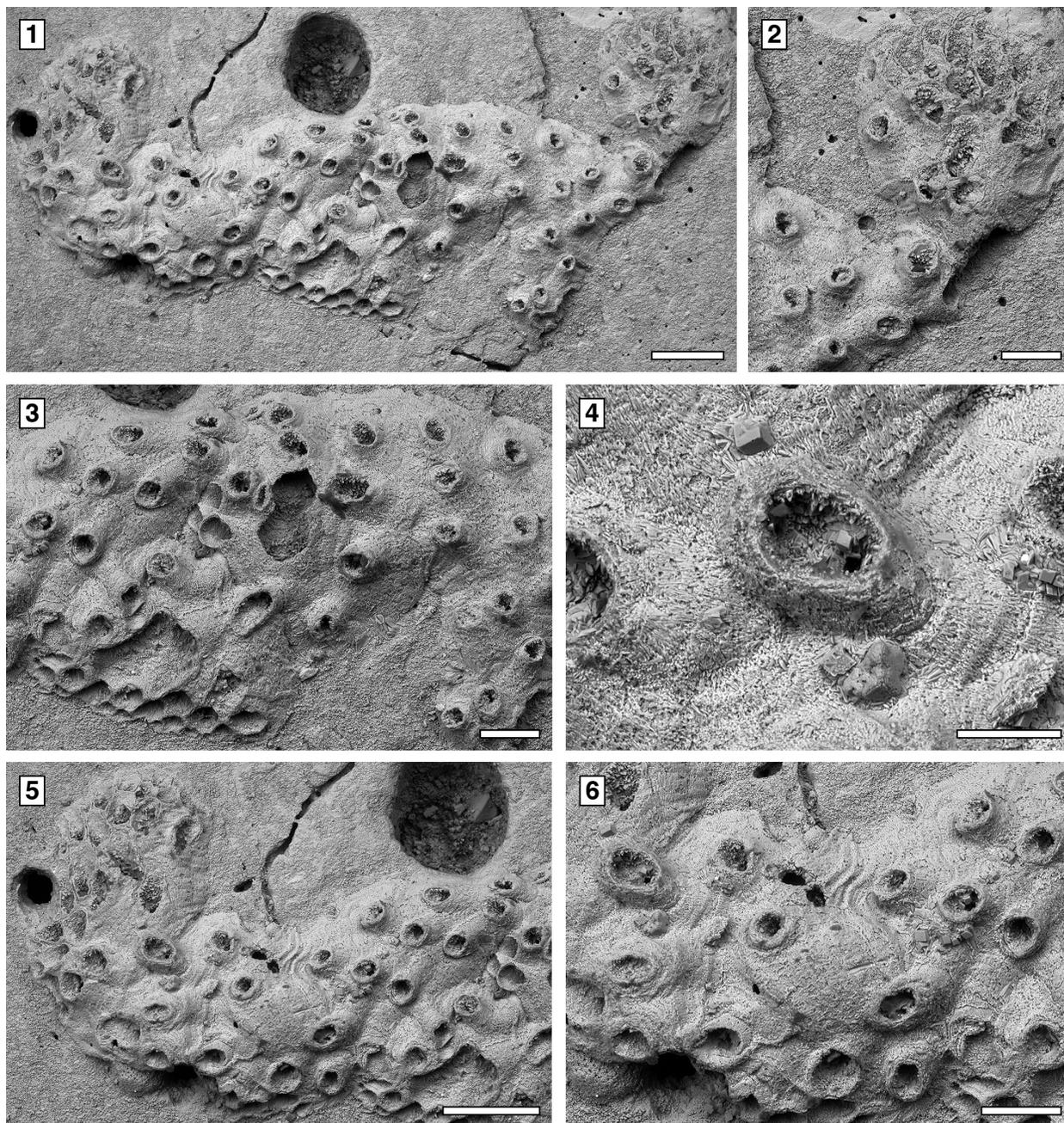


FIGURE 4. *Oncousoecia* sp., UF 305758 (Shell 111), Pliocene, lower Tamiami Formation Units 10/11. **1**, general view of the ribbon-like colony (400 μ m). **2**, close-up of the left lobe (200 μ m). **3**, part of the colony including several autozooids, a broken gonozooid and basal lamina with two rows of kenozooids (200 μ m). **4**, close-up of an autozooid with elliptical aperture and wrinkled, pseudoporous frontal wall (100 μ m). **5**, part of the colony with complete gonozooid and oocypore and distal fringe of basal lamina including several rows of kenozooids (400 μ m). **6**, close-up of the gonozooid (200 μ m).

Remarks. *Oncousoecia* sp. is the only cyclostome species present in our sample. Species-level identification is difficult on the basis of a single colony, given the paucity and high variability of skeletal characters among cyclostomes.

Order Cheilostomata Busk, 1852
 Superfamily Membraniporoidea Busk, 1852
 Family Membraniporidae Busk, 1852
 Genus *Acanthodesia* Canu and Bassler, 1919
Acanthodesia cf. *denticulata* (Busk, 1856)
 Figure 5

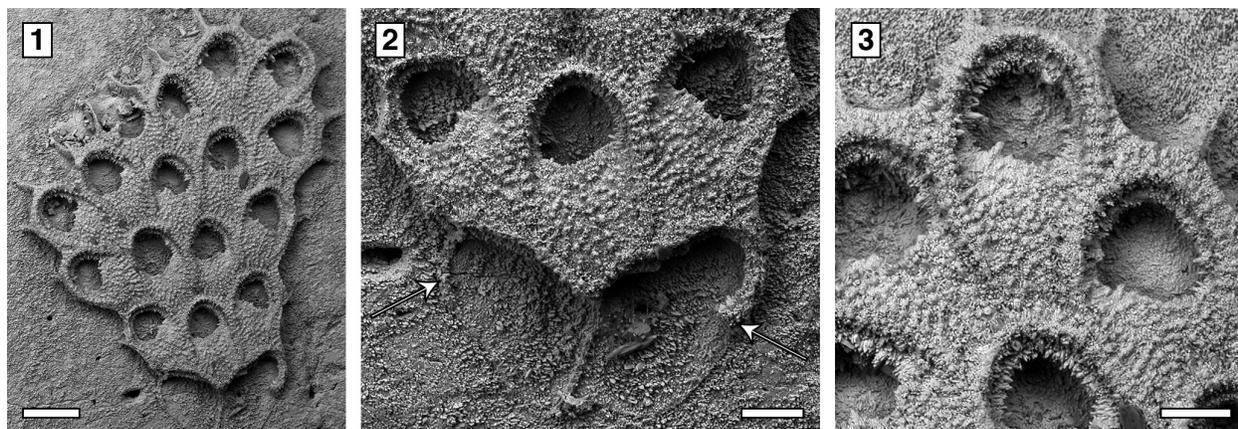


FIGURE 5. *Acanthodesia* cf. *denticulata* (Busk, 1856), UF 305759 (Shell 370), Pliocene, lower Tamiami Formation Units 10/11. **1**, general view of the small colony available (200 µm). **2**, close-up of the outline of the twinned ancestrula (see arrows) and first budded zooids (100 µm). **3**, close-up of autozooids with cryptocystal spine-like projections in the opesia (100 µm).

cf. 1856 *Membranipora denticulata* Busk, p. 176, pl. 7, figures 1, 2.

cf. 1873 *Biflustra denticulata* Smitt, p. 18, pl. 4, figures 89–91.

cf. 1923 *Hemiseptella filimargo* Canu and Bassler, p. 71, pl. 10, figure 9.

cf. 1928a *Hemiseptella denticulata* Canu and Bassler, p. 62, pl. 9, figure 9.

cf. 1928a *Hemiseptella hexagonalis* Canu and Bassler, p. 63, pl. 28, figure 9.

cf. 2005 *Biflustra denticulata* Winston, p. 6, figures 1–3.

Figured material. UF 305759 (Shell 370). Pliocene, lower Tamiami Formation.

Description. Colony encrusting, multiserial, unilaminar, comprising only 15 zooids up to the fourth generation (including early astogeny) and the outline of the ancestrular complex (Figure 5.1). Ancestrula twinned, with zooids adjacent to each other for about half of their length, then diverging outwards at an angle of 90°, 316–360 µm long by 198–210 µm wide (Figure 5.2, see arrows); a single autozooid budded laterally from both ancestrular zooids and placed centrally, two autozooids budded distolaterally, one from each ancestrular zooid. Autozooids distinct, boundaries marked by a fine fissure and also by a beaded mural rim distally, arranged quincuncially, subrectangular to club-shaped with rounded distal margin and concave proximal margin, elongate (mean L/W 1.76). Mural rim salient distally but indistinct elsewhere. Gymnocyath absent. Cryptocyst very narrow distally, extensive proximally occupying about two-thirds of the zooidal surface, flat to slightly convex proximal to the opesia, pustulose. Opesia reduced to one-third

of the zooidal frontal surface, roughly transversely D-shaped and equidimensional with a variable number of cryptocystal spine-like projections (Figure 5.3).

Measurements (µm). ZL 344±28, 292–388 (1, 10); ZW 195±10, 179–213 (1, 10); OL 124±5, 114–128 (1, 7); OW 125±4, 120–132 (1, 7).

Remarks. The Tamiami Formation species is very similar to the recent *Acanthodesia denticulata* in its general appearance, but differs from this species in having smaller autozooids and in the lack of gymnocyath tubercles on the proximal corners of the zooids. However, both of these differences may be due to the small size of the colony, which is limited to the early astogenetic stages. The illustration of *Hemiseptella filimargo* Canu and Bassler, 1923 from the Pliocene of Virginia shows what may be the same species. We prefer to use the genus *Acanthodesia* Canu and Bassler, 1919 rather than *Biflustra* d'Orbigny, 1852 because of the membraniporiform encrusting colonies, the type species of *Biflustra* having an erect vincularian colony (see also Taylor and Tan, 2015; Di Martino et al., 2017; Di Martino and Taylor, 2018).

Acanthodesia sp.

Figure 6

Figured material. UF 305760 (Shell 500). Pliocene, lower Tamiami Formation.

Description. Colony encrusting, multiserial, unilaminar, fan-shaped (Figure 6.1). Autozooids distinct, boundaries marked by a fine fissure and also by a beaded mural rim distally, arranged in alternating, parallel rows, subrectangular to club-shaped with rounded distal margin and straight to

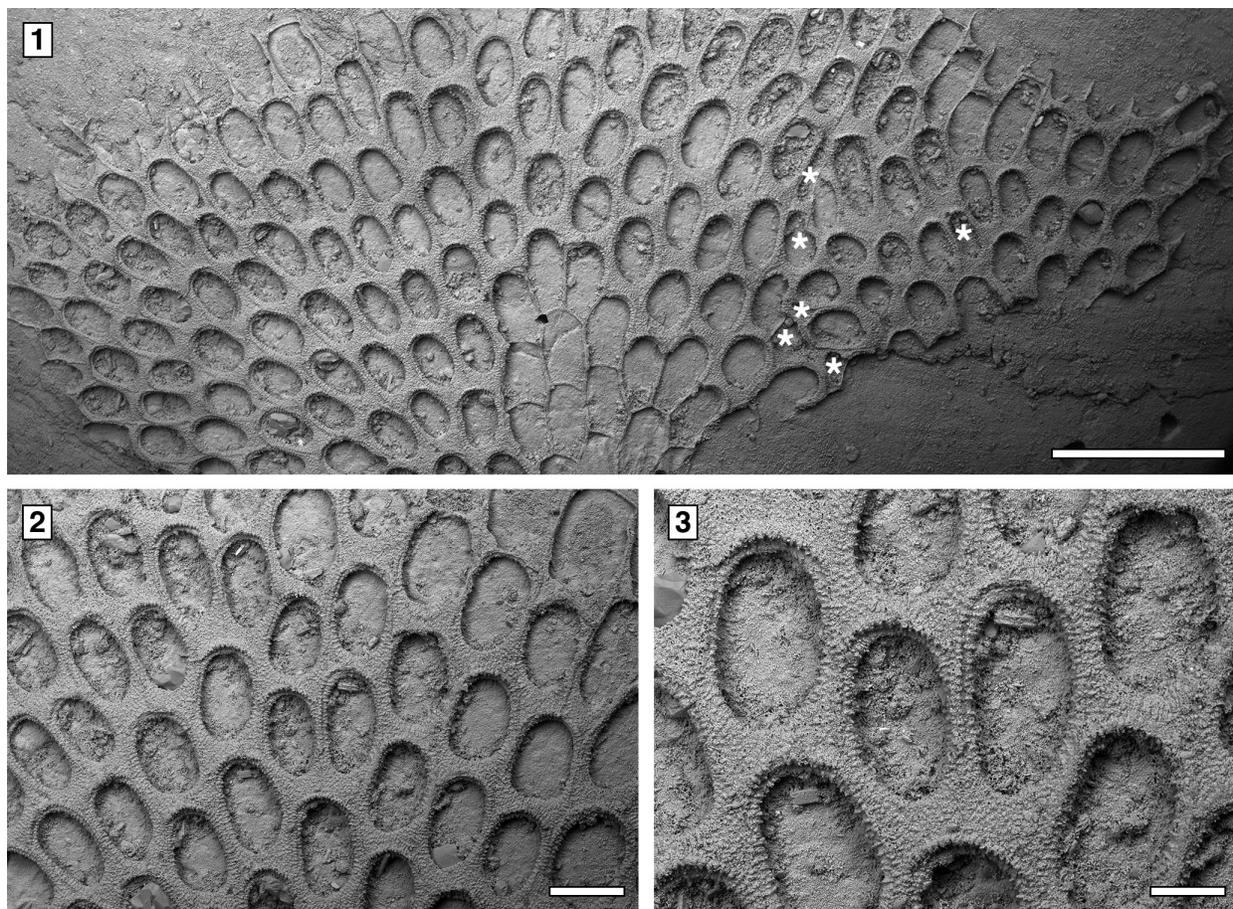


FIGURE 6. *Acanthodesia* sp., UF 305760 (Shell 500), Pliocene, lower Tamiami Formation Units 10/11. **1**, general view of the extensive sheet-like colony (2 mm), with asterisks indicating kenozooids and/or irregularly shaped autozooids forming at the edges of abutting lobes/colonies. **2**, group of autozooids (400 µm). **3**, close-up of slender autozooids placed at row bifurcation (200 µm).

concave proximal margin, elongate (mean L/W 1.46) (Figure 6.2). Autozooids situated immediately before row bifurcations similar in size to autozooids elsewhere; autozooids at row bifurcations slender (mean L/W 1.91) (Figure 6.3). Mural rim salient only distally. Gymnocyst absent. Cryptocyst very narrow distally and laterally, gently sloping down towards the opesia, slightly broader proximally and up to one-quarter of zooidal length, flat to slightly concave, pustulose, with pustules encircling the opesia aligned in radial rows and sometimes minute denticles projecting into it; proximomedial plate absent. Opesia ovoidal to elliptical, occupying nearly all frontal surface (Figure 6.3). Kenozooids and/or irregularly shaped autozooids forming at the edges of abutting colonies (Figure 6.1, see asterisks).

Measurements (µm). ZL 532±75, 445–716 (2, 20); ZW 366±30, 318–416 (2, 20); OL 406±39, 345–

492 (2, 20); OW 266±36, 204–330 (2, 20); ZL* 621±106, 444–804 (2, 10); ZW* 325±48, 250–379 (1, 10). *Zooids located at row bifurcations.

Remarks. The paucity of morphological characters makes the taxonomy of this genus particularly difficult. In previous works, fossil species from North America were frequently attributed to different varieties of *Acanthodesia savartii* (Audouin, 1826), whose type material is missing. The Tamiami Formation species is distinguished from known Cenozoic North American congeners in having larger zooids and also autozooids located immediately before row bifurcations similar in size to autozooids from elsewhere in the colony. Indeed, most species of *Acanthodesia* have exceptionally larger zooids preceding row bifurcations. However, *Acanthodesia oblongula* (Ulrich and Bassler, 1904) also lacks large zooids at row bifurcations (see Canu and Bassler, 1923, pl. 10, figures 1–3).

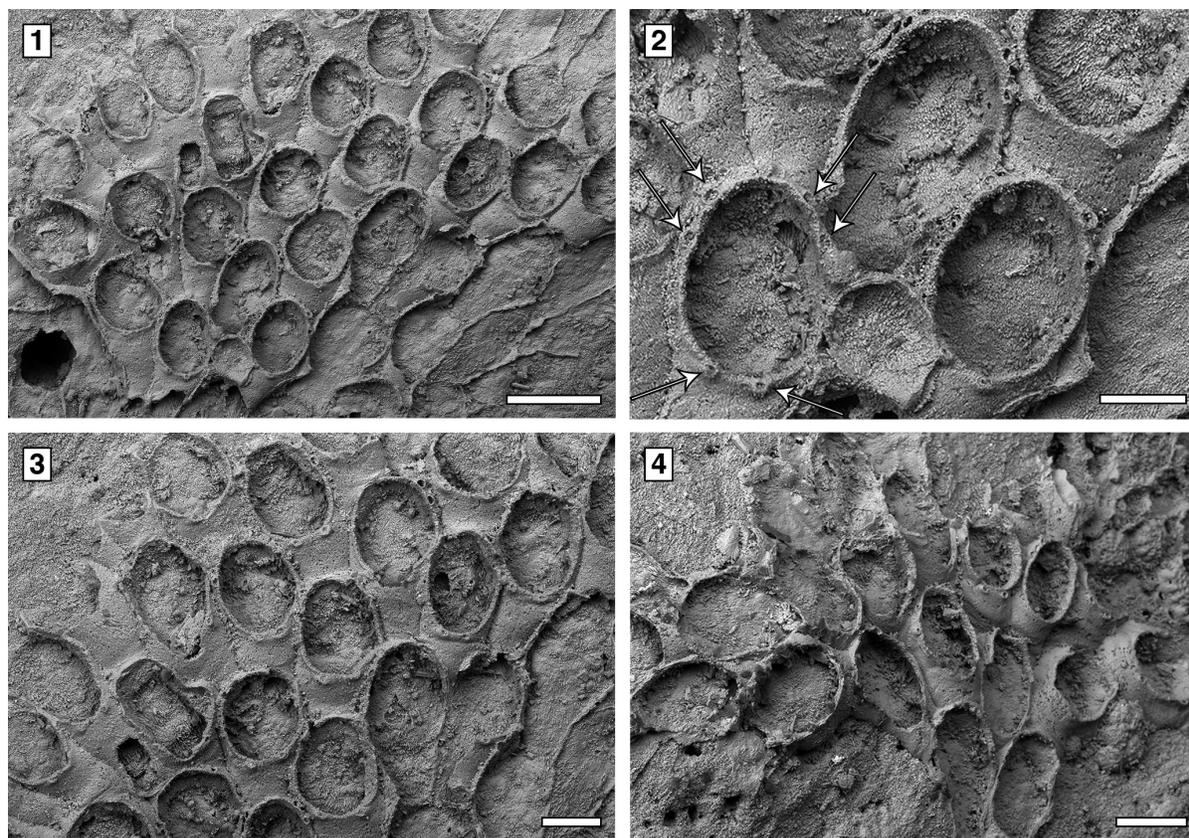


FIGURE 7. *Osburnea* aff. *biscuta* (Osburn, 1950), UF 305761a, b (Shell 374), Pliocene, lower Tamiami Formation Units 10/11. **1**, general view of the small colony (a) (400 µm). **2**, close-up of autozooids with arrows showing two proximomedial and four distolateral spine bases (200 µm). **3**, group of autozooids including those at row bifurcation (200 µm). **4**, tilted view of small, fan-shaped colony (b) (200 µm).

Family Electridae d'Orbigny, 1851
 Genus *Osburnea* Nikulina, 2010
Osburnea aff. *biscuta* (Osburn, 1950)
 Figure 7

aff. 1950 *Electra biscuta* Osburn, p. 37, pl. 3, figures 7, 8.

Figured material. UF 305761a, b (two small colonies) (Shell 374). Pliocene, lower Tamiami Formation.

Description. Colony encrusting, multiserial, unilaminar, forming fan-shaped patches (Figure 7.1, 4). Autozooids distinct with deep interzooidal grooves, quincuncially arranged, club-shaped, elongate (mean L/W 1.73) (Figure 7.2–3). Gymnocyist smooth, lacking pores or pits, narrow distally and laterally, extended proximally to about one-quarter of the frontal surface, rarely two-thirds. Opesia extensive, elliptical, surrounded by an inconspicuous, crenulated rim of cryptocyst. Four spine bases distolaterally and two median spine bases proximally to the opesia on the gymnocyist indenting the cryptocystal rim, 25–30 µm in diame-

ter (Figure 7.2, see arrows). Avicularia and kenozooids absent. Ancestrula not seen.

Measurements (µm). ZL 536±60, 440–622 (1, 15); ZW 309±26, 273–342 (1, 15); OL 341±40, 286–363 (1, 10); OW 271±14, 247–295 (1, 10).

Remarks. This species resembles the recent *Osburnea biscuta* (Osburn, 1950), from the Pacific coast of Panama, in having a gymnocyist commonly extended proximally to one-quarter of the frontal surface, four distolateral spines and more than a single proximomedial spine: the number of proximomedial spines in the Pliocene species is constantly two, while it varies from one to five in the recent species. *Osburnea* aff. *biscuta* also differs from the nominal species in having a less conspicuous cryptocyst surrounding the extensive opesia and, based on the size range reported in Osburn (1950), much larger zooids (0.45–0.62 mm long by 0.27–0.34 mm wide vs 0.26–0.30 mm long by 0.18–0.22 mm wide in the fossil and recent species, respectively). The *Osburnea biscuta* species group is presently restricted to the Pacific coast of

Panama. If the affinity of the Tamiami Formation species is confirmed, it may indicate a change in the biogeographical distribution driven by the closure of the Panamanian gateway. This is the second fossil record of *Osburnea*, the oldest being *O. aquitana* (Nikulina and Taylor, 2009) reported from the Miocene of France.

Superfamily Calloporoidea Norman, 1903

Family Calloporidae Norman, 1903

Genus *Amphiblestrum* Gray, 1848

Amphiblestrum constrictum Ulrich and Bassler,

1904

Figure 8

- 1904 *Amphiblestrum constrictum* Ulrich and Bassler, p. 413, pl. 115, figures 6, 7, pl. 118, figure 15.
- 1923 *Amphiblestrum constrictum* Canu and Bassler, p. 44, pl. 13, figures 1–6.
- 1923 *Amphiblestrum tenuiparietis* Canu and Bassler, p. 45, pl. 13, figure 7.

Figured material. UF 305762 (Shell 5); UF 305763 (Shell 329); UF 305764 (Shell 28). Pliocene, lower Tamiami Formation.

Description. Colony encrusting, multiserial, unilaminar, sheet-like (Figure 8.1). Pore-chamber windows visible at colony growing edge (Figure 8.1, 3), a slightly larger one placed distally and two per side placed distolaterally, elliptical, 95–125 μm long by 35–40 μm wide. Autozooids distinct with deep interzooidal furrows, arranged quincuncially, rounded hexagonal, slightly longer than wide (mean L/W 1.28). Gymnocyst inconspicuous, visible proximally and laterally on some zooids; cryptocyst elongate oval to pyriform, defined by a slightly raised, crenulated rim, extended usually one-quarter to one-third of zooidal frontal length, rarely half, granular, with granules aligned in rows radiating from the proximal margin of the opesia, reduced laterally and very narrow distally, sloping inwards. Opesia pear-shaped, constricted distolaterally by two stout, rounded, condyle-like projections of the cryptocyst. One or two distolateral oral spine bases present in some autozooids (Figure 8.4, see arrows). Adventitious avicularia columnar, placed on the proximolateral corner of most autozooids, pear-shaped (Figure 8.3, 5); rostrum rounded, proximolaterally or distolaterally directed; two stout condyles. Interzooidal avicularia absent. Ovicell hyperstomial, globular; endooecium calcified, granular, crescentic; ectooecium reduced to a marginal band of smooth calcification bordering the endooecium (Figure 8.6). Ancestrula tatiform with an uncertain number of circumopesia spines, ovoidal, about 380 μm long by 320 μm wide, surrounded by

seven autozooids (Figure 8.2, 7); gymnocyst more developed proximally, tapering laterally, disappearing distally, but completely overgrown in large colonies by later autozooids; cryptocyst very narrow, crenulated; opesia subcircular, about 200 μm in diameter. A distal (300–340 μm long by 150–250 μm wide) and two distolateral zooids (310–410 μm long by 230–260 μm wide) budded directly from the ancestrula, similar to later autozooids but smaller. Intramural buds and closure plates observed (Figure 8.4).

Measurements (μm). ZL 465 \pm 34, 406–523 (2, 20); ZW 363 \pm 38, 294–437 (2, 20); OL 251 \pm 26, 206–289 (2, 20); OW 223 \pm 18, 190–259 (2, 20); AvL 133 \pm 12, 112–149 (2, 12); AvW 100 \pm 9, 89–119 (2, 12); OvL 167 \pm 10, 149–186 (2, 15); OvW 233 \pm 15, 207–260 (2, 15).

Remarks. *Amphiblestrum constrictum* Ulrich and Bassler, 1904 was originally described from the Miocene St. Mary's Formation of Maryland and subsequently reported from the Pliocene of Virginia and the Pleistocene of South Carolina by Canu and Bassler (1923). Canu and Bassler (1923) also described a new species of *Amphiblestrum* from the Pliocene of Florida, *A. tenuiparietis*, distinguished from *A. constrictum* in having circumopesia spines in the ancestrula and distal opesia spines in the autozooids. However, SEM images of the holotype of *A. constrictum* (Figure 9.1–3; USNM 68459) revealed the presence of 1–4 distal opesia spine bases in some autozooids. Unfortunately, the ancestrula was not observed in the holotype of *A. constrictum*, but our Pliocene Florida specimens show that spines in the ancestrula are often masked owing to recrystallization or covered by the subsequently budded periancestrular autozooids. No other significant differences can be observed between these two species, and *A. tenuiparietis* (Figure 9.4–6; USNM 68460) is here considered to be a junior subjective synonym of *A. constrictum*. As Canu and Bassler (1923) already stated, intracolony variability for species in this genus is high. Some variations we observed in both our specimens and the holotype of *A. constrictum* include: the direction of avicularia, which are mostly directed proximally and only exceptionally distally; the extension of the ectooecial calcification; and the presence/absence of spine bases and their number.

Genus *Aplousina* Canu and Bassler, 1927

Aplousina grandis (Canu and Bassler, 1923)

Figure 10

- 1923 *Membrendoecium grande* Canu and Bassler, p. 36, pl. 11, figures 10–12.

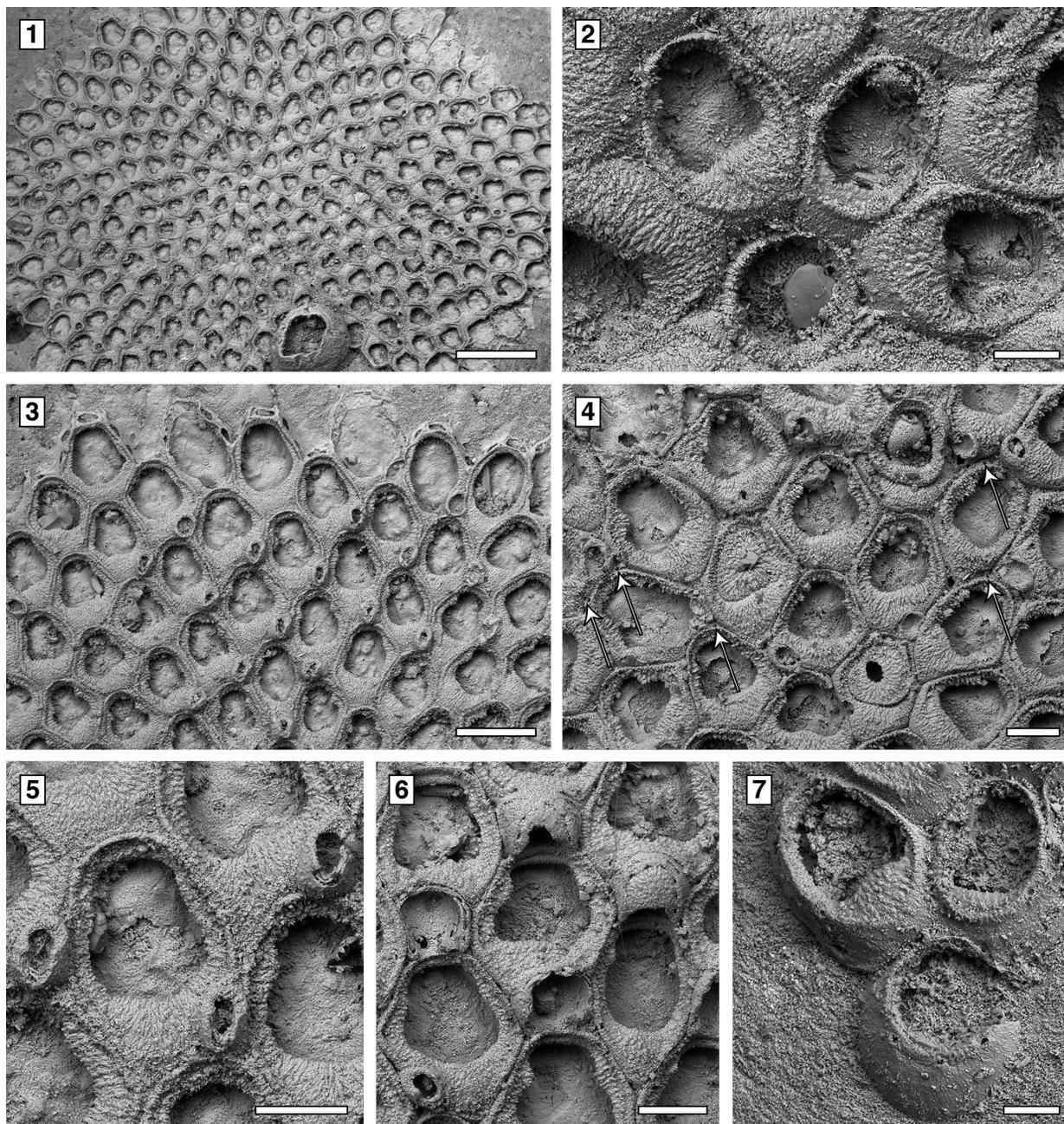


FIGURE 8. *Amphiblestrum constrictum* Ulrich and Bassler, 1904, Pliocene, lower Tamiami Formation Units 10/11. **1–3**, UF 305762 (Shell 5). **1**, general view of an extensive, circular, sheet-like colony overgrown by a barnacle (1 mm). **2**, close-up of a partially overgrown tatiform ancestrula and first budded autozooids (100 μ m). **3**, group of autozooids including colony growing edge, showing pore-chamber windows and adventitious avicularia (500 μ m). **4–6**, UF 305763 (Shell 329). **4**, group of autozooids showing intramural buds and closure plates (200 μ m), with arrows indicating distolateral oral spine bases. **5**, close-up of adventitious avicularia (200 μ m). **6**, group of ovicellate zooids (200 μ m). **7**, UF 305764 (Shell 28), close-up of the tatiform ancestrula and first budded zooids (100 μ m).

Figured material. UF 305765 (Shell 13); UF 305766 (Shell 504). Pliocene, lower Tamiami Formation.

Description. Colony encrusting, multiserial, unilaminar, sheet-like (Figure 10.1). Pore-chambers

lacking. Autozooids distinct with shallow interzooidal furrows, arranged quincuncially, highly variable in shape, from ovoidal to hexagonal, lozenge-shaped or pear-shaped, very large, averaging 1.5 times longer than wide. Autozooids located at col-

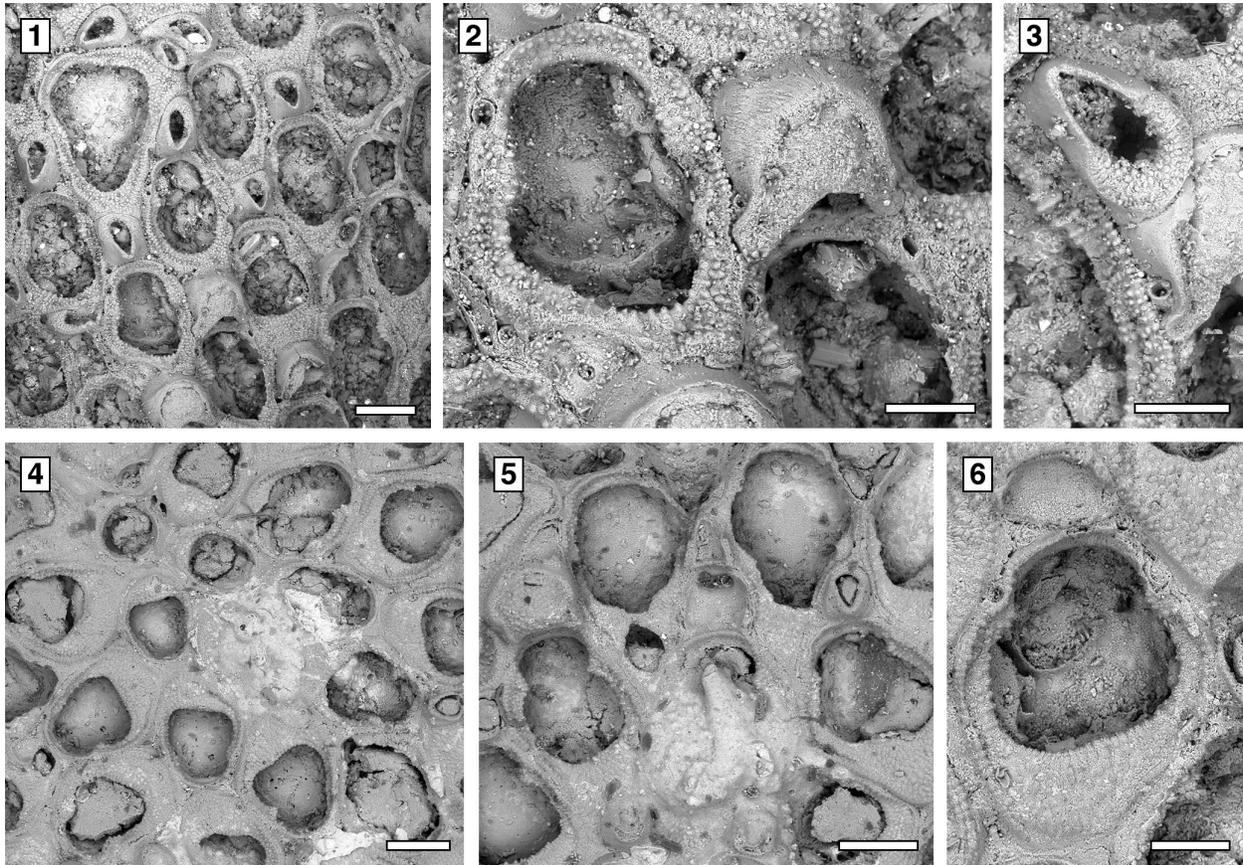


FIGURE 9. 1–3, *Amphiblestrum constrictum* Ulrich and Bassler, 1904, holotype USNM 68459, Miocene, Maryland, USA. 1, part of the colony (200 μ m). 2, close-up of an autozooid with four distolateral spine bases and an ovicell (100 μ m). 3, close-up of the adventitious avicularium (100 μ m). 4–6, *Amphiblestrum tenuiparietis* Canu and Bassler, 1923, holotype USNM 68460, Pliocene, Florida, USA. 4, part of the colony (200 μ m). 5, close-up including autozooids, an ovicellate zooid and an adventitious avicularium (200 μ m). 6, close-up of a zooid with incomplete ovicell and distolateral spine bases (100 μ m).

ony bifurcation either paired and narrow with the two zooids as large as the preceding autozooid in the row, or irregularly shaped and much larger than the preceding autozooid in the row (Figure 10.2–3). Frontal surface almost entirely occupied by the opesia, which assumes the shape of the autozooid. A very narrow rim of smooth gymnocyst occasionally visible proximally and distally; a narrow, low-beaded rim of cryptocyst encircling the opesia. Ovicell endozooidal, oecium hood-like, granular (Figure 10.3). Avicularia and spines absent. Intramural buds common (Figure 10.2). Irregularly shaped kenozooids formed along the edges between contacting colonies (Figure 10.4). Ancestrula pear-shaped, similar to later autozooids in size and in the development of the gymnocyst and the cryptocyst, budding one distal and two distolateral zooids (Figure 10.5–6). First budded zooids smaller than the ancestrula and later autozooids.

Measurements (μ m). ZL 786 \pm 44, 690–857 (1, 15); ZW 519 \pm 49, 399–602 (1, 15); OL 686 \pm 52, 577–775 (1, 15); OW 446 \pm 33, 383–509 (1, 15); OvL 153 \pm 12, 128–168 (1, 15); OvW 340 \pm 23, 294–373 (1, 15).

Remarks. *Aplousina grandis* was reported from the Pliocene Duplin Formation of North Carolina and the lower Pleistocene Waccamaw Formation of North Carolina (Canu and Bassler, 1923). The species was first attributed to the genus *Membrendoecium* Canu and Bassler, 1917, which was later placed in synonymy with *Antropora* Norman, 1903 by Silén (1941). In introducing the genus *Aplousina* in 1927, the authors referred this species to the new genus because of the lack of avicularia compared to other species included in *Membrendoecium* (Canu and Bassler, 1927, p. 3). A very similar species, *Aplousina gigantea* Canu and Bassler, 1928a, is at the present day very abundant in the eastern Gulf of Mexico and the Straits of Florida

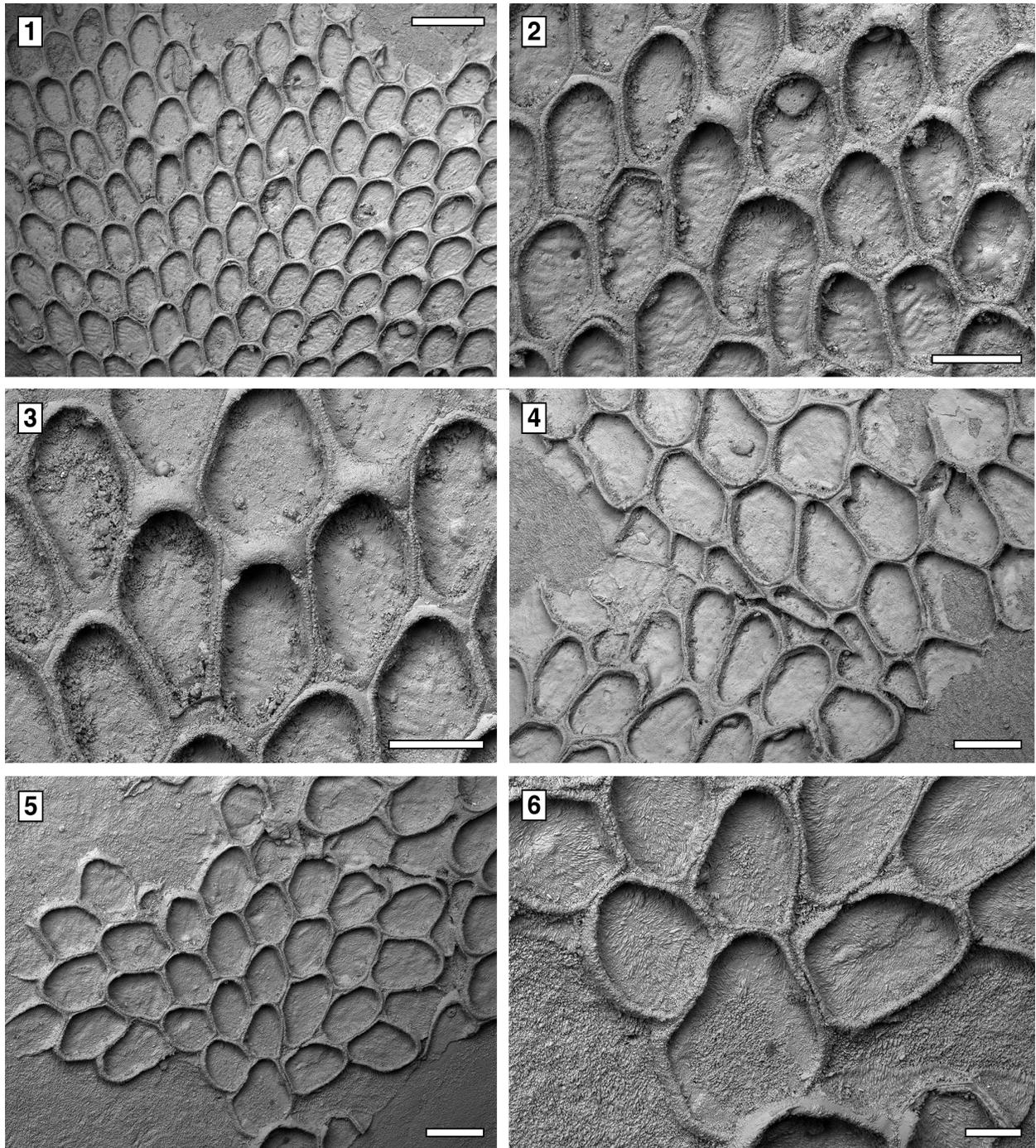


FIGURE 10. *Aplousina grandis* (Canu and Bassler, 1923), Pliocene, lower Tamiami Formation Units 10/11. **1–3**, UF 305765 (Shell 13). **1**, general view of an extensive, sheet-like colony (1 mm). **2**, group of ovicellate and non-ovicellate zooids, some with intramural buds (500 μ m). **3**, close-up of three ovicellate zooids (400 μ m). **4–6**, UF 305766 (Shell 504). **4**, irregularly shaped kenozooids formed along the encounter edge of two colonies (500 μ m). **5**, small colony with ancestrula (500 μ m). **6**, close-up of the ancestrula and first budded autozooids (200 μ m).

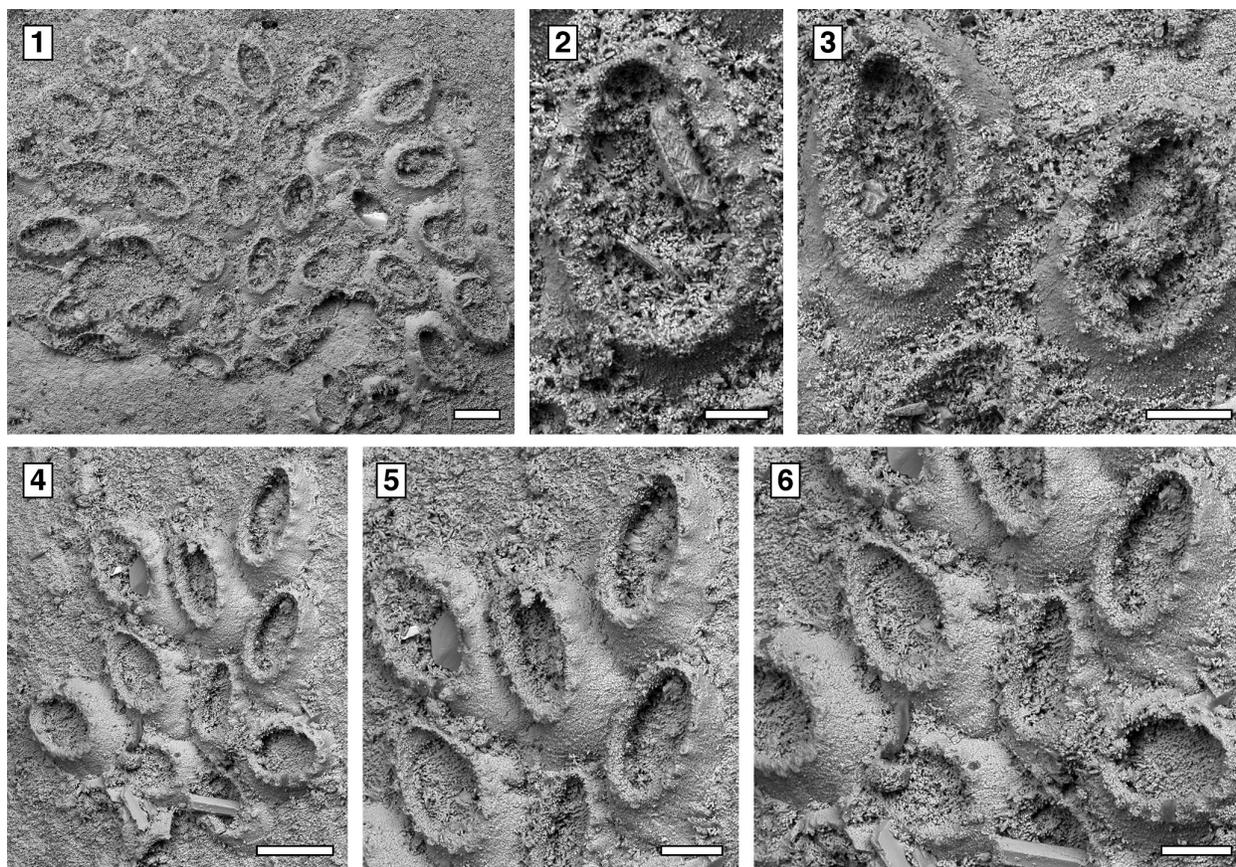


FIGURE 11. *Cauloramphus?* sp., UF 305767a, b (Shell 324), Pliocene, lower Tamiami Formation Units 10/11. **1**, general view of the small colony (a) (200 μ m). **2**, close-up of an autozooid showing the numerous circumopesia spine bases (60 μ m). **3**, close-up of two autozooids with a large gap in between likely to have been occupied by the connecting tubular chambers typical of some species of *Cauloramphus* (100 μ m). **4**, tilted view of small colony (b) (200 μ m). **5**, group of tilted autozooids, the top right autozooid showing cut-off tubular chambers on the lateral walls (100 μ m). **6**, close-up of some autozooids (100 μ m).

(Winston, 2016). Canu and Bassler (1928a) distinguished *A. grandis* from *A. gigantea* based on zooid size, with zooids larger in the living species than in the fossil. However, the zooid size range in fossil specimens overlaps with that of recent material. Additional differences are the shape of the zooids, highly variable in *A. grandis*, but consistently lozenge-shaped in *A. gigantea*, and the greater thickness of the walls in the fossil species compared to the modern.

Genus *Cauloramphus* Norman, 1903

Cauloramphus? sp.

Figure 11

Figured material. UF 305767a, b (two small colonies) (Shell 324). Pliocene, lower Tamiami Formation.

Description. Colony encrusting, multiserial, unilaminar (Figure 11.1, 4). Autozooids distinct, delineated by conspicuous interzooidal furrows,

irregularly arranged, oval, longer than wide (mean L/W 1.59). Gymnocyte smooth, convex, broad and sloping proximally and laterally; cryptocyst a nearly vertical shelf, granular, narrow. Opesia extensive, occupying most of the zooidal frontal surface, oval, surrounded by an uncertain number of oral spine bases (seemingly about 17–18), 14–20 μ m in diameter (Figure 11.2–3, 11.5–6). Avicularia, ovi-cells and ancestrula not observed.

Measurements (μ m). ZL 370 \pm 35, 324–450 (2, 15); ZW 233 \pm 32, 176–278 (2, 15); OL 254 \pm 28, 205–309 (2, 15); OW 149 \pm 21, 117–193 (2, 15).

Remarks. The only two specimens of *Cauloramphus?* sp. (Figure 11.1, 11.4) found, encrust the same *Anomia* shell. The colonies are small, with 25 and 11 zooids preserved, respectively. Unfortunately, the poor preservation prevents clear observation of the connections among autozooids, but it is apparent from some zooids that they are connected via narrow tubular chambers spaced out

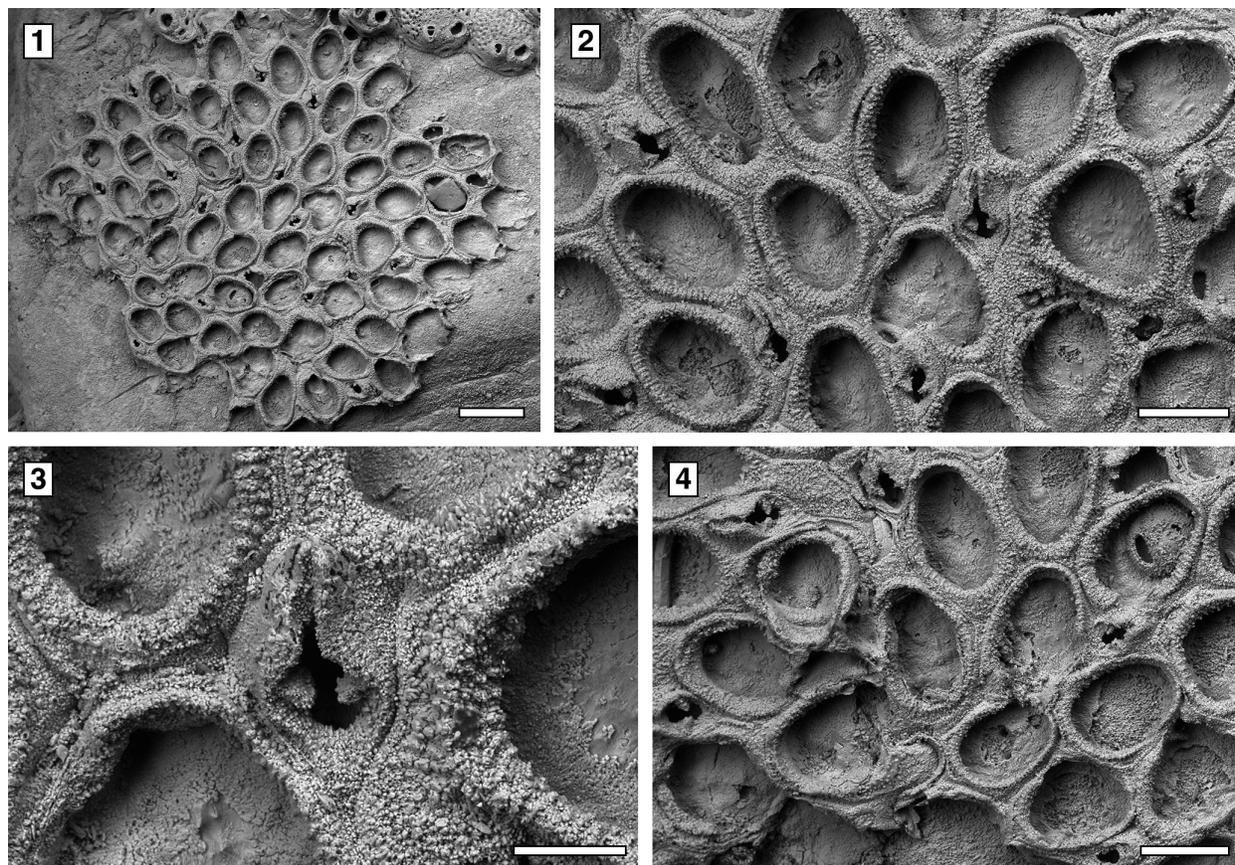


FIGURE 12. *Copidozoum cf. parvirostris* (Canu and Bassler, 1923) comb. nov., UF 305768 (Shell 12), Pliocene, lower Tamiami Formation Units 10/11. **1**, general view of the colony (400 µm). **2**, group of autozooids and interzooidal avicularia (200 µm). **3**, close-up of an interzooidal avicularium (100 µm). **4**, intramural buds (200 µm).

with lacunae (Figure 11.5). The recent *Cauloramphus opertus* Canu and Bassler, 1928a from the Gulf of Mexico is similar in size to the fossil species. It is characterized by having 20 opesial spines forming a costate frontal shield plus four erect orificial spines and long, pedunculate, horn-shaped avicularia. Because of both preservation and small size of the colonies available in the Tamiami Formation material, we are unable to observe the original spine arrangement and we cannot rule out the former presence of pedunculate avicularia in this species. An alternative attribution would be to *Retevirgula* Brown, 1948, but this genus has ovicells, which have not been observed in the scant material available.

Genus *Copidozoum* Harmer, 1926
Copidozoum cf. parvirostris (Canu and Bassler, 1923) comb. nov.

Figure 12

cf. 1923 *Callopora parvirostris* Canu and Bassler, p. 41, pl. 12, figure 3.

Figured material. UF 305768 (Shell 12). Pliocene, lower Tamiami Formation.

Description. Colony encrusting, multiserial, unilaminar (Figure 12.1). Pore-chamber windows present (Figure 12.2). Autozooids distinct with deep interzooidal furrows, quincuncially arranged, oval to rounded polygonal, longer than wide (mean L/W 1.37). Gymnocyst extremely reduced and discontinuously visible only along the margins of some zooids; cryptocyst narrow, forming a raised mural rim encircling the opesia, coarsely granular with a striated appearance owing to the granules aligned in radial rows, then sloping steeply inwards. Opesia extensive, occupying most of the zooidal frontal surface, variable in shape from oval to elliptical or pear-shaped. Spine bases absent. Avicularia interzooidal, frequent, scattered throughout the colony, placed on a rhomboidal cystid (Figure 12.3); gymnocyst more developed laterally; rostrum raised, acicular, directed distally; two robust condyles as pivots. Ovicells not observed. Intramural buds

present in both zooids and avicularia (Figure 12.3–4). Ancestrula not seen.

Measurements (µm). ZL 373±35, 337–427 (1, 10); ZW 271±19, 245–313 (1, 10); OL 284±21, 255–315 (1, 10); OW 185±17, 158–217 (1, 10); AvL 218±17, 190–242 (1, 9); AvW 151±18, 129–177 (1, 9).

Remarks. As already noted by Echols (1960), *Callopora parvirostris* Canu and Bassler, 1923, from the Miocene of Maryland is very similar to *C. tenuirostre*, the main difference being the greater size of the zooids. Canu and Bassler (1923) also distinguished *Callopora parvirostris* from *Copidozoum tenuirostre* based on the size of the interzooidal avicularia, which they stated is much smaller in the former species compared to the latter species. Unfortunately, Canu and Bassler (1923) failed to provide a size range for the avicularia in *C. parvirostris*. However, based on the image provided (Canu and Bassler, 1923, pl. 12, figure 3), the length of an avicularium seems to be about half the length of a zooid (ZL 0.55–0.60 mm), thus the avicularia in the holotype of *C. parvirostris* are about the same size of the avicularia in *C. cf. parvirostris* from the Tamiami Formation. The Tamiami Formation specimen differs from the holotype in having smaller autozooids (ZL 0.34–0.43 mm versus 0.55–0.60 mm).

Superfamily Microporoidea Gray, 1848

Family Onychocellidae Jullien, 1882

Genus *Floridina* Jullien, 1882

Floridina regularis Canu and Bassler, 1923

Figure 13

1923 *Floridina regularis* Canu and Bassler, p. 57, pl. 14, figures 7, 8.

2008 *Floridina regularis* Knowles, p. 87, figures 3.1, 4.1–4.5.

2018 *Floridina regularis* Taylor et al., figure 11c, d.

Figured material. UF 305769 (Shell 1); UF 305770 (Shell 18); UF 305771 (Shell 22); UF 305772 (Shell 24); UF 305773 (Shell 324). Pliocene, lower Tamiami Formation.

Description. Colony encrusting, multiserial, uni- to multilaminar (Figure 13.1, 3). Autozooids distinct with shallow interzooidal furrows, quincuncially arranged, subhexagonal with rounded distal margin and straight to slightly concave, sometimes undulose, proximal margin, longer than wide (mean L/W 1.23). Gymnocyst minimal, sometimes visible along the zooidal proximal margin. Cryptocyst shelf occupying about half of zooidal frontal length, coarsely granular, depressed centrally, and raised at the edges, which appear striated because of finer, densely aligned granules. Opesia bell-

shaped with straight or slightly concave proximal margin, rarely trifoliate with lateral opesiules or a convex proximal margin. Oral spines absent. A very narrow, granular cryptocyst shelf sometimes visible projecting proximally from the distal margin of the opesia. Vicarious avicularia infrequent, longer than autozooids, lozenge-shaped or sub-pentagonal (Figure 13.2); rostrum slightly raised, rounded triangular, laterally showing a narrow band of smooth gymnocyst; opesia placed centrally, ovoidal to elliptical; cryptocyst coarsely granular and depressed centrally as in autozooids; granules surrounding the opesia give a denticulate appearance to the outline. Ovicellate zooids slightly larger than autozooids; opesia generally larger than in autozooids and more elongate (mean L/W 1.54 vs 1.41 in autozooids) and with the distal edge formed by the cryptocyst of the next distal zooid (Figure 13.3). Irregularly shaped kenozooids, variable in size, observed along the edges of two abutting colonies (Figure 13.4); cryptocyst as in autozooids, variably developed; opesia located centrally, variable in size and shape. Ancestrula similar to later autozooids but smaller, 260–265 µm long by 210–215 µm wide; opesia bell-shaped, 85–115 µm long by 115–130 µm wide proximal to constrictions, 90–115 µm wide distal to constrictions (Figure 13.5); surrounded by six autozooids. One distal and two distolateral zooids budded directly from the ancestrula, 230–300 µm long by 195–240 µm wide and bell-shaped opesia 105–115 µm long by 110–120 µm proximally and 85–95 µm wide distally. Reparative structures often observed in autozooids (Figure 13.6). Closure plates with scars of the operculum observed in some colonies (Figure 13.7).

Measurements (µm). ZL 385±24, 341–429 (2, 20); ZW 313±20, 275–337 (2, 20); OL 155±6, 141–164 (2, 20); OWmin 110±9, 93–120 (2, 20); OWmax 147±11, 125–165 (2, 20); AvL 396±42, 310–476 (4, 20); AvW 213±41, 169–343 (4, 20); AvOpL 150±28, 96–194 (4, 20); AvOpW 75±22, 41–119 (4, 20); ZL* 414±62, 331–552 (3, 10); ZW* 329±60, 253–430 (3, 10); OL* 222±17, 193–243 (3, 10); OW*min 144±4, 138–151 (3, 10); OW*max 183±10, 169–206 (3, 10). *Maternal zooids.

Remarks. *Floridina regularis* was first described from the Pliocene Duplin Formation of North Carolina (Canu and Bassler, 1923) and later found in other North American Pliocene and Pleistocene deposits, including the upper Tamiami Formation of Florida, varying in abundance (Knowles, 2008). On the *Anomia* shells, *F. regularis* is extremely abundant (see Table 1). *Floridina regularis* differs from

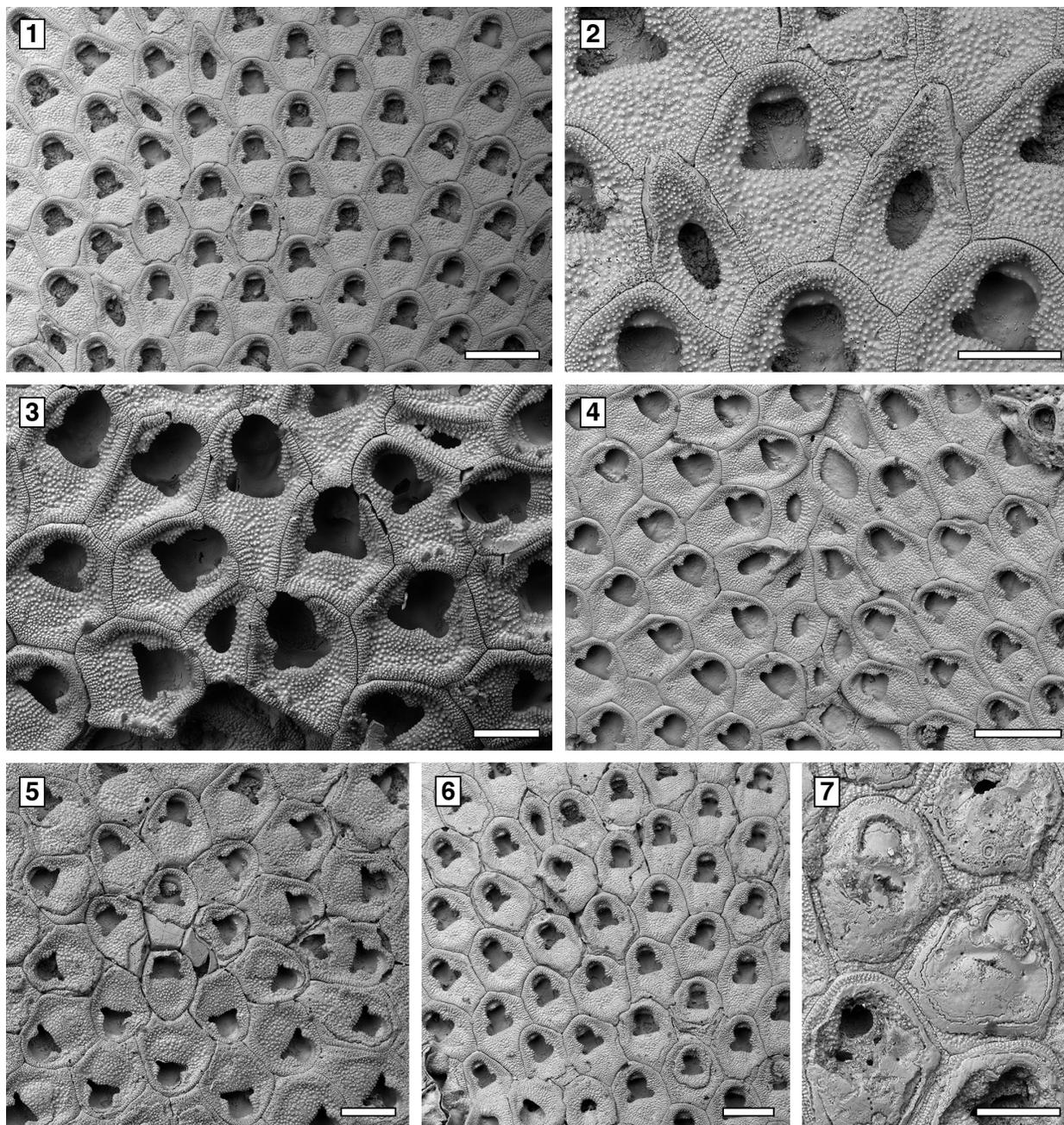


FIGURE 13. *Floridina regularis* Canu and Bassler, 1923, Pliocene, lower Tamiami Formation Units 10/11. **1–2**, UF 305769 (Shell 1). **1**, group of autozooids and vicarious avicularia (400 μ m). **2**, close-up of an autozooid flanked by two vicarious avicularia (200 μ m). **3**, UF 305770 (Shell 18), group of fertile zooids (200 μ m). **4**, UF 305771 (Shell 22), irregularly shaped kenozooids formed at the encounter edge between two abutting colonies (400 μ m). **5**, UF 305772 (Shell 24), ancestrula and early astogeny (300 μ m). **6**, UF 305770 (Shell 18), intramural buds (200 μ m). **7**, UF 305773 (Shell 324), closure plates with scars of the operculum (200 μ m).

F. antiqua (Smitt, 1873), the type species of the genus, in having more-angular zooids and a less-pronounced curvature of the proximal margin of the opesia, which often appears more bell-shaped than trifoliate. It also differs from *F. subantiqua* Di Martino, Taylor, and Portell, 2017 from the Chipola For-

mation of Florida in having smaller autozooids and vicarious avicularia lacking lateral constrictions.

Family Microporidae Gray, 1848

Genus *Micropora* Gray, 1848

Micropora stellata sp. nov.

Figure 14

TABLE 1. List of bryozoan species with abundances (number of colonies) found in Units 10/11 of the upper Pliocene lower Tamiami Formation of Florida, USA.

	Species	Authors	No. colonies
1	<i>Acanthodesia</i> cf. <i>denticulata</i>	(Busk, 1856)	1
2	<i>Acanthodesia</i> sp.	—	11
3	<i>Amphiblestrum constrictum</i>	Ulrich and Bassler, 1904	109
4	<i>Aplousina grandis</i>	(Canu and Bassler, 1923)	165
5	<i>Cauloramphus?</i> sp.	—	3
6	<i>Celleporaria</i> sp.	Canu and Bassler, 1923	24
7	<i>Copidozoum</i> cf. <i>parvirostris</i>	(Canu and Bassler, 1923)	8
8	<i>Cyclocolpota perforata</i>	Canu and Bassler, 1923	247
9	<i>Cycloperiella rubra</i>	Canu and Bassler, 1923	94
10	<i>Floridina regularis</i>	Canu and Bassler, 1923	554
11	<i>Hippaliosina rostrigera</i>	(Smitt, 1873)	7
12	<i>Hippopleurifera mucronata</i>	(Smitt, 1873)	51
13	<i>Metropieriella reversa</i>	(Ulrich and Bassler, 1904)	190
14	<i>Micropora stellata</i>	sp. nov.	9
15	<i>Microporella</i> cf. <i>protea</i>	Winston, 2005	1364
16	<i>Microporella sarasotaensis</i>	sp. nov.	592
17	<i>Microporella tamiamiensis</i>	sp. nov.	446
18	<i>Oncousoecia?</i> sp.	—	9
19	<i>Osburnea</i> aff. <i>biscuta</i>	(Osburn, 1950)	3
20	<i>Plesiocleidochasma</i> cf. <i>vestitum</i>	(Canu and Bassler, 1923)	106
21	<i>Pourtalesella chiarae</i>	sp. nov.	510
22	<i>Puellina scripta?</i>	(Reuss, 1848)	917
23	<i>Reptadeonella umbilicata</i>	(Lonsdale, 1845)	109
24	<i>Smittoidea maleposita</i>	(Canu and Bassler, 1923)	25
25	<i>Spiniflabellum laurae</i>	sp. nov.	2
26	<i>Stephanollina vorax</i>	(Canu and Bassler, 1923)	78
27	<i>Stylopoma leverhulme</i>	Di Martino, Taylor and Portell, 2017	13
28	<i>Trypostega composita</i>	sp. nov.	97
29	<i>Watersipora?</i> sp.	—	9

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Type material. Holotype UF 305774 (Shell 33); paratype UF 305775 (Shell 31). Pliocene, lower Tamiami Formation.

Etymology. From the Latin ‘stellatus, -a, -um’ meaning ‘starry, with stars’, referring to the finely denticulate, star-shaped pseudopores in the frontal shield.

Diagnosis. Colony encrusting; autozooids rhomboidal to hexagonal. Pore-chamber windows present. Frontal shield granulated and irregularly pseudoporous; pseudopores star-shaped because of a radiating denticulation. Opesiules paired, symmetrical, bordered by a raised margin, proximolateral to orifice. Orifice transversely D-shaped with a

prominent knob on each side. Oral spines absent. Avicularia interzooidal, distal to the orifice in non-ovicellate zooids, proximolaterally directed, cross-bar complete. Ovicells with median, chevron-shaped suture, smooth and a tubercle.

Description. Colony encrusting, multiserial, unilaminar. Pore-chamber windows visible at colony growing edge (Figure 14.1) or along the perimeter of broken autozooids, numerous, closely spaced in a continuous row all along the zooidal margin, elliptical, 30–45 µm long by 20–25 µm wide. Autozooids distinct, outlined by a thin interzooidal incision and a slightly raised, thickened (about 30 µm), granulated margin most pronounced where it borders the opesiules (Figure 14.2), quincuncially

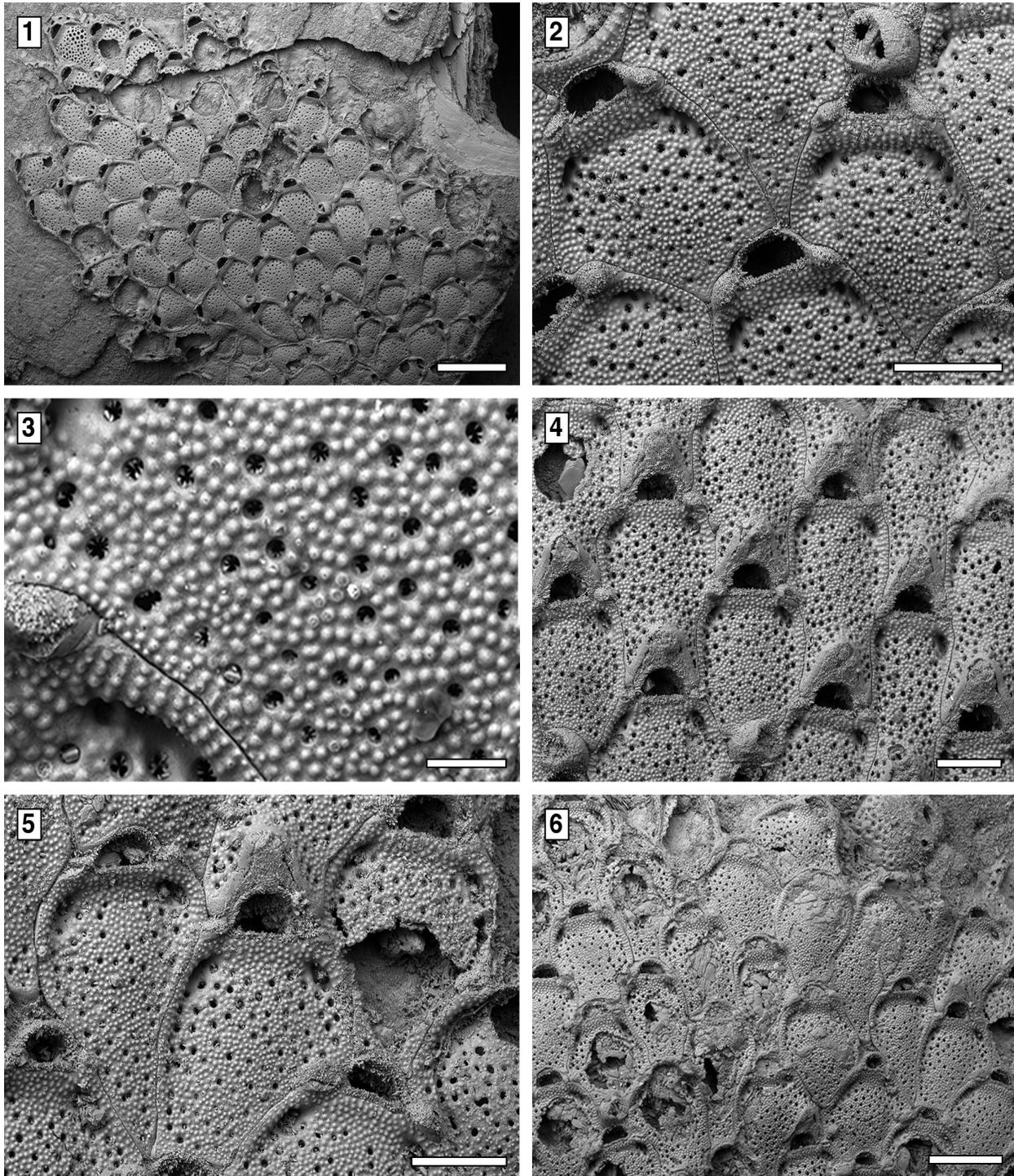


FIGURE 14. *Micropora stellata* sp. nov., Pliocene, lower Tamiami Formation Units 10/11. **1–3**, paratype UF 305775 (Shell 31). **1**, general view of the colony (600 µm). **2**, close-up of two autozooids with prominent knobs on each side of the orifice and a distal, interzooidal avicularium (200 µm). **3**, close-up of pseudopores with radiating denticulation giving them a star-shaped appearance (50 µm). **4–6**, holotype UF 305774 (Shell 33). **4**, group of ovicellate zooids (200 µm). **5**, close-up of an ovicellate zooid (200 µm). **6**, part of the colony with closure plates (400 µm).

arranged, rounded rhomboidal to irregularly hexagonal, longer than wide (mean L/W 1.32). Frontal shield flat to slightly convex, finely granulated, irregularly pseudoporous and with paired opesiules; pseudopores 30–65 (on average 50) unevenly scattered throughout the frontal, rounded, 8–12 μm in diameter, with radiating denticulation giving them a star-shaped appearance (Figure 14.3); opesiules bean- to spindle-shaped, symmetrical, 50–80 μm long by 20–25 μm wide, placed proximolateral to the orifice. Orifice transversely D-shaped, broader than long, elevated from the frontal surface, which starts to become raised below its proximal margin, forming an imperforate, coarsely granular, sloping shelf about 60–70 μm wide; a smooth, prominent knob, 50–70 μm in diameter, flanking the orifice on each side in most ovicellate and non-ovicellate zooids (Figure 14.2, 4). Oral spines absent; closure plates observed (Figure 14.6). Avicularia interzooidal, usually lying distal to the orifice in non-ovicellate zooids, on a raised, sloping, smooth cystid, almost elliptical in outline (Figure 14.1–2); rostrum triangular, occupying most of the raised top of the cystid, proximolaterally directed, either on the left or right; crossbar complete. Ovicells partly immersed in the frontal shield of the next distal zooid, with a median, chevron-shaped, smooth suture terminating in an apical tubercle (Figure 14.4–5). Intramural buds observed. Ancestrula not seen.

Measurements (μm). ZL 504 \pm 46, 410–573 (2, 20); ZW 380 \pm 38, 315–436 (2, 20); OL 79 \pm 6, 70–91 (2, 18); OW 140 \pm 12, 116–163 (2, 18); AvL 156 \pm 14, 135–187 (2, 15); AvW 109 \pm 8, 97–126 (2, 15); OvL 154 \pm 12, 138–173 (1, 12); OvW 224 \pm 13, 193–242 (1, 12).

Remarks. For their Neogene North American specimens, Canu and Bassler (1923) used the name *Micropora coriacea* (Esper, 1791), a European species that, since its first description, has been recorded worldwide at all latitudes and depths, both living and fossil. The advent of the SEM has helped to disentangle this complex of species, showing consistent differences in skeletal characters of material from different geological ages, regions, and habitats. Winston (2005) distinguished the western Atlantic material previously attributed to *M. coriacea* as the new species *M. acuminata*, which mainly differs from the new Tamiami Formation species in having a pointed tip to the ovicell suture. *Micropora angustiscapulis* Winston, Vieira, and Woollacott, 2014 from southwest Brazil is similar in having star-shaped pseudopores and robust knobs flanking the orifice but differs in

having smaller autozooids (342–450 μm long by 234–360 μm wide vs 410–573 μm long by 315–436 μm wide) and smaller orifices (45–54 μm long by 90–108 μm wide vs 70–91 μm long by 116–163 μm wide); unfortunately, the holotype and only specimen available of *M. angustiscapulis* lacks avicularia and ovicells, essential features for a more thorough comparison. Star-shaped pseudopores were also observed in *Micropora robusta* Cook, 1985 (NHMUK 1972.3.3.1; Figure 15.1–2) from west Africa, but this species differs in having a larger orifice, finer granules on the frontal shield, and interzooidal avicularia that are directed distolaterally. Large, robust knobs are present in *Micropora nodimagna* Ramalho and Calliari, 2015 from southern Brazil, which differs from the Tamiami Formation species in the shape of the ovicell, which is squat, more prominent and has a broader, triangular suture of smooth calcification. The fossil specimen of *Micropora coriacea* sensu Canu and Bassler (1923) from the Pleistocene of California (USNM 68480; Figure 15.3–4) differs from the new species in having interzooidal avicularia directed distolaterally and rounded pseudopores lacking denticulation, although the latter character may be dependent on the quality of preservation.

Superfamily Cribrilinoidea Hincks, 1879

Family Cribrilinidae Hincks, 1879

Genus *Puellina* Jullien, 1886

Puellina scripta (Reuss, 1848)

Figures 16–18

- 1848 *Cellepora scripta* Reuss, p. 82, pl. 9, figure 28.
- 1923 *Puellina radiata* forma *scripta* Canu and Bassler, p. 89, pl. 15, figure 12, pl. 35, figure 1.
- 1987 *Puellina scripta* Bishop and Househam, p. 58, figures 98, 99.

Figured material. UF 305776 (Shell 10); UF 305777 (Shell 343); UF 305778 (Shell 22); UF 305779 (Shell 3). Pliocene, lower Tamiami Formation.

Description. Colony encrusting, multiserial, unilaminar, developing extensive sheets (> 200 zooids) (Figure 16.1). Pore-chamber windows visible at colony growing edge, all along zooidal margins, numerous, closely spaced, rounded to elliptical, 30–45 μm long by 20–35 μm wide (Figure 17.4). Autozooids distinct with deep interzooidal furrows, quincuncially arranged, oval to rounded hexagonal, longer than broad (mean L/W 1.43). Gymnocyte generally negligible but more extensive proximally in some zooids (Figure 17.1–3). Frontal shield flat, formed by 12–18 (within colony com-

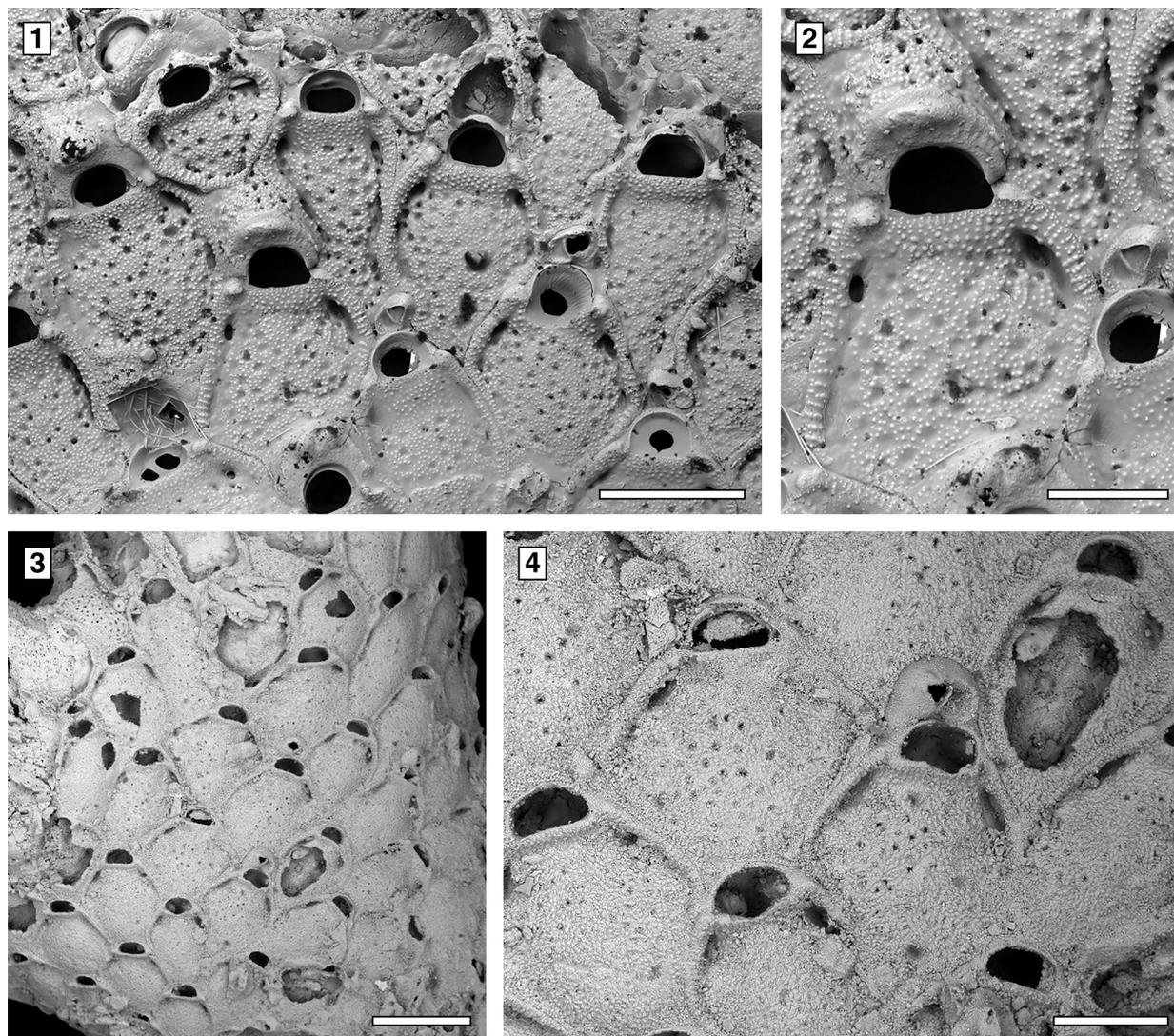


FIGURE 15. 1–2, *Micropora robusta* Cook, 1985, holotype NHMUK 1972.3.3.1, Recent, off Cape Coast, Ghana (27 m depth). 1, group of ovicellate and non-ovicellate zooids, some with closure plates, and interzooidal avicularia (400 μ m). 2, close-up of an ovicellate zooid and an interzooidal avicularium (200 μ m). 3–4, *Micropora coriacea* sensu Canu and Bassler (1923), USNM 68480, Pleistocene, California, USA. 3, group of autozooids and interzooidal avicularia (500 μ m). 4, close-up of some autozooids and an interzooidal avicularium (200 μ m).

only 13, exceptionally 17–18) fused costae, 30–50 μ m at the largest tip, with usually four rounded intercostal lacunae, about 5 μ m in diameter. Distal pair of costae (apertural bar) joined medially, outlining a short triangular area and forming a scarcely developed, median, suboral tubercle in a few zooids (Figure 16.2), otherwise flat, and leaving a central circular lacuna slightly larger than intercostal lacunae, up to 10 μ m in diameter, rarely visible in frontal view. Orifice transversely D-shaped with straight proximal margin, smooth to slightly crenulated in some zooids, constantly bearing five oral spine bases in non-ovicellate zooids and four in ovicellate zooids; spines stout, robust, 20 μ m in

diameter at the base, tapering towards the tips (10 μ m minimum diameter). Avicularia interzooidal, infrequent, placed on an irregularly polygonal cystid of smooth gymnocyst often overgrown by neighbouring autozooids (Figures 16.3, 17.3–6); rostrum ogival, directed distally or distolaterally, lying on the depression between autozooids, smooth; two faint condyles as pivots. Ovicells prominent, helmet-shaped, slightly wider than long, smooth, with a median suture and faint radiating costae, in some zooids more visible than in others (Figures 16.2–3, 17.1–2, 5). Intramural buds present in both autozooids and avicularia (Figure 17.3, 5–6). Ancestrula tafiform, oval, 160–200 μ m long by 100–130 μ m

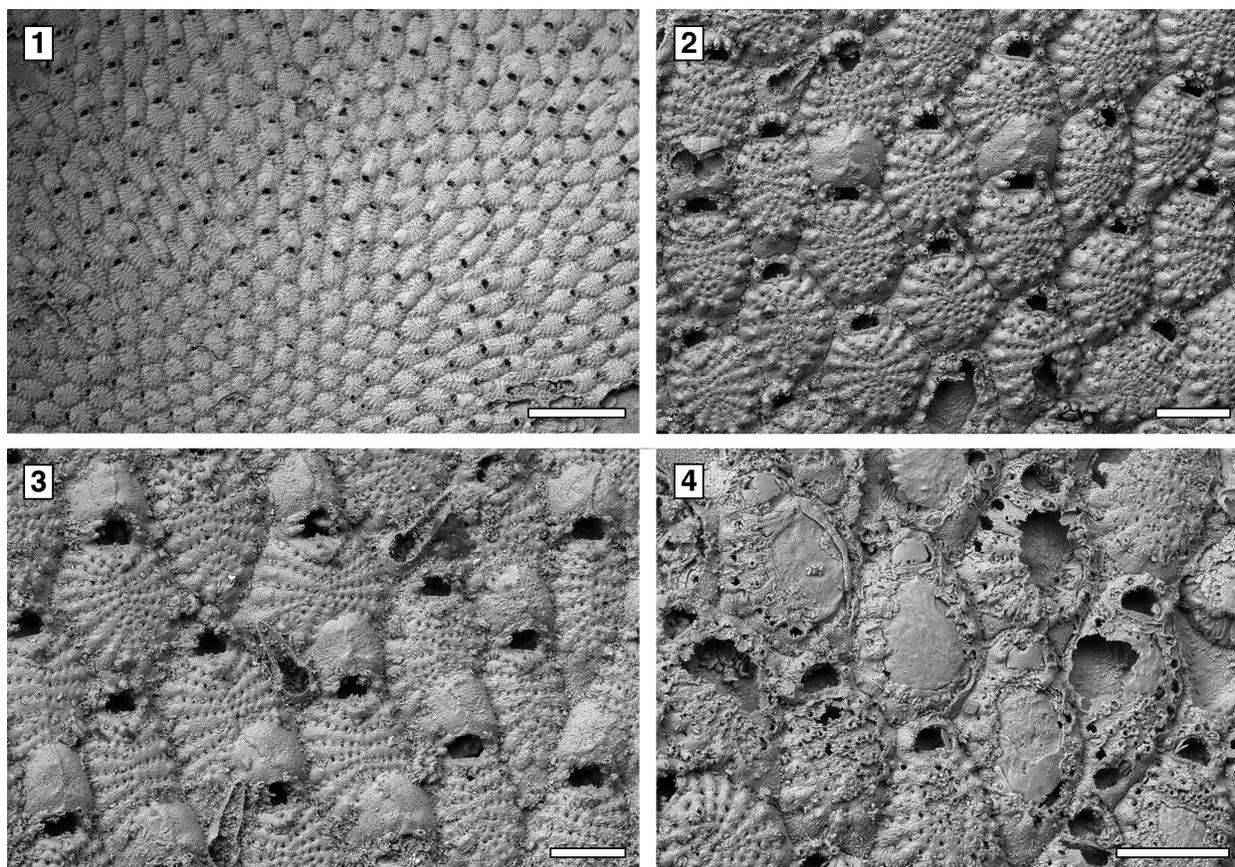


FIGURE 16. *Puellina scripta* (Reuss, 1848), UF 305776 (Shell 10), Pliocene, lower Tamiami Formation Units 10/11. **1**, general view of part of an extensive, sheet-like colony (1 mm). **2–3**, groups of ovicellate and non-ovicellate zooids and interzooidal avicularia (200 μ m). **4**, autozooids with peculiar preservation and closure plates with opercular scars (200 μ m).

wide, surrounded by six autozooids (Figure 18.1–2); gymnocyst more extensive proximally, tapering laterally, negligible distally; opesia elliptical to oval, occupying most of the frontal surface, 100–130 μ m long by 95–110 μ m wide, surrounded by an uncertain number of spines, at least nine. One distal and two distolateral zooids budded directly from the ancestrula, similar to later autozooids but smaller, 190–260 μ m long by 160–195 μ m wide, and with fewer costae forming the frontal shield (up to 10). Costulate kenozooid seen only once as a regeneration of the ancestrula (Figure 18.3–4). Closure plates with opercular scars observed (Figure 16.4). **Measurements (μ m).** ZL 421 \pm 56, 329–542 (3, 25); ZW 295 \pm 39, 214–364 (3, 25); OL 65 \pm 7, 52–75 (3, 20); OW 88 \pm 10, 75–108 (3, 20); AvL 222 \pm 49, 132–303 (3, 12); AvW 118 \pm 14, 96–140 (3, 12); OvL 175 \pm 15, 151–203 (3, 25); OvW 196 \pm 18, 157–232 (3, 25).

Remarks. *Puellina scripta* was first described from the Miocene (Badenian) of Austria and subsequently reported worldwide. However, with the

advent of the SEM, several new species have been distinguished within those records originally attributed to *P. scripta*. Compared to the holotype images depicted by Bishop and Househam (1987, p. 58, figures 98–99), the Tamiami Formation species is similar in the size and shape of zooids and avicularia, in having the suboral mucro poorly developed or totally absent, the frontal shield almost flat with costae separated by four intercostal lacunae, and five robust oral spines. The main difference is the number of costae forming the frontal shield, which is usually 16 in the holotype while is commonly 13 in the Florida specimens, although this still falls in the range of variation of the Florida species. The Tamiami Formation species is also similar to *Puellina radiata* forma *scripta* (Reuss, 1848) sensu Canu and Bassler (1923, p. 89, pl. 35, fig. 1) reported from the Pliocene of Virginia. Among recent western Atlantic species, those characterized by five oral spines in non-ovicellate zooids differ from the Tamiami Formation species in the following characters: *Puellina saginata* Win-

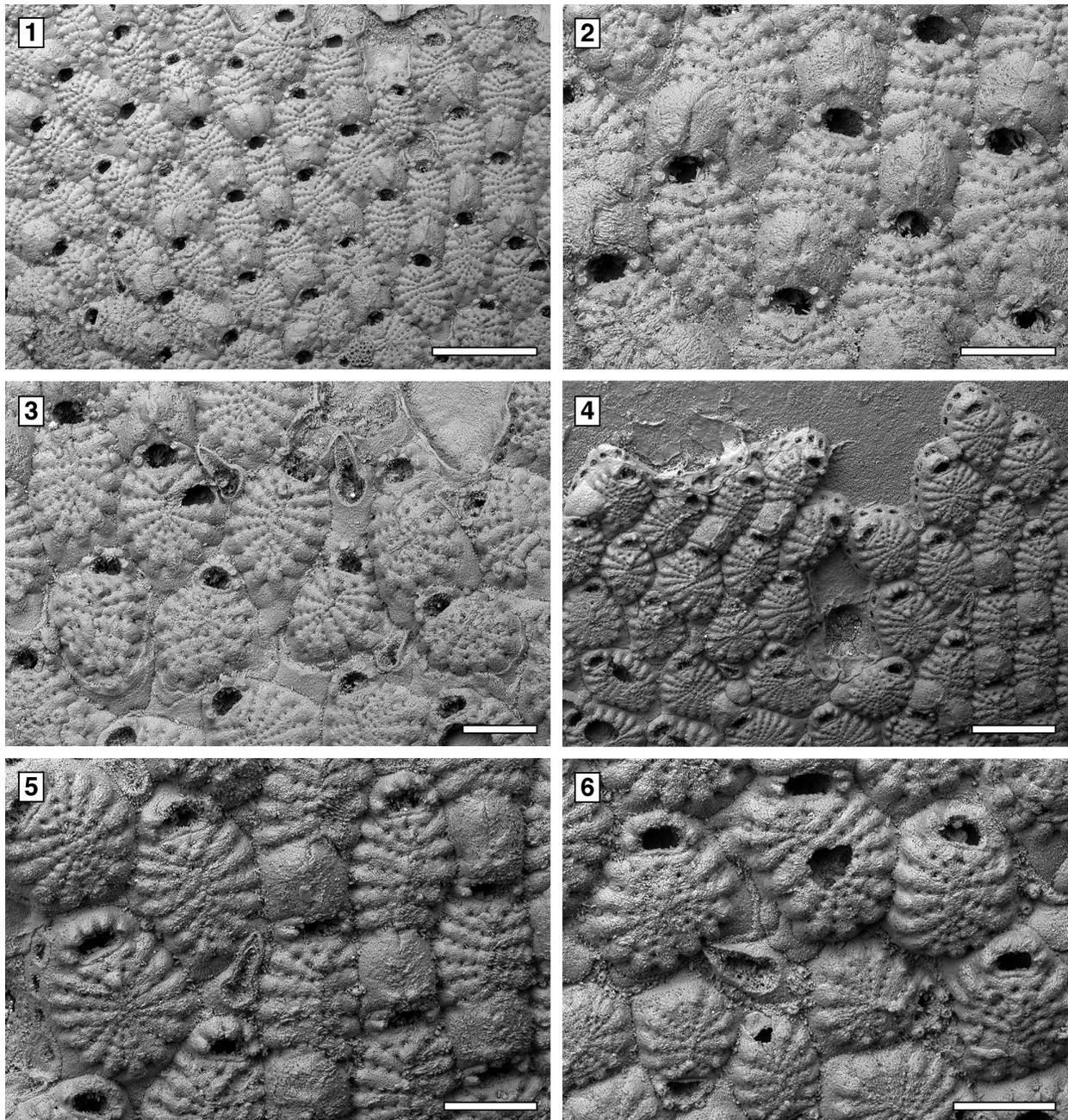


FIGURE 17. *Puellina scripta* (Reuss, 1848), UF 305777 (Shell 343), Pliocene, lower Tamiami Formation Units 10/11. **1**, general view of part of an extensive, sheet-like colony (500 μ m). **2**, groups of ovicellate zooids showing a median suture and faint radiating costae (200 μ m). **3**, autozooids with more extensive, smooth gymnocyst proximally, reparative buds and interzooidal avicularia (200 μ m). **4**, autozooids at colony growth edge showing pore-chamber windows (400 μ m). **5**, close-up of ovicellate and non-ovicellate zooids showing the robust oral spines and interzooidal avicularium with intramural bud (200 μ m). **6**, group of autozooids and interzooidal avicularium intramurally budded (200 μ m).

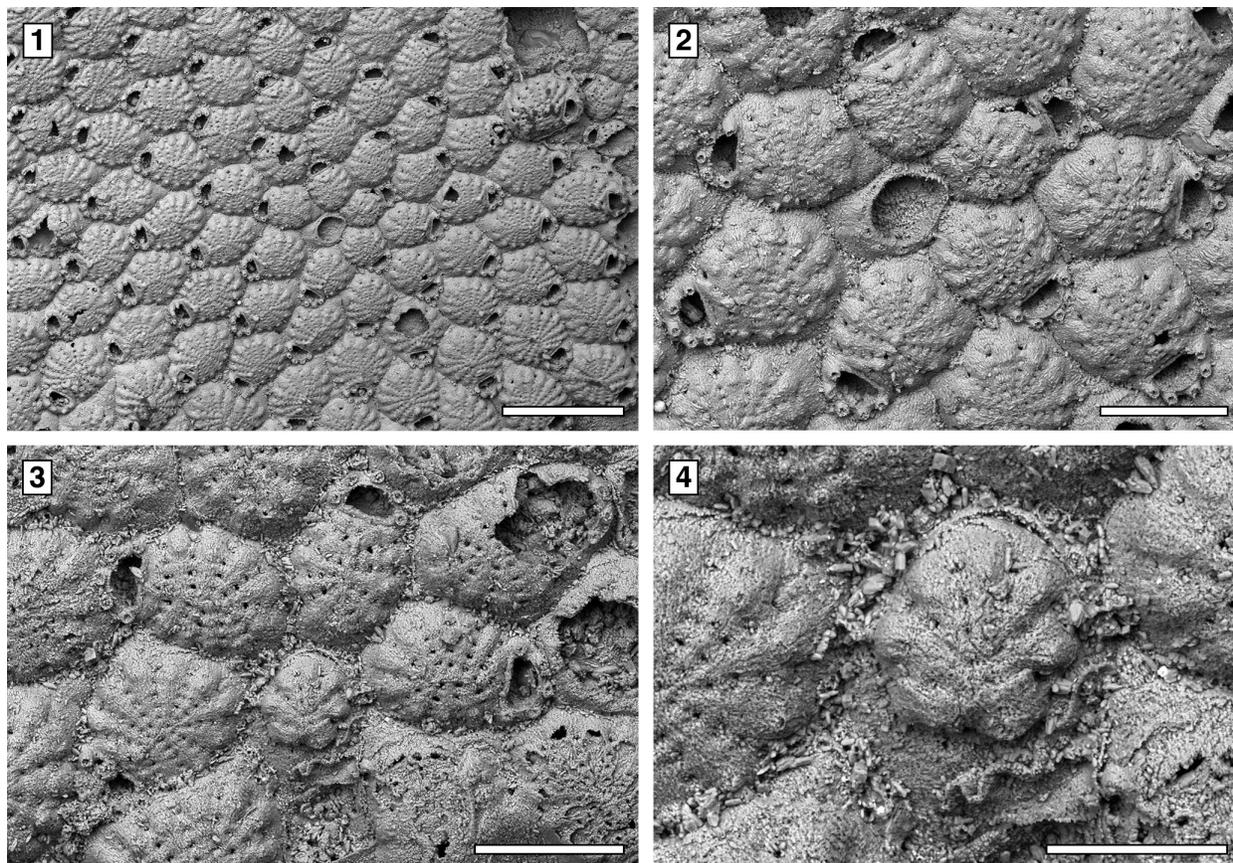


FIGURE 18. *Puellina scripta* (Reuss, 1848), Pliocene, lower Tamiami Formation Units 10/11. **1–2**, UF 305778 (Shell 22). **1**, tatform ancestrula and early astogeny (500 μ m). **2**, close-up of the tatform ancestrula and periancestrular zooids (200 μ m). **3–4**, UF 305779 (Shell 3). **3**, costulate kenozooid as regeneration of the ancestrula and first budded autozooids (200 μ m). **4**, close-up of the costulate kenozooid regenerated from the tatform ancestrula (100 μ m).

ston, 2005 has a more extensive gymnocyst surrounding the margins of the zooids and a greater number of intercostal lacunae; *P. testudinea* Winston, 2005 has long pointed avicularia (270 μ m long) and outer edges of costae raised into a tubercle; *P. capronensis* Winston, 2005 has smaller zooids (including orifice and ovicells), a large sub-oral pore, lacks avicularia and usually encrusts very small substrates such as sand or gravel grains; *P. minervae* Winston, 2016 has two large lacunae adjacent to the V-shaped first pair of costae.

A peculiar type of dissolution/abrasion has been observed in some specimens of *P. scripta* from the Tamiami Formation (Figure 13.4). Its origin is unknown.

Genus *Spiniflabellum* Di Martino and Rosso, 2015
Spiniflabellum laurae sp. nov.

Figure 19

zoobank.org/D90E131A-92DB-4DF0-9228-3AA17C6716AC

Type material. Holotype UF 305780 (Shell 30); paratype UF 305781 (Shell 135; not figured). Pliocene, lower Tamiami Formation.

Etymology. Named after Dr. Laura J. Cotton (University of Bristol) who helped to collect the specimens.

Diagnosis. Colony encrusting; autozooids elliptical. Frontal shield formed by 12–13 costae separated by four intercostal pores. One to three prominent pematidia on each costa. Distalmost pair of costae bifid and converging across the orifice. Primary orifice somewhat trapezoidal. Two platy elements, terminating in two coalescent oral spines located distolaterally, converging towards the zooid midline and touching distally. Pore-chamber windows visible at colony edges.

Description. Colony encrusting, multiserial, unilaminar (Figure 19.1). Pore-chamber windows visible along zooidal margins at colony growing edge, oval, small, 20–25 μ m long (Figure 19.1). Autozooids distinct with deep interzooidal furrows, ellipti-

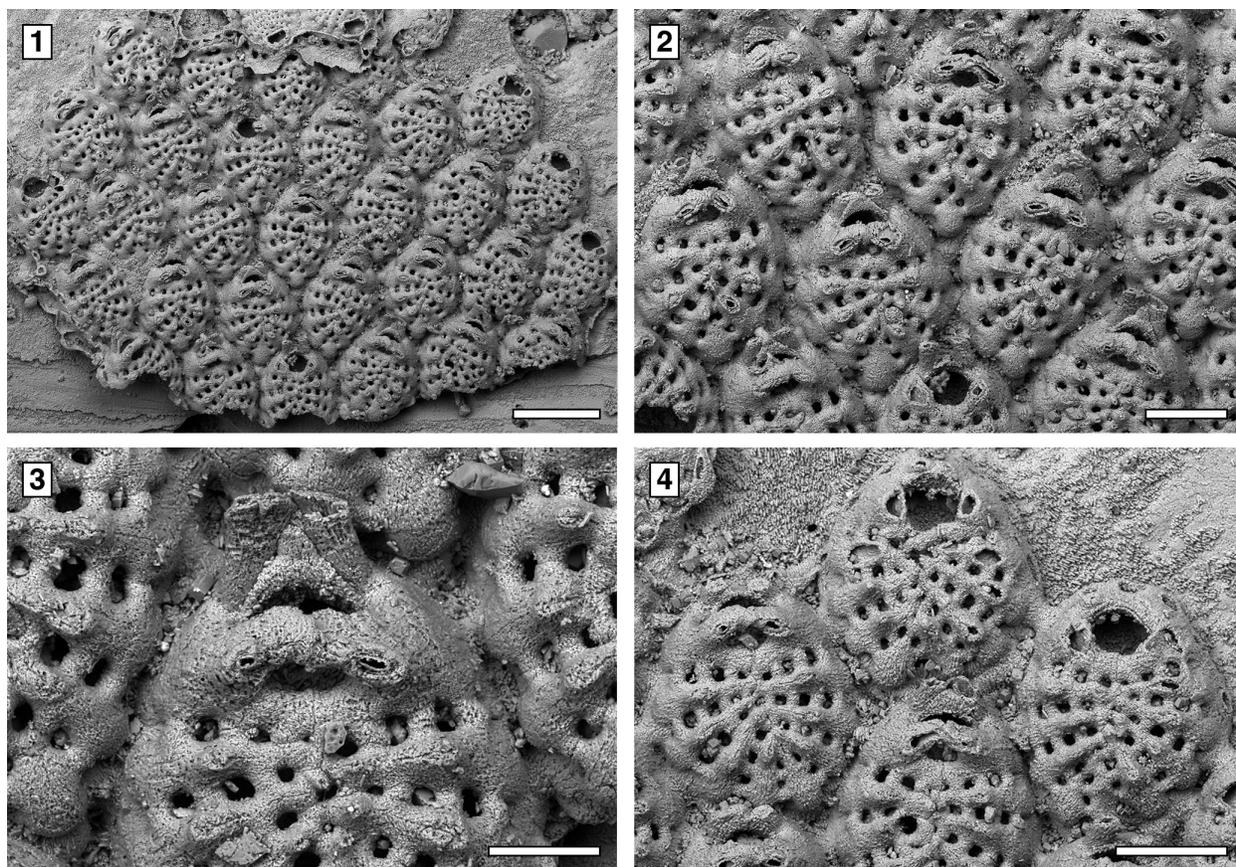


FIGURE 19. *Spiniflabellum laurae* sp. nov., holotype UF 305780 (Shell 30), Pliocene, lower Tamiami Formation Units 10/11. **1.** general view of the small colony (400 μm). **2.** group of autozooids (200 μm). **3.** close-up of the orifice over-arched by the distalmost pair of costae forming an eight-shaped pseudospiramen and bearing distally two platy elements terminating in two coalescent spines (100 μm). **4.** autozooids at colony growing edge showing pore-chamber windows and orifice (200 μm).

cal, longer than wide (mean L/W 1.32), quincuncially arranged. Frontal shield convex, formed by 12–13 narrow costae (20–30 μm), each separated by four oval to subcircular intercostal pores, 5–10 μm in diameter (Figure 19.2). Costae converging and fused at zooidal midline; 1–3 prominent tubular pematidia on each costa but usually broken off. Distalmost pair of costae smooth, bifid, the anterior bifurcation converging across the orifice, forming a wide arch and figure-eight-shaped to elliptical, proximomedian pseudospiramen (Figure 19.3), obscuring the primary orifice in frontal view; the arch bears two hollow pematidia. Primary orifice trapezoidal, about 45 μm long by 60 μm wide, with rounded distal margin and straight or slightly convex proximal margin (Figure 19.4). Two platy elements each terminating in two coalescent, hollow spines placed distolaterally to the orifice, converging and touching distally along zooidal midline (Figure 19.3). Avicularia, kenozooids and ovicells not observed. Ancestrula not seen.

Measurements (μm). ZL 274 \pm 19, 252–300 (1, 10); ZW 208 \pm 15, 190–230 (1, 10).

Remarks. *Spiniflabellum laurae* sp. nov. is the second fossil species of this genus to be described. An early Miocene species, *S. jacksoni* Di Martino, Taylor, and Portell, 2017, was described from the Chipola Formation in Florida. *Spiniflabellum laurae* sp. nov. differs from *S. jacksoni* in the number of costae forming the frontal shield (12–13 costae in the former species, 16–21 in the latter species), in the number of pematidia per costa (variably 1–3 in *S. laurae* sp. nov. and constantly single and central in *S. jacksoni*), and in the position of the coalescent spines, which are placed distolateral to the orifice, converging towards the zooidal midline and touching in the Pliocene species, but are always completely separated in the Miocene species. The recent *Spiniflabellum spinosum* (Canu and Bassler, 1928a), reported from Cuba and the Strait of Florida, differs from the new species in having 16–18 costae forming the frontal shield and a larger, char-

acteristically bat-shaped pseudospiramen. All species of *Spiniflabellum* are rare, with a single or a few colonies found, and all lack ovicells. However, more material needs to be found to confirm that the absence of ovicells is not a sampling artifact.

Superfamily Hippothooidea Busk, 1859
Family Trypostegidae Gordon, Tilbrook and
Winston in Winston, 2005
Genus *Trypostega* Levinsen, 1909
Trypostega composita sp. nov.

Figure 20

zoobank.org/B4CFD4C4-1A4F-4887-B720-9E6A54249424

Type material. Holotype UF 305782 (Shell 21); paratypes UF 305783 (Shell 261) and UF 305784 (Shell 17). Pliocene, lower Tamiami Formation.

Etymology. From the Latin 'compositus, -a, -um', meaning 'compound' and referring to the characteristic small subcolonies produced by frontal eruptive budding.

Diagnosis. Colony encrusting, sheet-like with frontal subcolonies. Basal pore-chambers present. Autozoid frontal shield gymnocrystal, smooth, evenly pseudoporous. Orifice dimorphic, cleithridiate; suboral umbo absent. Ovicell kenozooidal, globular, evenly pseudoporous. Zoeciules highly variable in size and shape, often distal or in-between autozooids, isolated or forming clusters.

Description. Colony encrusting, multiserial, uni- to multilaminar, sheet-like with several, small, circular subcolonies produced by frontal eruptive budding (Figure 20.1, 4). Basal pore-chamber windows visible in autozooids at colony growth edge, circular to transversely elliptical, variable in size, 20–35 µm long by 10–20 µm wide, evenly spaced (Figure 20.1, 6). Autozooids arranged quincuncially, distinct, with shallow interzooidal furrows, elongate (mean L/W 1.64), rounded rhomboidal or irregularly polygonal. Frontal shield slightly convex or flat, smooth, evenly perforated by 50–60 circular pseudopores, 5–10 µm in diameter; pseudopores distal to the orifice aligned in an arch. Orifice dimorphic, keyhole-shaped (cleithridiate), slightly longer than wide (mean L/W 1.22), with a horse-shoe-shaped anter separated by medially directed condyles from a U-shaped sinus, narrow and extended for one-quarter of total orifice length in non-ovicellate autozooids (Figure 20.3), but broader and shallower in ovicellate zooids (Figure 20.4); suboral umbo absent. Ovicell kenozooidal, globular, longer than wide, evenly perforated as the frontal shield of zooids, the associated zoeciule distinguished by its circular opening in the distal centre about 20 µm in diameter (Figure 20.5–7). Zoeciules often distal to an autozoid or laterally

between two autozooids, isolated or in clusters, variable in size and shape, with the frontal shield evenly pseudoporous and a larger, rounded, sinuate opening, 10–20 µm in diameter (Figure 20.2, 6–8). Closure plates often occur (Figure 20.8).

Measurements (µm). ZL 438±40, 376–510 (3, 20); ZW 267±31, 185–328 (3, 20); OL 81±4, 75–90 (3, 20); OW 66±3, 62–72 (3, 20); OvL 266±29, 224–310 (2, 18); OvW 226±17, 196–254 (2, 18); ZcL 179±60, 100–340 (3, 20); ZcW 137±54, 68–272 (3, 20).

Remarks. Canu and Bassler (1923) referred to specimens of *Trypostega* found in Neogene formations of North America as the widespread extant *Trypostega venusta* (Norman, 1903), which is now known to be limited to the northeast Atlantic Ocean, from southern Britain to Madeira and into the Mediterranean Sea (Tilbrook, 2006). *Trypostega venusta* differs from *T. composita* sp. nov. in having a well-developed, suboral umbo constantly present in each autozoid. Among Eocene North American species, *T. elongata* Canu and Bassler, 1920 differs in having carinated ovicells; *T. inornata* (Gabb and Horn, 1862) differs in having an imperforate frontal shield; and *T. undulata* Canu and Bassler, 1920 differs in having very distinct transverse undulations. The early Miocene *T. vokesi* Di Martino, Taylor, and Portell, 2017 from the Chipola Formation of Florida has fewer, larger pseudopores. Among recent Atlantic species, *T. ilhabelae* Winston and Vieira, 2013 differs in having a large, suboral umbo; *T. striatula* (Smitt, 1873) has marked longitudinal striations; and *T. tropicalis* Winston, Vieira and Woollacott, 2014 has a broader sinus.

Superfamily Adeonoidea Busk, 1884

Family Adeonidae Busk, 1884

Genus *Reptadeonella* Busk, 1884

Reptadeonella umbilicata (Lonsdale, 1845) comb. nov.

Figure 21

1845 *Cellepora umbilicata* Lonsdale, p. 507.

1923 *Adeona heckeli* Canu and Bassler, p. 158, pl. 24, figures 1, 2.

Figured material. UF 305785 (Shell 101); UF 305786 (Shell 18); UF 305787 (Shell 19). Pliocene, lower Tamiami Formation.

Description. Colony encrusting, multiserial, uni- or multi-laminar, sheet-like (Figure 21.1, 4). Autozooids delimited by slightly raised lateral margins, zooidal boundaries becoming unclear in later ontogeny owing to secondary calcification spreading from marginal areolar pores (Figure 21.6), arranged in alternating rows in unilaminar colonies,

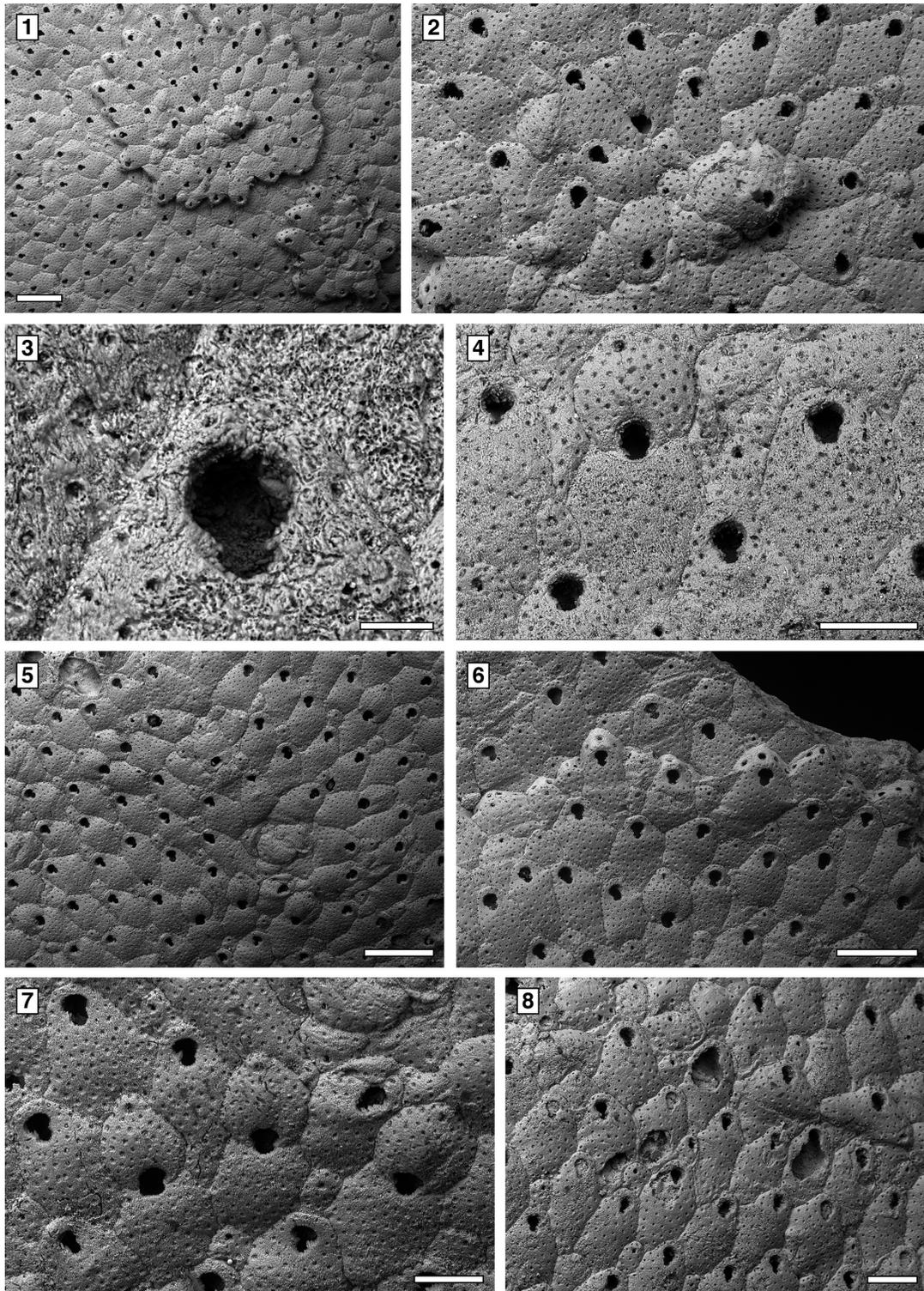


FIGURE 20. *Trypostega composita* sp. nov., Pliocene, lower Tamiami Formation Units 10/11. **1–2**, holotype UF 305782 (Shell 21). **1**, view of part of the colony with two subcolonies produced by frontal budding (500 μ m). **2**, group of autozooids and clustered zoeciules (200 μ m). **3–7**, paratype UF 305783 (Shell 261). **3**, close-up of the orifice of a non-ovicellate autozooid (40 μ m). **4**, group of ovicellate and non-ovicellate zooids showing the dimorphic orifices (200 μ m). **5**, group of ovicellate and non-ovicellate zooids and zoeciules (500 μ m). **6**, growing edge of a subcolony showing pore-chamber windows and zoeciules in clusters (400 μ m). **7**, group of ovicellate and non-ovicellate zooids (200 μ m). **8**, paratype UF 305784 (Shell 17), autozooids, some with closure plates (200 μ m).

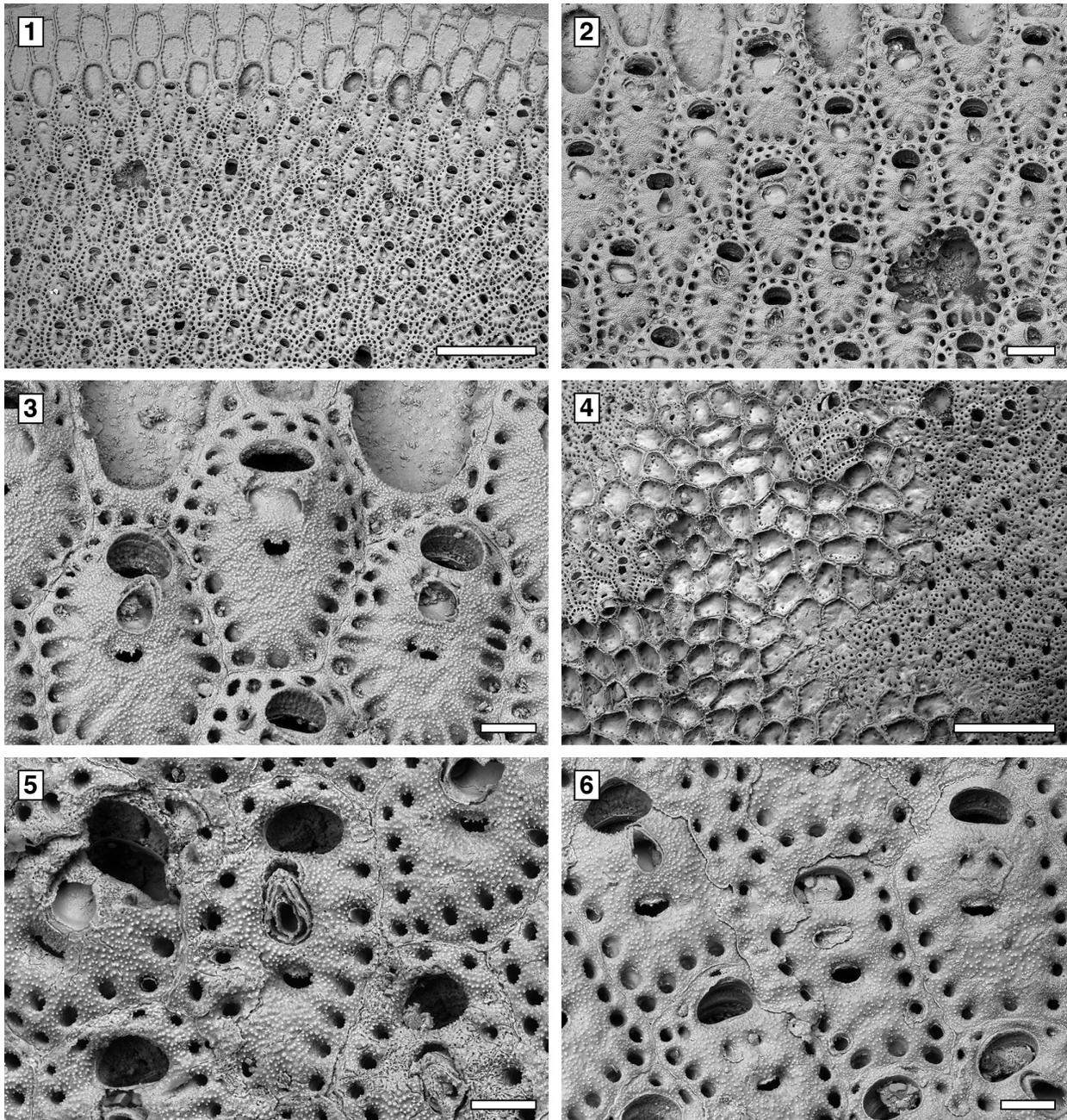


FIGURE 21. *Reptadeonella umbilicata* (Lonsdale, 1845) comb. nov., Pliocene, lower Tamiami Formation Units 10/11. **1–3**, UF 305785 (Shell 101). **1**, view of part of an extensive, unilaminar colony including the growing edge (1 mm). **2**, group of autozooids with or without the suboral avicularium and a fertile zooid in the centre (200 μ m). **3**, close-up of the fertile zooid (100 μ m). **4–5**, UF 305786 (Shell 18). **4**, view of part of a multilaminar colony (1 mm). **5**, close-up of an autozooid with intramural buds in the suboral avicularium (100 μ m). **6**, UF 305787 (Shell 19), group of autozooids mostly lacking a suboral avicularium and calcification spreading from marginal areolar pores (100 μ m).

irregularly in subsequent layers in multilaminar colonies, rounded polygonal, elongate (mean L/W 1.53). Frontal shield flat to slightly convex, finely granular, with a single (rarely double) row of circular, marginal areolar pores, 20–30 μ m in diameter.

Primary orifice transversely elliptical, sunken; secondary orifice transversely elliptical to bean-shaped, the margin bordered by a thin rim of smooth calcification; peristome not prominent at level with zooid surface. Suboral avicularium pres-

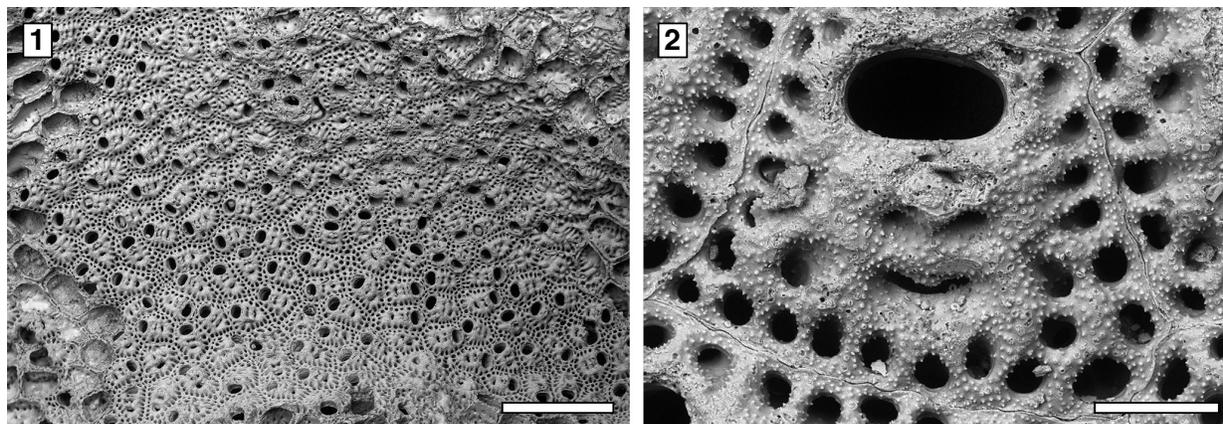


FIGURE 22. *Reptadeonella umbilicata* (Lonsdale, 1845) comb. nov., holotype NHMUK D53195, Pliocene, Petersburg, Virginia, USA. **1**, general view of the colony (1 mm). **2**, close-up of an autozoid lacking the suboral avicularium (100 µm).

ent or absent, drop-shaped, median, perpendicular or oblique on frontal shield, sloping proximally; rostrum pointed triangular, raised, directed distally or distolaterally, sometimes indenting the proximal margin of the secondary orifice; mandible hinged on two small condyles. If absent, in place of the avicularium one or two areolar pores are seen. No other avicularia. Spiramen crescent-shaped, placed 35–45 µm below the avicularium, about 60 µm wide by 15–20 µm long, distal margin with projecting denticles, usually set at level of zooid surface, seldom in a circular depression. Intramural buds common in suboral avicularia (Figure 21.5). Gonozooids similar to ordinary autozooids but with a transverse secondary orifice placed more proximally and overarched by a double row of marginal areolar pores distally; avicularium absent (Figure 21.2–3, 6). Ancestrula not seen.

Measurements (µm). ZL 468±41, 421–537 (2, 15); ZW 305±37, 252–360 (2, 15); OL 82±7, 69–91 (2, 15); OW 124±11, 114–148 (2, 15); AvL 101±9, 89–126 (2, 15); AvW 66±10, 55–87 (2, 15).

Remarks. With its multilayered encrusting colonies, this species fits better in the genus *Reptadeonella* Busk, 1884 than *Adeona* Lamouroux, 1812, which forms large, erect, cribrate colonies. Lonsdale (1845) described *R. umbilicata* from the Pliocene of Petersburg in Virginia. Although the holotype (D53195; Figure 22.1–2) lacks avicularia, all other characters resemble the Pliocene species from Florida. In both the Virginia (Figure 22.2) and Florida (Figure 21.3) colonies, among the zooids lacking avicularia only those with transverse orifices and a distal double row of marginal areolar pores are interpreted as gonozooids.

Superfamily Lepralielloidea Vigneaux, 1949

Family Lepraliellidae Vigneaux, 1949

Genus *Celleporaria* Lamouroux, 1821

Celleporaria sp.

Figure 23

Figured material. UF 305788 (Shell 31); UF 305789 (Shell 11); UF 305790 (Shell 107). Pliocene, lower Tamiami Formation.

Description. Colony encrusting, multiserial, mounded multilaminar (Figure 23.1, 4). Autozooids distinct, with deep interzooidal furrows, rectangular along the growing edge of the colony but erect and chaotically arranged in the central area of the colony, longer than wide (mean L/W 1.35). Frontal shield convex and smooth, imperforate apart from a few, small, circular marginal areolar pores scattered along the lateral zooidal margins, 20–30 µm in diameter. Orifice transversely D-shaped with concave proximal margin; two distolateral oral spine bases, 15–30 µm in diameter (Figure 23.2). Adventitious avicularium suboral, placed horizontally and parallel to one side of the proximal margin of the orifice, elliptical, laterally, and outwards directed; rostrum serrated and raised, crossbar complete. Interzooidal avicularia large, elliptical to spatulate, placed on a swollen cystid and irregularly scattered within the colony, randomly directed (Figure 23.2–4); rostrum rounded and raised, crossbar complete. Ovicells hood-shaped, widely open, imperforate, smooth, about 150 µm long by 320 µm wide (Figure 23.5).

Measurements (µm). ZL 546±46, 466–600 (2, 10); ZW 405±62, 324–513 (2, 10); OL 134±10, 118–152 (2, 10); OW 167±8, 155–177 (2, 10); AvL (suboral) 101±20, 80–147 (2, 10); AvW (suboral) 68±12, 57–

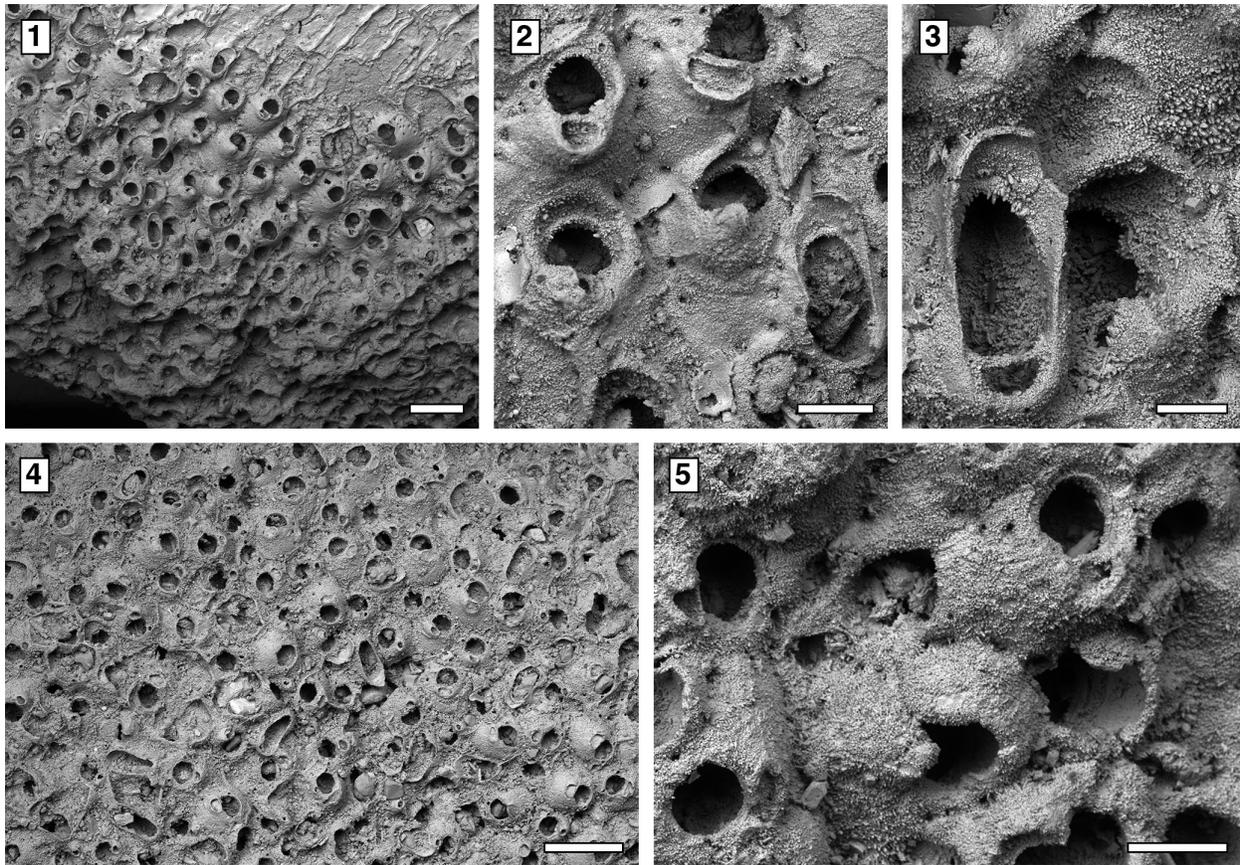


FIGURE 23. *Celleporaria* sp., Pliocene, lower Tamiami Formation Units 10/11. **1–3**, UF 305788 (Shell 31). **1**, general view of a multilaminar, mounded colony (600 μ m). **2**, close-up of autozooids with two, distolateral, oral spine bases and suboral avicularia (200 μ m). **3**, close-up of an interzooidal avicularium (100 μ m). **4**, UF 305789 (Shell 11), general view of a mounded colony with several interzooidal avicularia (500 μ m). **5**, ovicellate zooids (200 μ m).

96 (2, 10); AvL (interzooidal) 426 ± 29 , 381–460 (2, 10); AvW (interzooidal) 251 ± 22 , 222–286 (2, 10).

Remarks. *Celleporaria* cf. *bicornis* (Canu and Bassler, 1923) from the lower Miocene Chipola Formation of Florida (Di Martino et al., 2017) is similar in having two distolateral oral spine bases, a suboral avicularium with serrated rostrum, and large interzooidal avicularia with an elliptical to spatulate rostrum; it differs in having nodular frontal shields and ovicells, and an orifice with the proximal margin bearing three pointed denticles. *Celleporaria magnifica* (Osburn, 1914) has a similar orifice, but lacks oral spine bases, the interzooidal avicularia have duck-beak shaped rostra and sometimes a median triangular ligula, and zooidal length reaches 1 mm. The shape of the orifice is also similar in *C. albirostris* (Smitt, 1873). However, *C. albirostris* differs in having granular frontal shield and a prickly appearance produced by the sharply pointed avicularian rostra (Winston, 2005).

Family Bryocryptellidae Vigneaux, 1949
Genus *Cyclocolposa* Canu and Bassler, 1923
Cyclocolposa perforata Canu and Bassler, 1923
Figure 24

1923 *Cyclocolposa perforata* Canu and Bassler, p. 135, pl. 30, figures 6–14.

Figured material. UF 305791 (Shell 36); UF 305792 (Shell 10). Pliocene, lower Tamiami Formation.

Description. Colony encrusting, multiserial, unilaminar, sheet-like (Figure 24.1). Pore-chamber windows present (Figure 24.5). Autozooids distinct, with shallow furrows, arranged quincuncially, pentagonal to hexagonal, longer than wide (mean L/W 1.38). Frontal shield convex, coarsely granular, imperforate apart from one or two rows of circular marginal areolar pores, 15–25 μ m in diameter. Orifice subcircular, slightly wider than long (mean L/W 0.93), surrounded by a shortly raised, funnel-shaped peristome. An uncertain number (possibly four) of disto-oral spine bases present in early bud-

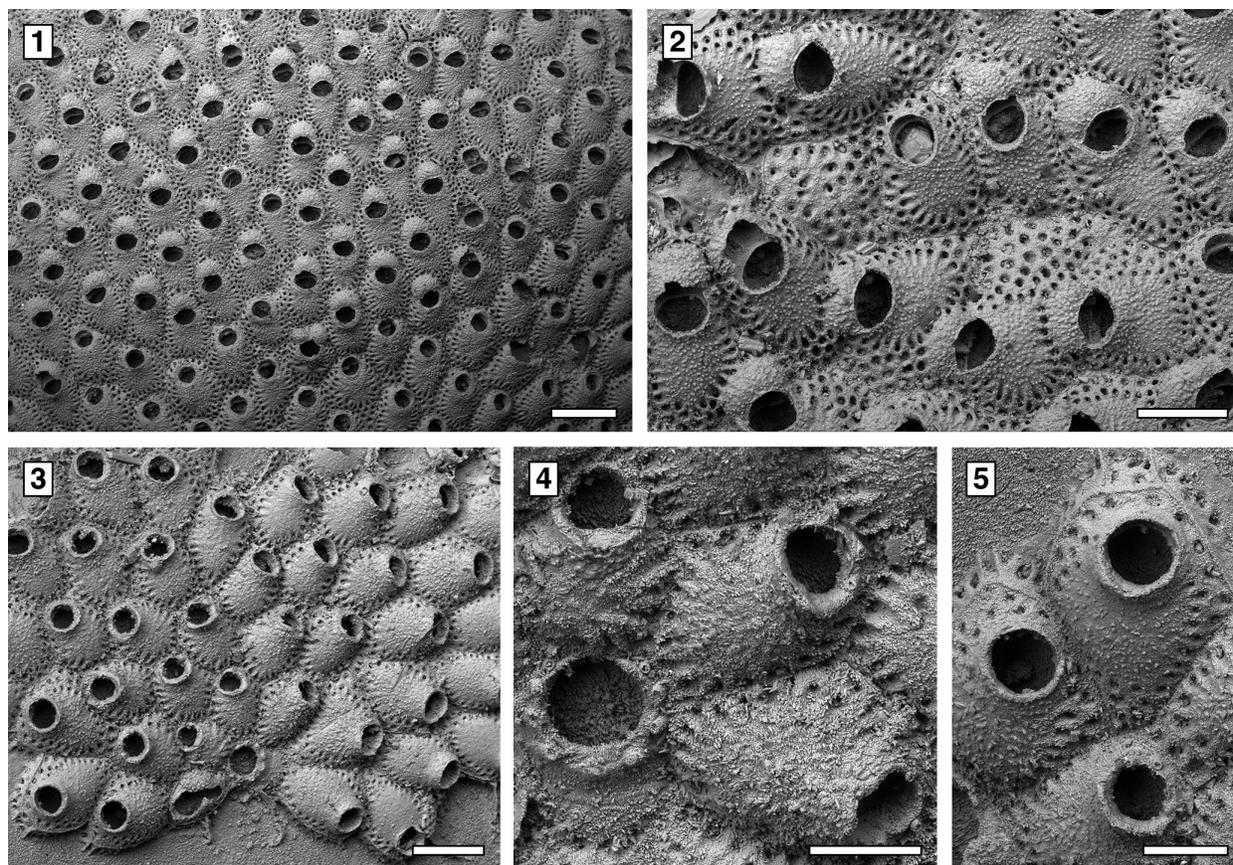


FIGURE 24. *Cyclocolpota perforata* Canu and Bassler, 1923, Pliocene, lower Tamiami Formation Units 10/11. **1–2**, UF 305791 (Shell 36). **1**, view of part of an extensive colony with most of the zooids ovicellate (500 μ m). **2**, group of ovicellate and non-ovicellate zooids and kenozooids (400 μ m). **3–5**, UF 305792 (Shell 10). **3**, tatiform ancestrula and early astogeny (400 μ m). **4**, close-up of the tatiform ancestrula and first budded autozooids (200 μ m). **5**, close-up of autozooids at the colony growing edge showing pore chamber windows (200 μ m).

ded autozooids up to the third generation, and randomly in later autozooids. Ovicells prominent, globular, slightly wider than long, imperforate and granular like the frontal shield (Figure 24.1–2). Avicularia absent. Kenozooids occasionally form at contacts between two colonies or fusing lobes of the same colony, irregularly shaped, with granular frontal shield lacking an opening but with up to three rows of areolar pores (Figure 24.2). Ancestrula tatiform, 320 μ m long by 280 μ m wide, surrounded by seven autozooids, a single zooid budded distally (330 μ m long by 285 μ m wide), and two zooids distolaterally (405–412 μ m long by 280–320 μ m wide), smaller than later autozooids; gymnocyst extensive proximally, occupying about one-third of the frontal surface; opesia transversely elliptical, 145 μ m long by 155 μ m wide, encircled by 12 oral spine bases (Figure 24.3–4).

Measurements (μ m). ZL 552 \pm 54, 462–638 (2, 20); ZW 394 \pm 50, 300–456 (2, 20); OL 168 \pm 13, 150–

192 (2, 14); OW 180 \pm 17, 144–215 (2, 14); OvL 210 \pm 28, 180–250 (1, 5); OvW 266 \pm 20, 238–292 (1, 5).

Remarks. Echols (1960) provided the unique report of *Cyclocolpota perforata* Canu and Bassler, 1923 from the Tamiami Formation, the species being reported previously only from the lower Pleistocene Waccamaw Formation of South Carolina. The genus name *Cyclocolpota* was first used by Canu and Bassler (1920, p. 431) and the type species – *C. perforata* (USNM 68617; Figure 25.1–3) – was mentioned, but not described, making both *Cyclocolpota* Canu and Bassler, 1920 and *C. perforata* Canu and Bassler, 1920 *nomina nuda*. The genus and species were validly published by Canu and Bassler (1923). *Cyclocolpota tenuiparietis* Canu and Bassler, 1923 (Figure 25.4–6; USNM 68618), from the Duplin Marl (now assigned to the upper Pliocene Jackson Bluff Formation) of Florida, differs from *Cyclocolpota perforata* in hav-

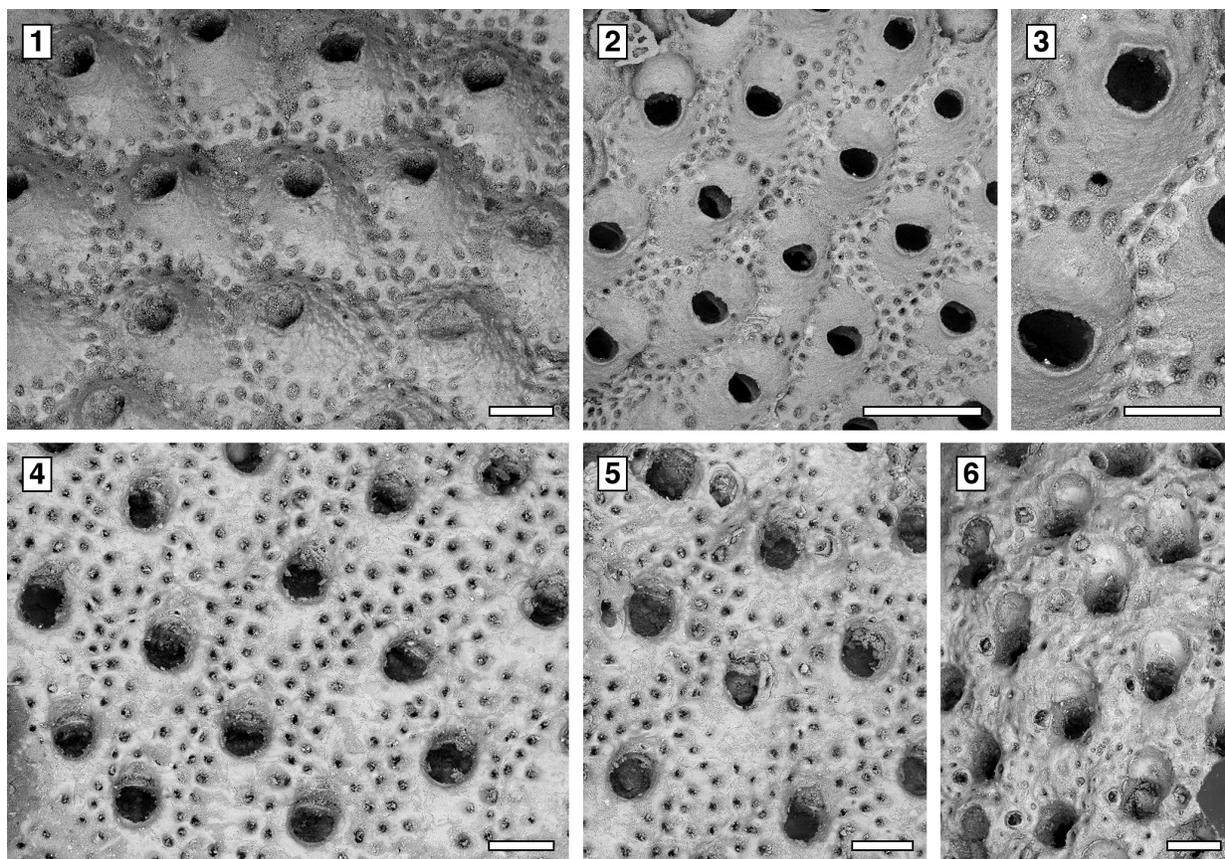


FIGURE 25. 1–3, *Cyclocolpota perforata* Canu and Bassler, 1923, syntype USNM 68617, early Pleistocene, South Carolina, USA. 1, group of autozooids (200 μm). 2, group of ovicellate zooids (500 μm). 3, close-up of an ovicellate and a non-ovicellate zooid (200 μm). 4–6, *Cyclocolpota tenuiparietis* Canu and Bassler, 1923, syntype USNM 68618, Pliocene, Florida, USA. 4, group of autozooids (200 μm). 5, group of autozooids, one with a latero-oral avicularium (200 μm). 6, group of zooids with incomplete ovicells (200 μm).

ing larger marginal areolar pores migrating towards the centre of the frontal shield and occasionally a round, latero-oral avicularium.

Family Romancheinidae Jullien, 1888

Genus *Hippopleurifera* Canu and Bassler, 1925

Hippopleurifera mucronata (Smitt, 1873)

Figure 26

- 1873 *Hippothoa mucronata* Smitt, p. 45, pl. 8, figure 169.
 2005 *Hippopleurifera mucronata* Winston, p. 54, figures 143–145.
 2017 *Hippopleurifera mucronata* Di Martino et al., p. 151, figure 41.

Figured material. UF 305793 (Shell 29). Pliocene, lower Tamiami Formation.

Description. Colony encrusting, multiserial, unilaminar, sheet-like (Figure 26.1). Pore-chamber windows present (Figure 26.2). Autozooids distinct, with shallow interzooidal furrows, rhomboidal to hexagonal, longer than wide (mean L/W 1.34).

Frontal shield convex, finely granular, with an imperforate central area, a row of marginal areolar pores and one or two less complete inner rows of pores, giving a faintly ribbed appearance. Marginal areolar pores subcircular, 20–30 μm in diameter. Orifice with two rounded condyles, dividing a semielliptical anter from a U-shaped sinus. Collar surrounding the orifice rises proximally in a pointed suboral umbo, often levelled-off, and bears 5–7 stout, closely spaced oral spine bases, 35–40 μm in diameter; only four spines visible in ovicellate zooids. Avicularia occasionally present, adventitious, placed laterally on the frontal shield at about mid-length (Figure 26.2, 4); rostrum rounded triangular, slightly curved, proximolaterally directed; no crossbar or condyles. Ovicells globose, slightly broader than long, with teardrop-shaped, marginal areolar pores, 40–50 μm long, between ribs of calcification radiating outward from a prominent, central tubercle (Figure 26.3–4).

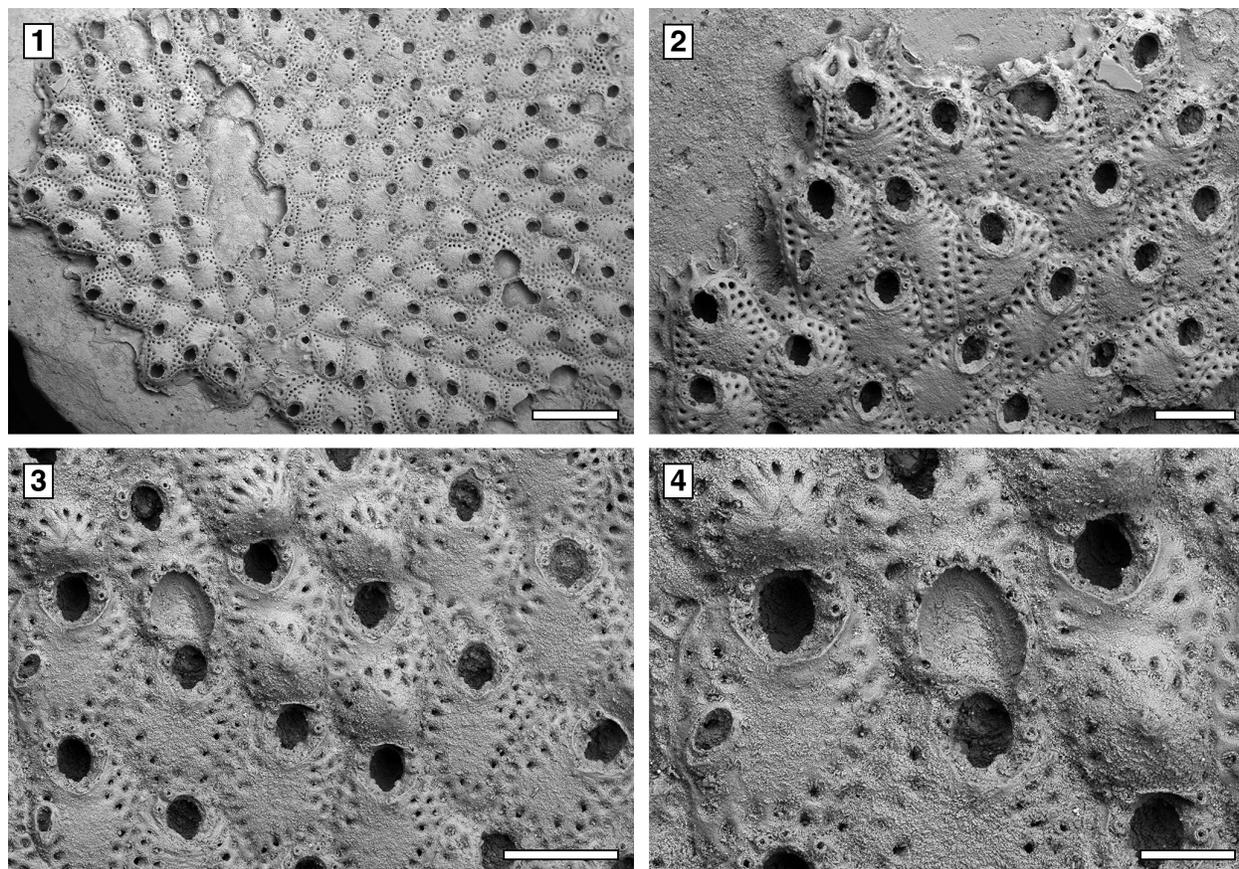


FIGURE 26. *Hippopleurifera mucronata* (Smitt, 1873), UF 305793 (Shell 29). Pliocene, lower Tamiami Formation Units 10/11. **1**, general view of the colony (1 mm). **2**, group of autozooids at colony growing edge showing pore-chamber windows (400 µm). **3**, group of ovicellate zooids (400 µm). **4**, close-up of ovicellate zooids, one with an adventitious avicularium (200 µm).

Measurements (µm). ZL 630±84, 510–862 (1, 20); ZW 470±49, 374–546 (1, 20); OL 173±12, 156–200 (1, 10); OW 131±8, 116–142 (1, 10); OvL 284±26, 232–322 (1, 10); OvW 303±22, 272–346 (1, 10); AvL 171±19, 150–188 (1, 5); AvW 102±11, 90–118 (1, 5).

Remarks. In addition to the Pliocene Tamiami Formation, the fossil record of *Hippopleurifera mucronata* includes the lower Miocene Chipola Formation of Florida (Di Martino et al., 2017) and the Pliocene Duplin Formation of North Carolina (Canu and Bassler, 1923). Although based only on a few zooids, the early Miocene specimen shows generally smaller measurements than the Tamiami Formation specimens. This difference might be related to temperature, the Chipola Formation representing a more tropical climate. At the present day, *H. mucronata* is a common inhabitant of the shelf off the southeast coast of the USA, Gulf of California, and the Caribbean (Winston, 2005).

Family Sfeniellidae Gordon, 2006
Genus *Stephanollina* Vigneaux, 1949
Stephanollina vorax (Canu and Bassler, 1923)
Figure 27

- 1923 *Gemelliporella vorax* Canu and Bassler, p. 111, pl. 19, figures 1–9.
2006 “*Gemelliporella*” *vorax* Gordon, pl. 2, figures 3, 4.
2017 *Stephanollina vorax* Di Martino and Taylor, p. 788.

Figured material. UF 305794 (Shell 34); UF 305795 (Shell 35); UF 305796 (Shell 22). Pliocene, lower Tamiami Formation.

Description. Colony encrusting, multiserial, unilaminar (Figure 27.1). Pore-chamber windows visible at colony growing edge, circular to transversely elliptical, variable in size, 30–70 µm long by 15–35 µm wide (Figure 27.1). Autozooids quincuncially arranged, rounded hexagonal, slightly longer than wide (mean L/W 1.15), distinct with shallow interzooidal furrows; zooidal boundaries becoming ill-

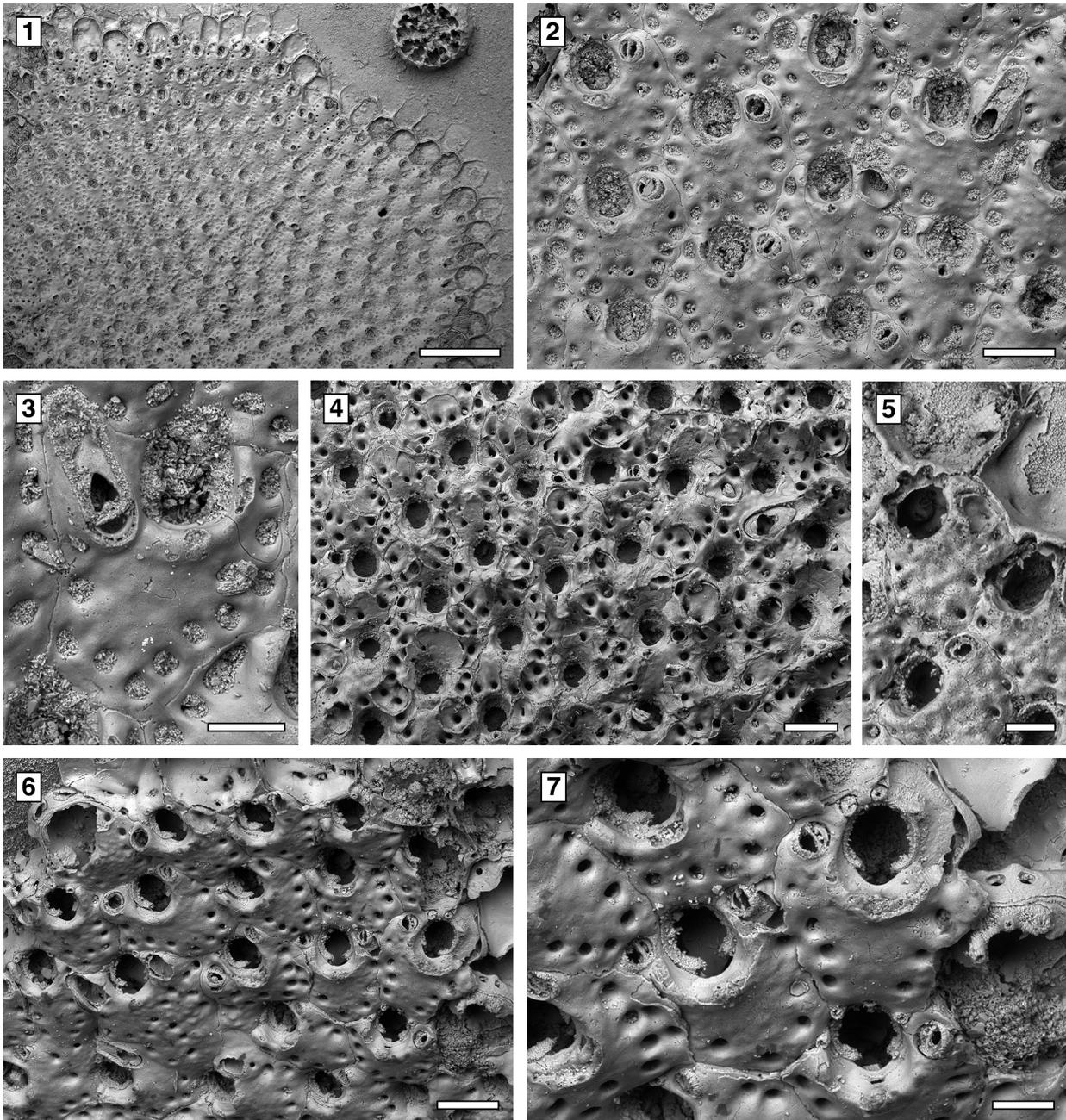


FIGURE 27. *Stephanollina vorax* (Canu and Bassler, 1923), Pliocene, lower Tamiami Formation Units 10/11. **1–3**, UF 305794 (Shell 34). **1**, general view of the colony (1 mm). **2**, group of autozooids with different types of adventitious avicularia (200 µm). **3**, close-up of an autozooid with a fusiform avicularium (100 µm). **4–5**, UF 305795 (Shell 35). **4**, general view of an 'old' part of a colony (200 µm). **5**, close-up of two autozooids showing the cormidial secondary orifice (100 µm). **6–7**, UF 305796 (Shell 22). **6**, group of autozooids at colony growing edge showing pore-chamber windows (200 µm). **7**, close-up of two autozooids (100 µm).

defined through ontogeny because of secondary calcification spreading from marginal areolar pores. Frontal shield slightly convex or flat, nodular, imperforate apart from a row of circular to drop-shaped marginal areolar pores, 15–50 µm long, along the proximal and lateral zooidal margins. Pri-

mary orifice longer than wide, keyhole shaped, with two robust, pointed condyles proximally placed and downwardly and proximomedially directed, separating an arched anter from a shallow, slightly narrower sinus with a concave proximal margin. Secondary orifice cormidial, formed by the frontal

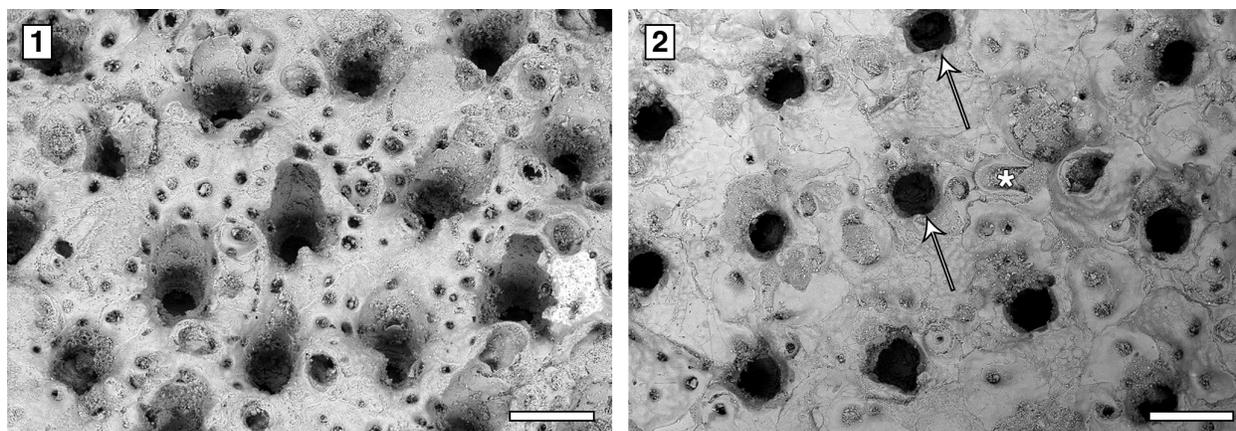


FIGURE 28. *Stephanollina vorax* (Canu and Bassler, 1923), syntypes. **1**, USNM 651310, Pleistocene, South Carolina, USA, group of autozooids (200 µm). **2**, USNM 651312, Pliocene, North Carolina, USA, group of autozooids with arrows indicating orifices in which condyles are preserved and asterisk indicating a fusiform avicularium partially covered by secondary calcification (200 µm).

shields of the surrounding distal zooids, forming a crenulated margin of lobes and cusps, the latter in the position of the oral spine bases (Figure 27.4–6). A small, pointed, suboral mucro develops with ontogeny (Figure 27.4). Commonly three oral spine bases, exceptionally four, about 20 µm in diameter. Adventitious avicularia budded from marginal areolar pores, distolateral to the orifice, single or paired, of two types; most commonly small, rounded to oval, placed on a swollen cystid sloping distally, directed laterally outwards or proximolaterally, with complete pivotal bar (type 1; Figure 27.2, 6); rarely large, fusiform to slightly spatulate, with arrowhead shaped opening, distally or distolaterally directed, the tip of the rostrum resting on the frontal shield of the next distal zooid, with complete pivotal bar (type 2; Figure 27.3–5). Avicularia lacking in some autozooids. Ovicells and ancestrula not observed.

Measurements (µm). ZL 393±48, 326–515 (2, 20); ZW 341±30, 286–404 (2, 20); OL 134±9, 120–146 (2, 10); OW 118±5, 111–123 (2, 10); AvL (type 1) 71±9, 60–85 (3, 10); AvW (type 1) 64±9, 54–80 (3, 10); AvL (type 2) 223±60, 158–295 (3, 5); AvW (type 2) 81±17, 65–110 (3, 5).

Remarks. *Stephanollina vorax* has been reported in the early Miocene through to the early Pleistocene of the eastern USA from North Carolina to Florida (Canu and Bassler, 1923). Di Martino and Taylor (2017) re-assigned the specimens of “*Gemelliporella*” *vorax* examined by Gordon (2006) to *Stephanollina* Vigneaux, 1949. These non-type specimens are from the Miocene of Wilmington in North Carolina and housed in the Canu Collection

at the Museum Nationale d’Histoire Naturelle, Paris. New SEM images of two syntypes (USNM 651310 separated from USNM 68575, and USNM 651312 separated from USNM 68576) (Figure 28) confirm this attribution. In the Tamiami Formation specimens ancestrulae and ovicells are lacking. Canu and Bassler (1923, p. 111) described the ancestrula as a small autozooid and the ovicell as “convex, little salient, deeply embedded in the distal zooecium”. Only incomplete ovicells were observed in the syntypes examined and in the specimens figured by Gordon (2006).

Superfamily Smittinoidea Levinsen, 1909

Family Smittinidae Levinsen, 1909

Genus *Smittioidea* Osburn, 1952

Smittioidea maleposita (Canu and Bassler, 1923)
comb. nov.

Figure 29

1923 *Smittina maleposita* Canu and Bassler, p. 144, pl. 31, figures 10, 11.

Figured material. UF 305797 (Shell 17); UF 305798 (Shell 258). Pliocene, lower Tamiami Formation.

Description. Colony encrusting, multiserial, multilaminar, sheet-like (Figure 29.1). Autozooids distinct with shallow interzooidal furrows, ill-defined later in ontogeny owing to frontal budding and secondary calcification, irregularly arranged, polygonal, almost as long as wide (mean L/W 1.03). Frontal shield flat to slightly convex, nodular, with one or two series of large conspicuous marginal areolar pores (Figure 29.2–3); areolae circular to elliptical, 40–45 µm long. Orifice slightly longer than wide, with triangular, pointed condyles proxi-

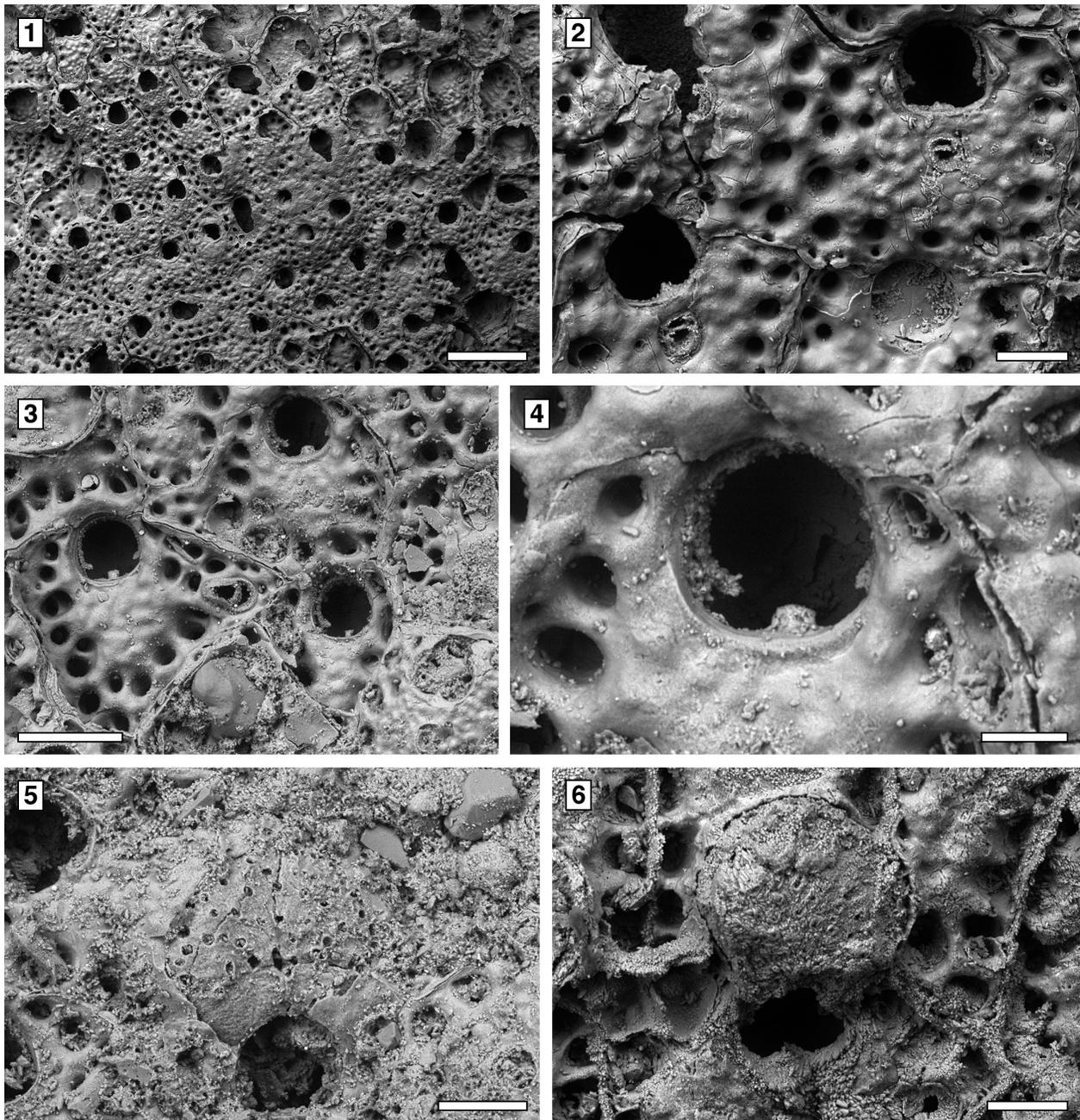


FIGURE 29. *Smittoidea maleposita* (Canu and Bassler, 1923) comb. nov., Pliocene, lower Tamiami Formation Units 10/11. **1–5**, UF 305797 (Shell 17). **1**, part of a multilaminar colony (500 μ m). **2**, close-up of two autozooids with median, suboral avicularium and closure plate (100 μ m). **3**, close-up of an autozooid with avicularium at the proximo-lateral corner (200 μ m). **4**, close-up of the orifice (50 μ m). **5**, close-up of an ovicell (100 μ m). **6**, UF 305798 (Shell 258), close-up of an ovicell (100 μ m).

mally directed (c. 15 μ m wide) and a rectangular lyrula occupying one-fourth of the proximal border (c. 25 μ m wide) (Figure 29.4). Oral spine bases absent in late ontogenetic zooids. Two types of adventitious avicularia, both with pivotal bar, are occasionally present: (1) common small, oval avicularia placed medially on the frontal shield, proxi-

mal to the orifice and directed proximally (Figure 29.2); (2) rare larger, pointed triangular avicularia, placed on the proximolateral zooidal corner and directed distolaterally (Figure 29.3). Ovicell globular, broader than long, 200 μ m long by 290 μ m wide; ectoecium calcified with scattered, small, circular pseudopores, 10–15 μ m in diameter, the

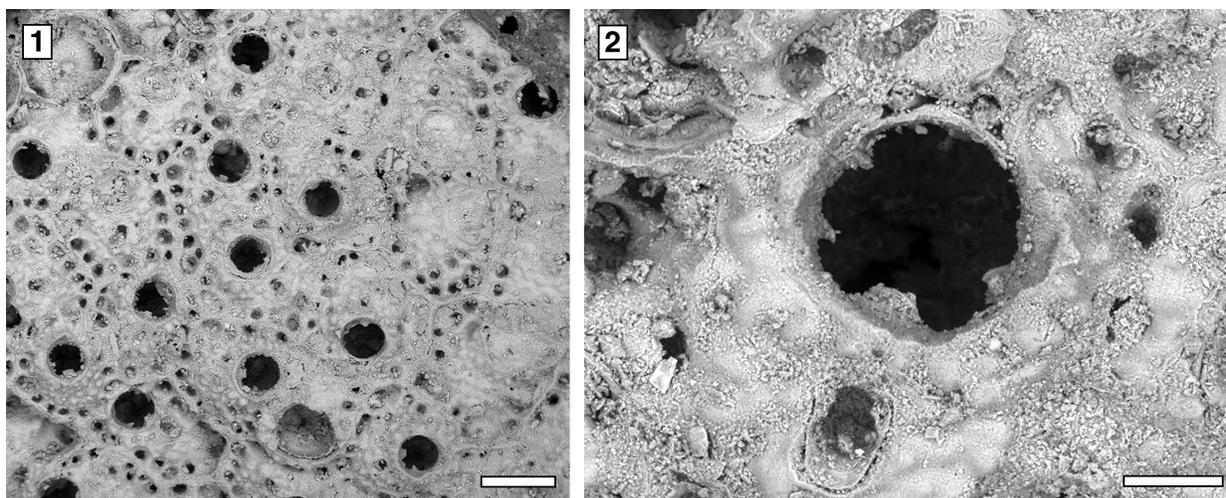


FIGURE 30. *Smittoidea maleposita* (Canu and Bassler, 1923) comb. nov., syntype, USNM 68641, Pleistocene, Florida, USA. **1.** group of autozooids (200 μ m). **2.** close-up of the orifice and suboral avicularium (50 μ m).

base covered by secondary calcification extending from the frontal shield of the next distal zooid (Figure 29.5–6). Closure plates common (Figure 29.2).

Measurements (μ m). ZL 389 \pm 60, 312–502 (3, 17); ZW 376 \pm 59, 294–487 (3, 17); OL 131 \pm 5, 127–139 (2, 5); OW 116 \pm 1, 114–117 (2, 5); AvL (type 1) 61 \pm 10, 46–78 (2, 8); AvW (type 1) 53 \pm 8, 45–64 (2, 8); AvL (type 2) 100 \pm 10, 93–107 (2, 2); AvW (type 2) 63 \pm 5, 59–66 (2, 2).

Remarks. The original generic attribution of this species seems unlikely because the genus *Smitina* is characterized by frontal shields that are uniformly pseudoporous. This species fits better in *Smittoidea*, which has frontal shields centrally imperforate, but with marginal areolar pores, and adventitious avicularia usually situated medially on the frontal, proximal to the orifice. *Smittoidea maleposita* comb. nov. is known only from the Pliocene–Pleistocene of Florida. Canu and Bassler (1923) described this species from the Caloosahatchee Formation at Shell Creek in the DeSoto County (Figure 30).

Family Bitectiporidae MacGillivray, 1895
Genus *Metroperiella* Canu and Bassler, 1917
Metroperiella reversa (Ulrich and Bassler, 1904)
comb. nov.
Figure 31

1904 *Lepralia?* *reversa* Ulrich and Bassler, p. 426, pl. 113, figures 1, 2.

1923 *Porella reversa* Canu and Bassler, p. 148, pl. 23, figures 5–10.

Figured material. UF 305799 (Shell 343); UF 305800 (Shell 23); UF 305801 (Shell 3); UF

305802 (Shell 461); UF 305803 (Shell 2). Pliocene, lower Tamiami Formation.

Description. Colony encrusting, multiserial, unilaminar, sheet-like (Figure 31.1, 5). Autozooids distinct, with shallow interzooidal grooves, arranged in alternating parallel rows, hexagonal to rectangular, longer than wide (mean L/W 1.74). Pore-chamber windows not seen. Frontal shield slightly convex, nodular, evenly perforated by 30–40 large, funnel-shaped pseudopores, 20–30 μ m in diameter, each situated at the center of a polygonal mesh. Orifice subcircular (mean L/W 0.98), with two small pointed condyles proximally directed defining a shallowly concave proximal sinus, surrounded by a thick, low peristomial rim (Figure 31.2). Oral spine bases absent. A single, small, adventitious avicularium with pivotal bar constantly present medially on the frontal shield, below the orifice touching the peristomial rim (Figure 31.1); rostrum pointed triangular, proximally directed. Closure plates occurring in both autozooids and avicularia (Figure 31.3). Ovicell globular, broader than long, united with the peristomial rim (Figure 31.3, 5); ectooecium calcified, smooth, with 10–15 large circular pseudopores, 15–35 μ m in diameter, the base covered by secondary calcification extending from the frontal shield of the next distal zooid. Irregularly shaped kenozooids lacking an opening present along the edges of juxtaposed colonies or filling gaps in the colony produced by self-overgrowth (Figure 31.4–5). Ancestrula tatiform, with an uncertain number of circumopesial spines usually overgrown by the six periancestrular autozooids (Figure 31.6); one distal and two distolateral autozooids budded directly

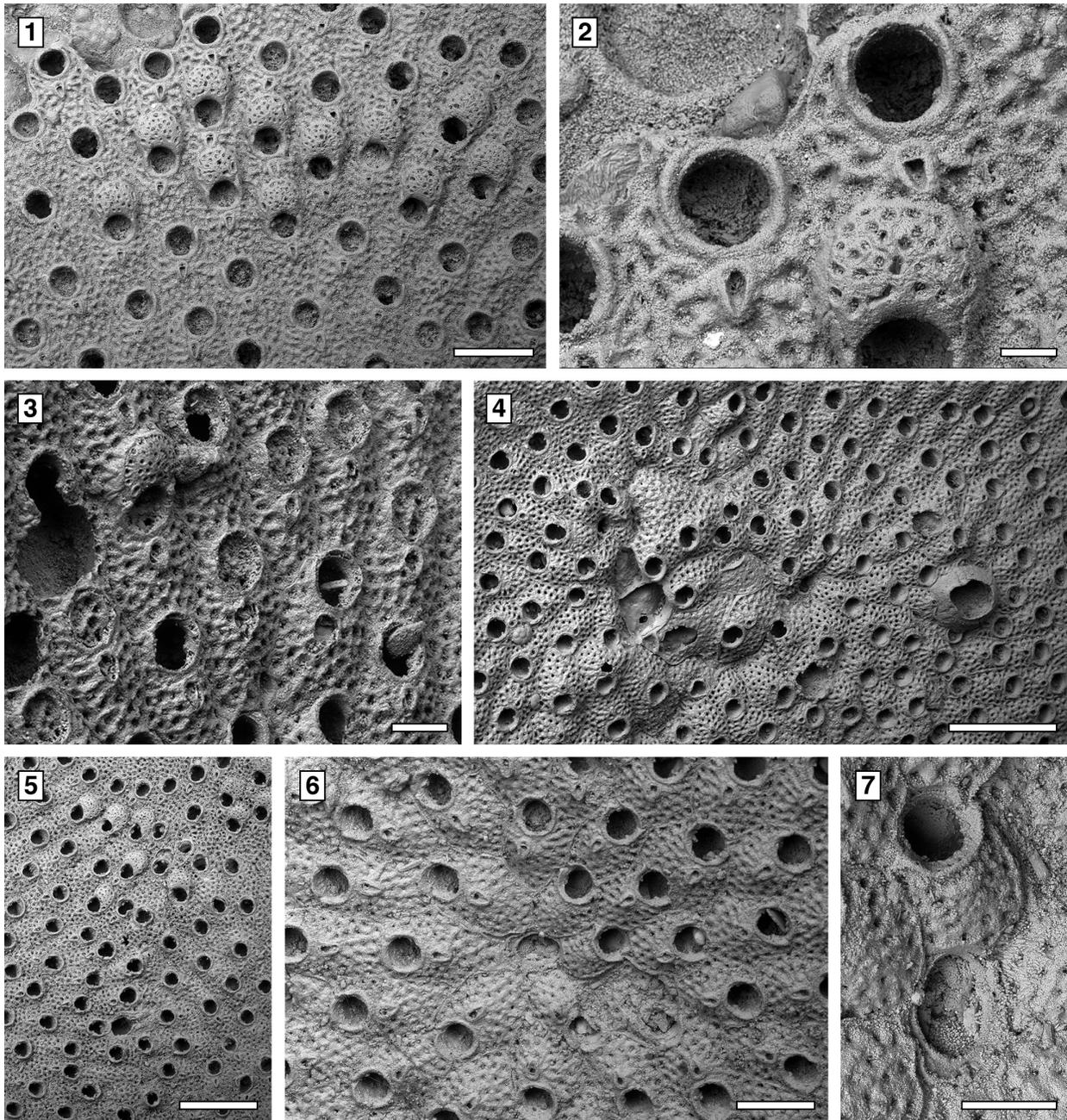


FIGURE 31. *Metroperiella reversa* (Ulrich and Bassler, 1904) comb. nov., Pliocene, lower Tamiami Formation Units 10/11. 1–2, UF 305799 (Shell 343). 1, group of ovicellate and non-ovicellate zooids (500 μ m). 2, close-up of two orifices with suboral avicularia and an ovicell (100 μ m). 3, UF 305800 (Shell 23), group of zooids with closure plates (200 μ m). 4, UF 305801 (Shell 3), group of autozooids and irregularly shaped kenozooids filling the gap resulting from self-overgrowth (1 mm). 5, UF 305802 (Shell 461), irregularly shaped kenozooids at the junction between two abutting colonies (1 mm). 6–7, UF 305803 (Shell 2). 6, tatiform ancestrula and early astogeny (400 μ m). 7, close-up of the tatiform ancestrula and first budded zooid (200 μ m).

from the ancestrula, smaller than later autozooids; distal periancestrular bud usually lacking an avicularium (Figure 31.7), which can be either present or absent in the two distolateral periancestrular buds.

Measurements (μ m). ZL 628 \pm 39, 576–697 (1, 10); ZW 360 \pm 34, 300–418 (1, 10); OL 180 \pm 11, 164–194 (1, 8); OW 183 \pm 13, 167–199 (1, 8); AvL 134 \pm 13, 118–166 (1, 12); AvW 90 \pm 8, 75–104 (1, 12); OvL

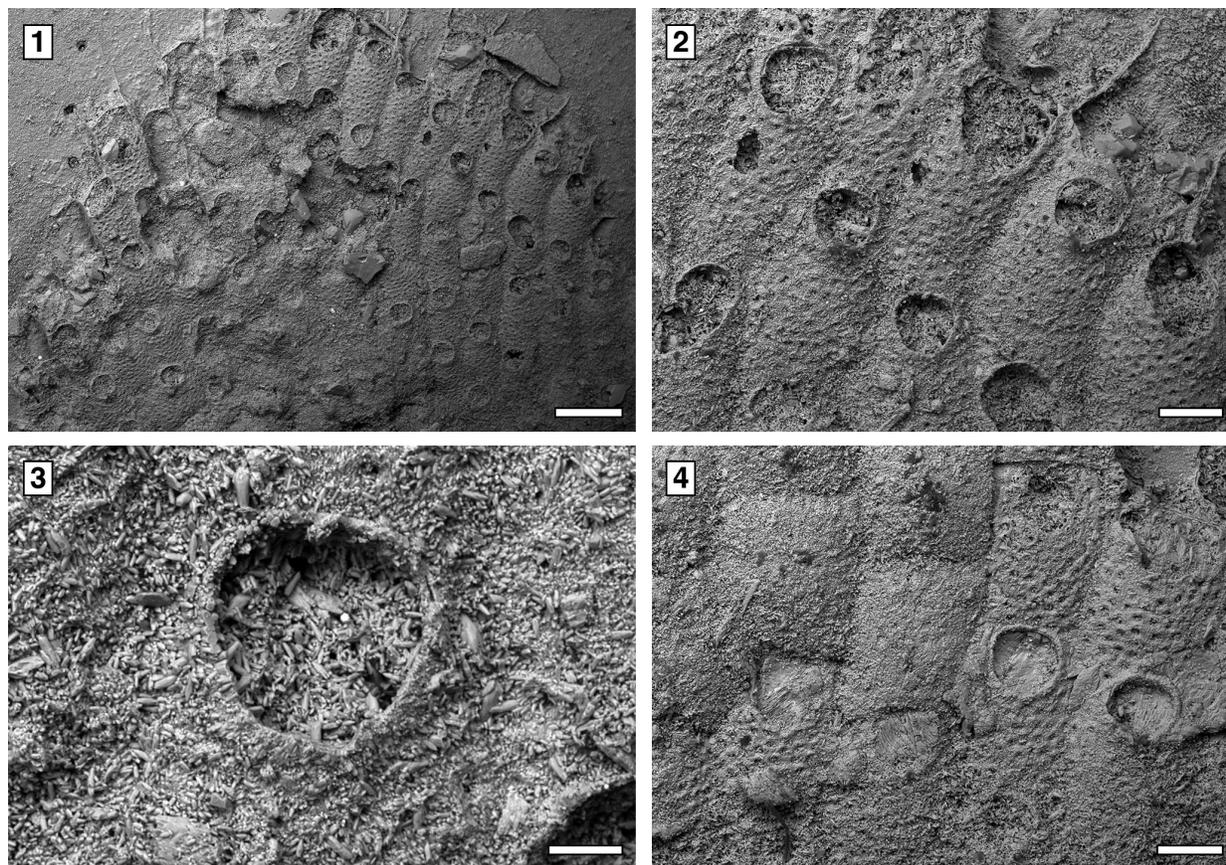


FIGURE 32. *Watersipora?* sp., UF 305804 (Shell 427), Pliocene, lower Tamiami Formation Units 10/11. **1.** part of a colony (500 µm). **2.** group of autozooids (200 µm). **3.** close-up of an orifice (50 µm). **4.** ?kenozooids lacking an aperture (200 µm).

214±14, 195–240 (1, 12); OvW 296±12, 280–315 (1, 12).

Remarks. This species was first described from the Miocene of Maryland and subsequently reported from the Pliocene of North Carolina (Canu and Bassler, 1923). It is better accommodated in *Metroperiella* Canu and Bassler, 1917 as *Porella* Gray, 1848, to which Canu and Bassler (1923) reassigned the species, is characterized by an imperforate frontal shield with marginal areolar pores only, a suboral avicularium usually embedded in the peristome and ovicells that are imperforate or have a single small pore.

Family Watersiporidae Vigneaux, 1949

Genus *Watersipora* Neviani, 1896

Watersipora? sp.

Figure 32

Figured material. UF 305804 (Shell 427). Pliocene, lower Tamiami Formation.

Description. Colony encrusting, multiserial, unilaminar (Figure 32.1). Autozooids distinct, with shallow interzooidal furrows, subrectangular, arranged

in alternating parallel rows, elongate (mean L/W 1.82). Frontal shield convex, granular, evenly pseudoporous (Figure 32.2); pseudopores numbering 45–55, rounded, about 10–15 µm in diameter. Orifice subcircular to oval, condyles seemingly present (Figure 32.3); oral spines absent. Avicularia, ovicells and ancestrula not observed. Apparent kenozooids lacking an aperture common (Figure 32.4).

Measurements (µm). ZL 640±50, 544–707 (1, 12); ZW 353±35, 312–402 (1, 12); OL 175±11, 160–190 (1, 8); OW 172±15, 152–198 (1, 8).

Remarks. The poor preservation of all available specimens precludes a more accurate identification for this species, which is tentatively referred to *Watersipora* based only on the shape of zooids and orifice and on the pseudoporous frontal shield. The specimen figured here is the only one preserving the shape of the orifice, although poorly and only for some zooids; all of the remaining specimens in the Tamiami Formation material consist of a few subrectangular ?kenozooids totally lacking

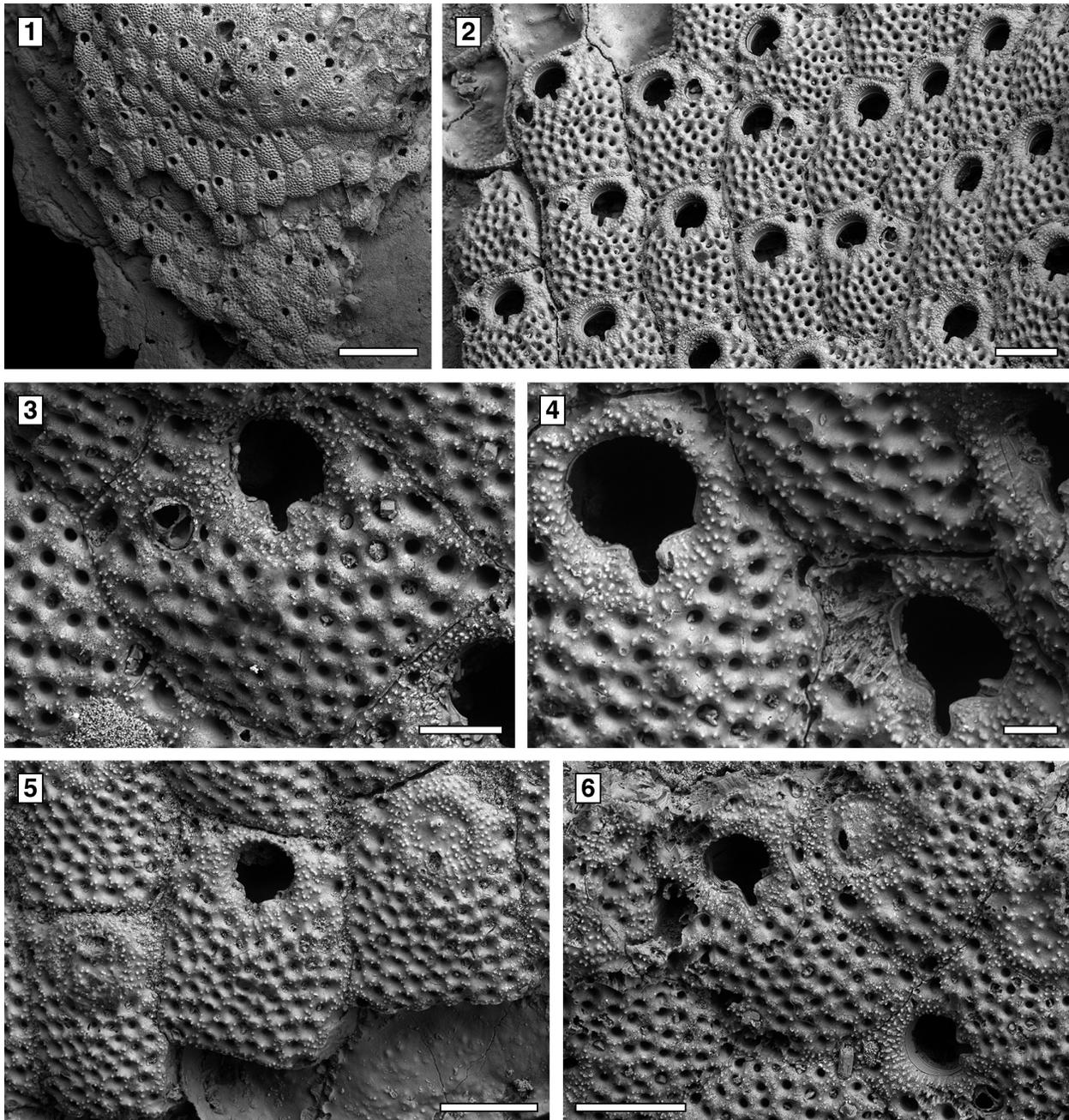


FIGURE 33. *Stylopoma leverhulme* Di Martino, Taylor, and Portell, 2017, UF 305805 (Shell 27), Pliocene, lower Tami-ami Formation Units 10/11. **1**, general view of a multi-layered colony (1 mm). **2**, group of autozooids (200 µm). **3**, close-up of an autozooid with an avicularium lateral to the orifice (100 µm). **4**, close-up of two orifices (50 µm). **5–6**, autozooids with closure plates (200 µm).

apertures. The general appearance of this species recalls some Neogene species that Canu and Bassler (1923) identified as *Dakaria*, which is now recognized as junior synonym of *Watersipora* (Vieira et al., 2014), and await revision.

Superfamily Schizoporelloidea Jullien, 1883
Family Schizoporellidae Jullien, 1883

Genus *Stylopoma* Levinsen, 1909
Stylopoma leverhulme Di Martino, Taylor, and
Portell, 2017
Figure 33

2017 *Stylopoma leverhulme* Di Martino, Taylor, and
Portell, p. 157, figure 46.

Figured material. UF 305805 (Shell 27). Pliocene, lower Tamiami Formation.

Description. Colony encrusting, multiserial, multilaminar (Figure 33.1). Autozooids with distinct interzooidal furrows, rectangular to irregularly polygonal, arranged in alternating parallel rows, slightly longer than wide (mean L/W 1.30). Frontal shield flat to slightly convex, sharply granular, evenly and densely perforated by 45–65 pseudopores, absent around the orifice (Figure 33.2–4); pseudopores circular, funnel-shaped, about 15 μm in external diameter and 12 μm in internal diameter, situated at the centres of a polygonal facets; marginal areolar pores placed at zooidal corners distinguished from pseudopores by being larger, 20–30 μm in diameter, and not evenly funnel-shaped. Primary orifice wider than long, 100 μm long (excluding sinus) by 120 μm wide, transversely D-shaped, proximal edge straight, sloping into a deep, slit-like sinus about 40 μm long by 20 μm wide (Figure 33.4); condyles robust, rectangular with rounded edges, smooth (Figure 33.6). Adventitious avicularium single or absent, small, teardrop-shaped, located laterally of the orifice, directed distally or distolaterally (Figure 33.2–3); rostrum pointed triangular, raised; crossbar complete. Vicarious avicularia and ovicells not observed. Closure plates common (Figure 33.5–6).

Measurements (μm). ZL 407 \pm 40, 342–476 (1, 15); ZW 312 \pm 46, 238–386 (1, 15); AvL 69 \pm 5, 65–76 (1, 4); AvW 57 \pm 2, 54–58 (1, 4).

Remarks. *Stylopoma leverhulme* was first described from the lower Miocene Chipola Formation of Florida. Although ovicells and vicarious avicularia were lacking in the few colonies available from the Tamiami Formation, the identity is confirmed by the zooidal measurements, the sharply granular appearance of the frontal shield, as well as the shape of the orificial sinus and the shape and position of the adventitious avicularium.

Family Stomachetosellidae Canu and Bassler, 1917

Genus *Cycloperiella* Canu and Bassler, 1923
Cycloperiella rubra Canu and Bassler, 1923
Figure 34

1923 *Cycloperiella rubra* Canu and Bassler, p. 137, pl. 21, figures 5–9.

Figured material. UF 305806 (Shell 15); UF 305807 (Shell 28). Pliocene, lower Tamiami Formation.

Description. Colony encrusting, multiserial, unilaminar, sheet-like (Figure 34.1, 4). Pore-chamber windows present along zooidal margins (Figure 34.1–2). Autozooids distinct, with shallow interzooi-

dal furrows, quincuncially arranged, hexagonal, longer than wide (mean L/W 1.33). Frontal shield convex, finely granular, densely perforated, except around the orifice, by 30–40 circular pseudopores, 15–20 μm in diameter; marginal areolar pores distinguishable at proximal and lateral zooidal corners, absent distally, larger than pseudopores, 25–45 μm in diameter. Orifice almost equidimensional, located terminally, the distal edge produced partly by the next distal zooid; two small, pointed condyles medially directed and proximally located separate a rounded anter from a broad, shallowly concave proximal margin (Figure 34.3). Oral spine bases absent. Avicularia adventitious, varying in number and distribution, placed laterally to the orifice or proximolaterally on the frontal shield, small, pear-shaped or pointed triangular, proximally or distolaterally directed (Figure 34.2, 6). Ovicells of the lepralielliform type, large, formed by the next distal zooid and occupying half of its frontal surface (Figure 34.4–6); ectooecium calcified, granular with numerous round pseudopores covered by the secondary calcification formed from the distal and distolateral neighbouring zooids and separated by thin raised sutures; a peristomial collar surrounds the orifice in ovicellate zooids (Figure 34.6). Ancistrula not observed.

Measurements (μm). ZL 545 \pm 53, 443–633 (3, 20); ZW 409 \pm 40, 320–461 (3, 20); OL 127 \pm 5, 119–136 (2, 12); OW 130 \pm 6, 122–143 (2, 12); AvL 108 \pm 12, 86–124 (2, 12); AvW 72 \pm 7, 61–84 (2, 12); OvL 228 \pm 14, 207–257 (1, 10); OvW 330 \pm 14, 308–353 (1, 10).

Remarks. Canu and Bassler (1923) reported *Cycloperiella rubra* from several Pliocene and Pleistocene deposits in North America, including the Pliocene Duplin Formation of North Carolina and the upper Pliocene Bowden Formation (now Bowden Member, Layton Formation) of Jamaica. The genus *Cycloperiella* was first described by Canu and Bassler (1920, p. 431) and the type species, *C. rubra*, mentioned as a *nomen nudum* (Figure 35; USNM 68620) before the authors made the name available three years later (Canu and Bassler, 1923, p. 137). The specific name '*rubra*' likely refers to the typical reddish colour of the fossils, which was probably the colour of the living colonies. Although the type material from the Pliocene of North Carolina has fewer pores (on average 20) in the frontal shield, we are skeptical that this is sufficient to introduce a new species for the Tamiami material until a more comprehensive revision has been undertaken of the genus.

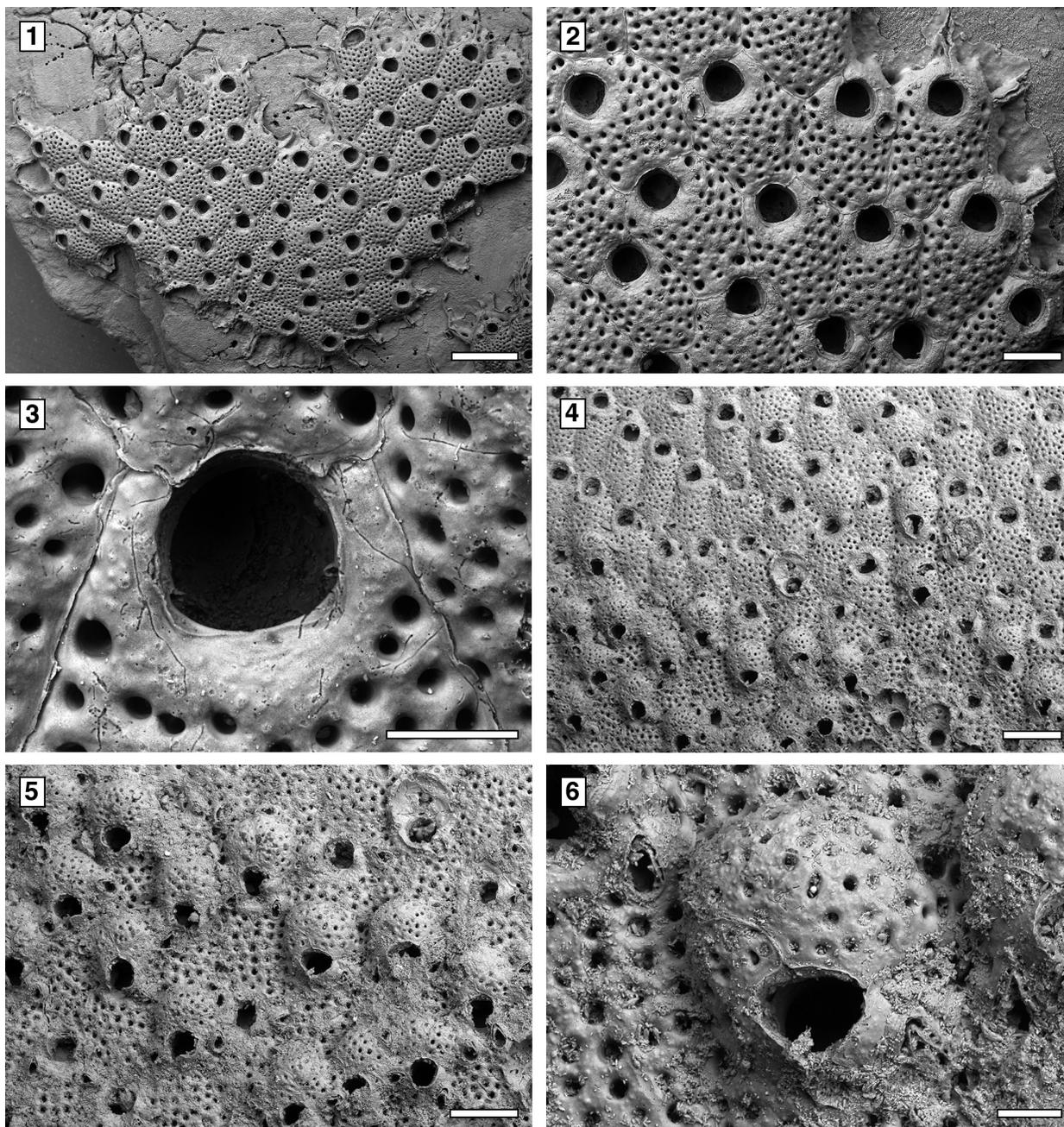


FIGURE 34. *Cycloperiella rubra* Canu and Bassler, 1923, Pliocene, lower Tamiami Formation Units 10/11. **1–3**, UF 305806 (Shell 15). **1**, general view of a colony (500 μ m). **2**, group of autozooids, some with avicularium lateral to the orifice (200 μ m). **3**, close-up of the orifice (100 μ m). **4–6**, UF 305807 (Shell 28). **4**, group of ovicellate and non-ovicellate zooids (500 μ m). **5**, group of ovicellate zooids (300 μ m). **6**, close-up of an ovicell with peristomial collar (100 μ m).

Family Hippaliosinidae Winston, 2005
 Genus *Hippaliosina* Canu, 1918
Hippaliosina rostrigera (Smitt, 1873)
 Figure 36

1873 *Escharella rostrigera* Smitt, p. 57, figures 203–205.

2005 *Hippaliosina rostrigera* Winston, p. 75, figures 200–206.

2017 *Hippaliosina rostrigera* Di Martino et al., p. 163, figure 50.

Figured material. UF 305808 (Shell 22). Pliocene, lower Tamiami Formation.

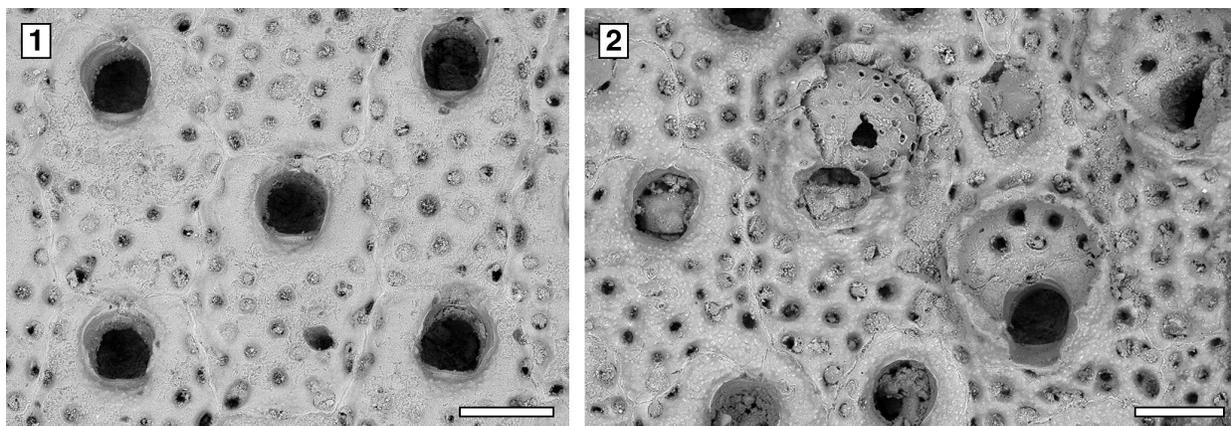


FIGURE 35. *Cycloperiella rubra* Canu and Bassler, 1923, syntype USNM 68620, Pliocene, North Carolina, USA. **1**, group of autozooids (200 μm). **2**, ovicellate zooids (200 μm).

Description. Colony encrusting, multiserial, unilaminar, sheet-like (Figure 36.1). Pore-chamber windows oval, 15–25 μm long, visible along zooidal margins. Autozooids distinct, bordered by narrow grooves, hexagonal to polygonal in shape with undulose margins, longer than broad (mean L/W 1.60). Frontal shield flat to slightly convex, coarsely granular, imperforate apart from a continuous row of marginal areolar pores, teardrop-shaped, outwardly sloping, 10–30 μm long. Orifice distally placed, keyhole-shaped (i.e., cleithriate), longer than broad; two pointed, downwardly directed condyles separating a horseshoe-shaped anter from a bowl-shaped sinus. Adventitious avicularia lateral to the orifice usually single, occasionally paired, their bases slightly above the proximal margin of the orifice or distally placed, distally and inwardly directed, pivotal bar absent (Figure 36.2). Maternal

zooids and kenozooids not observed. Ancestrula not seen.

Measurements (μm). ZL 402 \pm 54, 319–482 (1, 10); ZW 251 \pm 27, 208–285 (1, 10); AvL 131 \pm 18, 97–147 (1, 10); AvW 66 \pm 9, 43–77 (1, 10).

Remarks. *Hippaliosina rostrigera* ranges from the early Miocene to recent. It has been recorded in Virginia, North Carolina, South Carolina, and Florida, as well as Costa Rica. The zooidal measurements of the Tamiami Formation specimens are similar to those of the Chipola Formation specimens described by Di Martino et al. (2017), but smaller than the type specimen figured in Winston (2005).

Family Microporellidae Hincks, 1879

Genus *Microporella* Hincks, 1877

Microporella sarasotaensis sp. nov.

Figure 37

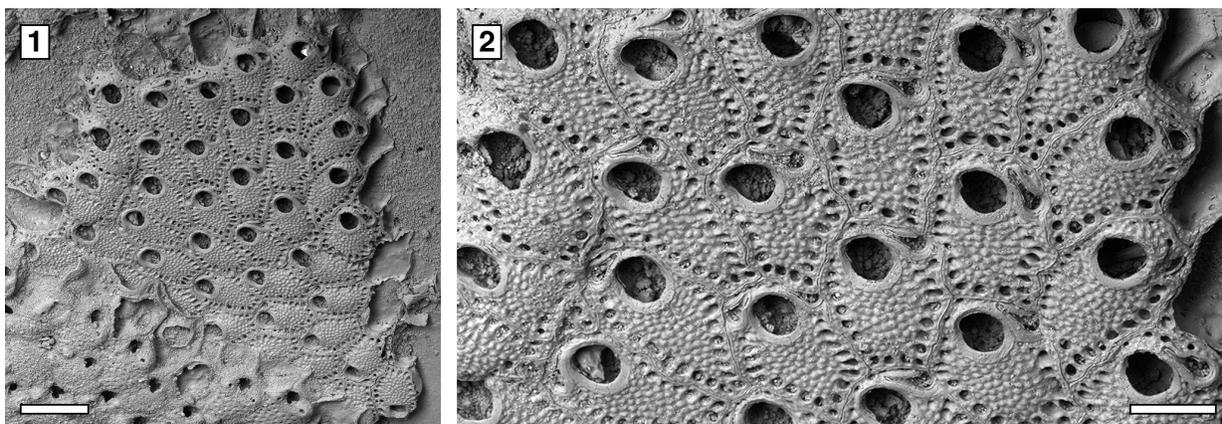


FIGURE 36. *Hippaliosina rostrigera* (Smitt, 1873), UF 305808 (Shell 22), Pliocene, lower Tamiami Formation Units 10/11. **1**, tilted view of a colony (400 μm). **2**, group of autozooids with adventitious avicularia (200 μm).

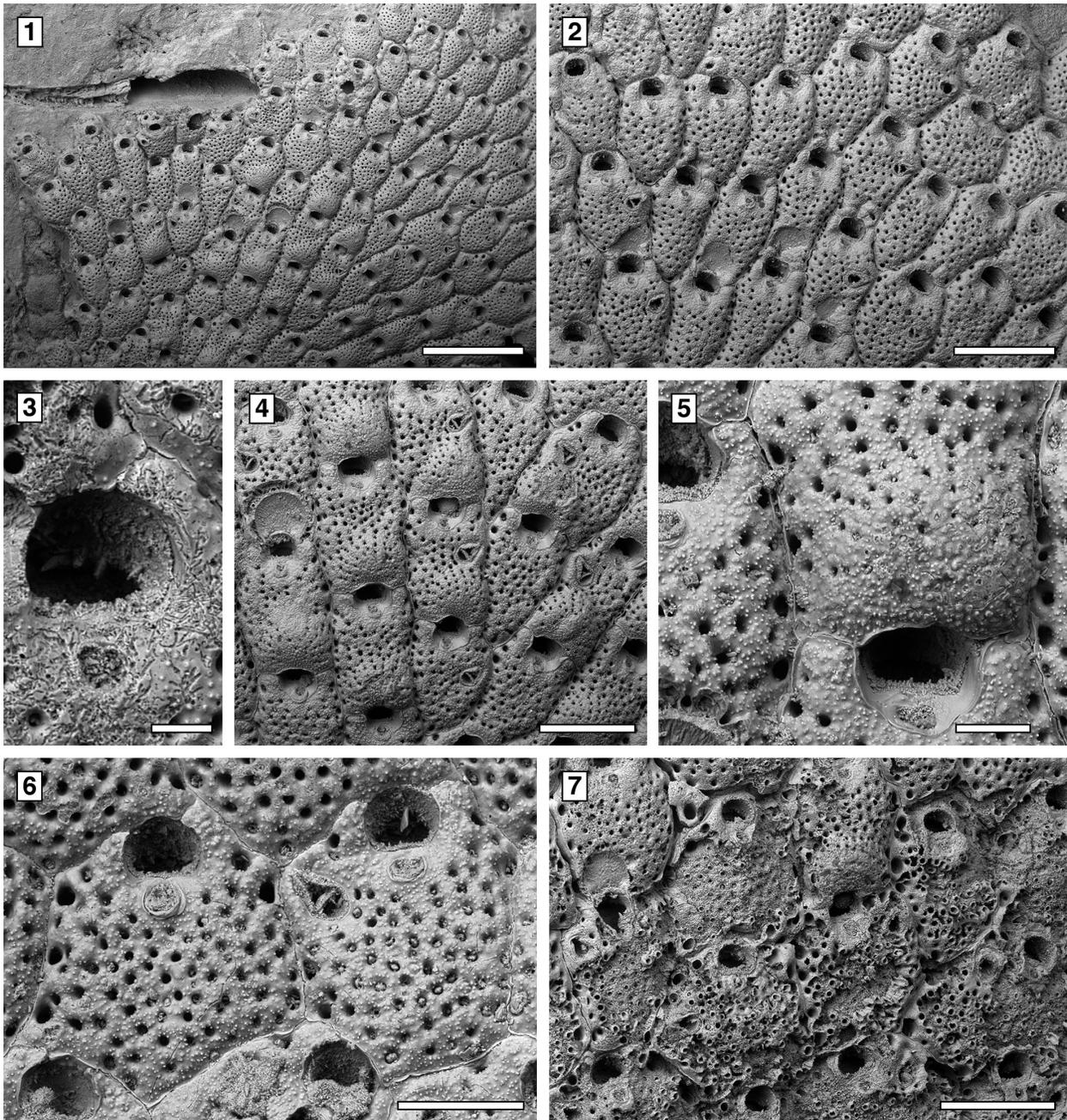


FIGURE 37. *Microporella sarasotaensis* sp. nov., Pliocene, lower Tamiami Formation Units 10/11. **1–5**, holotype UF 305809 (Shell 374). **1**, general view of part of the colony (1 mm). **2**, group of autozooids (500 μ m). **3**, close-up of the cribrate ascopore (50 μ m). **4**, group of ovicellate zooids (400 μ m). **5**, close-up of an ovicell (100 μ m). **6**, paratype UF 305810 (Shell 14), close-up of two autozooids (200 μ m). **7**, paratype UF 305811 (Shell 21), dissolution or abrasion of some zooids (400 μ m).

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Type material. Holotype UF 305809 (Shell 374); paratypes UF 305810 (Shell 14) and UF 305811 (Shell 21). Pliocene, lower Tamiami Formation.

Etymology. Named after the type locality, Sarasota.

Diagnosis. Colony encrusting forming extensive sheets. Pore-chamber windows present. Autozooids with finely granular and evenly pseudoporous frontal shield, except around the orifice and ascopore; marginal areolar pores at zooidal corners. Ascopore rounded, cribrate, adjacent to the orifice. Orifice semicircular, with denticulate proximal mar-

gin. Oral spines absent. Avicularium adventitious, single, located at about zooidal mid-length, triangular to acicular, directed distolaterally, with complete crossbar. Ovicell finely granular around an imperforate centre.

Description. Colony encrusting, multiserial, unilaminar, developing extensive sheets (>100 zooids) (Figure 37.1). Pore-chamber windows visible at colony growing edge, numerous, closely spaced, rounded to elliptical, 35–60 µm long by 25–30 µm wide (Figure 37.1). Autozooids distinct, with shallow interzooidal furrows and a thin lamina of smooth, raised calcification, hexagonal to rectangular or irregularly polygonal, commonly arranged in well-defined longitudinal rows (Figure 37.2, 4) but irregularly arranged when different lobes of the same colony converge, longer than wide (mean L/W 1.43). Frontal shield convex, finely granular, evenly pseudoporous, except for an imperforate band about 50 µm wide surrounding the orifice and including the ascopore (Figure 37.2); pseudopores 40–55, on average 45, circular, 10–15 µm in diameter; a few scattered marginal areolar pores located at zooidal corners, larger than pseudopores, rounded to elliptical, 30–40 µm long; ascopore rounded, 50–65 µm in diameter, cribrate (Figure 37.3), placed very close to the orifice, embedded in the proximal part of the peristome in ovicellate zooids (Figure 37.5), surrounded by a raised margin of smooth calcification, more extensive proximally. Orifice semicircular, with denticulate proximal margin, no condyles; oral spine bases absent (Figure 37.2, 6). Avicularia adventitious, generally single, sometimes absent, placed on either side of the autozooid, at about mid-length, much below the ascopore, teardrop-shaped (Figure 37.2, 4–6); rostrum rounded triangular, sometimes pointed acicular, directed distolaterally; crossbar complete; intramural buds observed in avicularia. Ovicell embedded in the distal zooid, finely granular proximally, with a few, scattered pseudopores distally, smaller than those on the frontal shield (Figure 37.4); orifice in ovicellate zooid hidden distally by the recumbent ovicell, deeper than in non-ovicellate zooids because of the development of a peristome that also includes the ascopore (Figure 37.5). Ancestrula not seen.

Measurements (µm). ZL 535±63, 424–672 (2, 20); ZW 375±52, 293–489 (2, 20); OL 108±10, 95–128 (2, 15); OW 138±12, 122–159 (2, 15); AvL 112±14, 86–132 (2, 20); AvW 77±10, 61–97 (2, 20); OvL 246±22, 213–281 (2, 15); OvW 323±24, 289–385 (2, 15).

Remarks. Among fossil and modern western Atlantic species of *Microporella*, only a few lack oral spines, which is one of the main features of *M. sarasotaensis* sp. nov., distinguishing it from congeneric species found in the Tamiami Formation material. *Microporella bifoliata* Ulrich and Bassler, 1904 and *M. inflata* Ulrich and Bassler, 1904 both from the Miocene of Maryland, differ in having erect, bifoliate colonies and smooth-surfaced zooids and ascopores more distant from the orifice, respectively; *M. cucullata* Canu and Bassler, 1928b, from Brazil differs in having a distally raised peristome; *M. normani* Canu and Bassler, 1928a, allegedly found alive in Madeira and as a fossil in the Pliocene of Panama, differs in having a larger avicularium closer to the orifice and a crescentic ascopore.

Microporella tamiamiensis sp. nov.

Figure 38

zoobank.org/E368247E-2354-4357-99DE-C9A8AE397372

Type material. Holotype UF 305812 (Shell 3); paratype UF 305813 (Shell 29). Pliocene, lower Tamiami Formation.

Etymology. Named after its type lithostratigraphic unit, the Tamiami Formation.

Diagnosis. Colony encrusting, forming extensive sheets. Pore-chamber windows present. Autozooids with finely granular and evenly and densely pseudoporous frontal shields, except around the orifice and ascopore. Ascopore crescentic, denticulate, placed in a depression below the orifice and surrounded by a slightly raised collar. Orifice semicircular, with denticulate proximal margin. Oral spines 3–5, commonly 3, hidden in ovicellate zooids. Avicularium adventitious, single, located at zooidal lateral corners, triangular, directed distolaterally, with complete crossbar. Ovicell evenly pseudoporous; peristome developing as two lateral flaps.

Description. Colony encrusting, multiserial, unilaminar, developing extensive sheets (>100 zooids) (Figure 38.1). Pore-chamber windows visible at colony growing edge, numerous, closely spaced, rounded to elliptical, 30–50 µm long by 25–30 µm wide (Figure 38.3, 5). Autozooids distinct, with shallow interzooidal furrows, rounded hexagonal to irregularly polygonal, quincuncially arranged, slightly longer than wide (mean L/W 1.22). Frontal shield convex, finely granular, evenly and densely pseudoporous, except for an imperforate band about 95 µm large, surrounding the orifice and the ascopore (Figure 38.2–3); pseudopores 80–110, on average 90, circular, 12–20 µm in diameter; marginal areolar pores undistinguished from pseu-

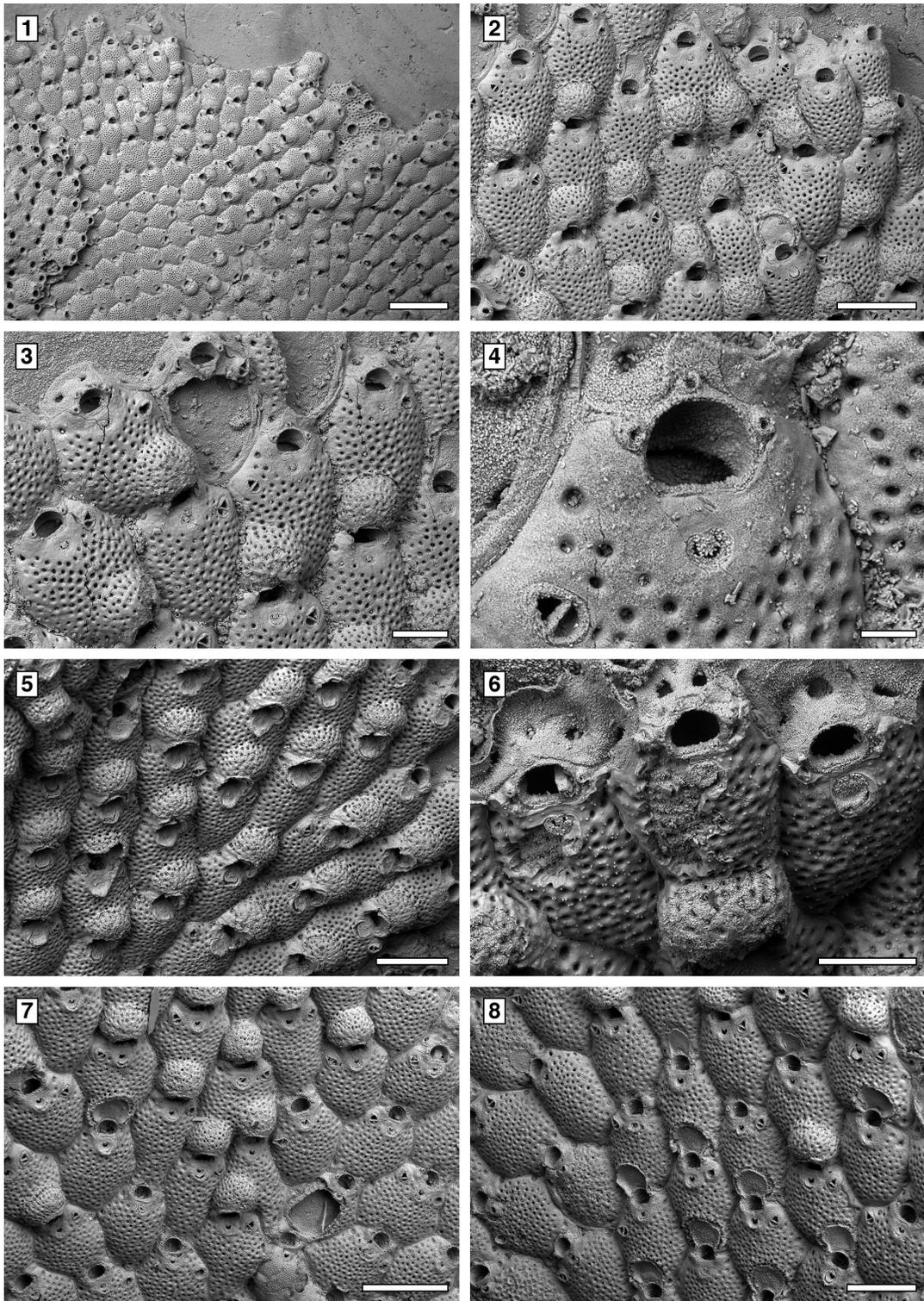


FIGURE 38. *Microporella tamiamiensis* sp. nov., Pliocene, lower Tamiami Formation Units 10/11. **1–6**, holotype UF 305812 (Shell 3). **1**, general view of part of the colony (1 mm). **2**, group of ovicellate and non-ovicellate zooids (500 µm). **3**, group of zooids at colony growing edge showing pore-chamber windows (200 µm). **4**, close-up of an orifice with three oral spine bases (50 µm). **5**, group of tilted ovicellate zooids showing the lateral flaps (400 µm). **6**, close-up of zooids showing pore-chamber windows (200 µm). **7–8**, paratype UF 305813 (Shell 29), group of zooids showing a more developed proximal rim of the ascopore (**7**, 500 µm; **8**, 400 µm).

dopores, except for a few cases when located at zooidal corners, slightly larger and circular, about 25 µm in diameter; ascopore crescent-shaped, 35–65 µm in diameter, denticulate, placed in a depression about 35 µm below the orifice and surrounded by a slightly raised collar, often more developed proximally (Figure 38.4–5). Orifice semicircular, with denticulate proximal margin (Figure 38.4); no condyles. Oral spine bases 3 (one mediiodistal and two mediolateral; Figure 38.4) or 4 (one distal pair and one mediolateral pair; Figure 38.3), most commonly 3, 5 observed only once, 20–30 µm in diameter. Avicularia adventitious, generally single, sometimes absent, located on either side of the autozoid, commonly placed at the lateral corner at the same level as the ascopore, rarely at zooidal mid-length below the ascopore, teardrop-shaped (Figure 38.3–4); rostrum triangular, directed distolaterally; crossbar complete. Ovicell embedded in the proximal part of the distal zooid, finely granular and evenly pseudoporous (Figure 38.3, 7–8); no spines visible in ovicellate zooids, hidden by the development of a peristome consisting of two lateral flaps (Figure 38.5). Ancestrula not seen.

Measurements (µm). ZL 512±46, 444–652 (2, 20); ZW 419±52, 333–518 (2, 20); OL 82±6, 75–94 (2, 16); OW 102±4, 94–108 (2, 16); AvL 79±8, 64–95 (2, 15); AvW 54±7, 42–66 (2, 15); OvL 197±16, 173–231 (2, 20); OvW 253±20, 220–291 (2, 20).

Remarks. At first sight *Microporella tamiamiensis* sp. nov. is apparently very similar to its sympatric congener *Microporella sarasotaensis* sp. nov. Indeed, both species form very extensive sheets and the zooids are of similar length. They also share the finely granular frontal shield of the zooids and the denticulate proximal border of the orifice. However, there are several differences. In *M. tamiamiensis* sp. nov. the zooids are wider and have a lower L/W ratio (1.22 vs 1.43), orifices and avicularia are both smaller, frontal shields have double the number of pseudopores (on average 90 vs 45), oral spines are present, ovicells evenly pseudoporous, peristomes developed laterally as two flaps, and the ascopore is denticulate. Among fossil and living western Atlantic species with 3–4 oral spine bases, the following species differ from *M. tamiamiensis* sp. nov.: *Microporella curta* Almeida, Souza, Menegola, and Vieira, 2017 has zooids separated by raised walls, a nodular frontal shield and personate ovicells; *M. fissurifera* Canu and Bassler, 1923 has frontal shields with fewer pseudopores, the ascopore is closer to the orifice and there are large, slit-like pore-chamber windows; *M. protea* Winston, 2005 has a pustulose frontal

shield with fewer, larger pseudopores, smaller autozooids and ovicells but larger avicularium; *M. proxima* Ramalho, Muricy and Taylor, 2011 has smaller autozooids, a nodular frontal shield with fewer, larger pseudopores, and avicularia that are more proximally placed.

Microporella cf. *pontifica* Osburn, 1952
Figure 39

cf. 1952 *Microporella pontifica* Osburn, p. 383, pl. 44, figure 5.

Figured material. UF 305814 (Shell 15); UF 305815 (Shell 30); UF 305816 (Shell 12). Pliocene, lower Tamiami Formation.

Description. Colony encrusting, multiserial, unilaminar, generally forming small spots (<20 zooids) but sometimes developing more extensive patches (50–100 zooids) (Figure 39.1). Pore-chamber windows visible at colony growing edge, numerous, closely spaced, rounded to elliptical, 40–60 µm long by 25–45 µm wide (Figure 39.1). Autozooids distinct with shallow interzooidal furrows and a thin lamina of smooth, raised calcification, rounded hexagonal, quincuncially arranged, slightly longer than wide (mean L/W 1.18). Frontal shield undulose, smooth and nodular, evenly and densely pseudoporous, except laterally to the orifice and laterally and distally to the avicularium (Figure 39.2); pseudopores 60–75, on average 65, circular, 15–18 µm in diameter; a few, scattered, marginal areolar pores adjacent to the zooidal margin, rounded to elliptical, 20–25 µm long; ascopore crescent-shaped, 50–60 µm in diameter, denticulate (Figure 39.4), placed in a depression just below the orifice and surrounded by a slightly raised collar, often more developed proximally. Orifice semicircular, with slightly denticulate proximal and distal margins, no condyles; a crenulated margin of cusps and lobes, the latter in the position of the oral spines, is formed by the frontal shield of the distal zooid (Figure 39.4). Oral spine bases 5–7, most commonly 5, 7 observed only in a few zooids, 20–30 µm in diameter. Avicularia adventitious, generally single, rarely absent or paired, located on either side of the autozoid, commonly placed at the lateral zooidal corner, at the same level as the ascopore or below, teardrop-shaped; rostrum funnel-shaped, channeled, directed distolaterally; crossbar complete. Ovicell embedded in the proximal part of the next distal zooid, nodular and evenly pseudoporous like the frontal shield but with smaller pseudopores (Figure 39.2–3); no spines visible in ovicellate zooids, possibly hidden by the development of a collar consisting of two lateral flaps that can fuse into a bridge (personate

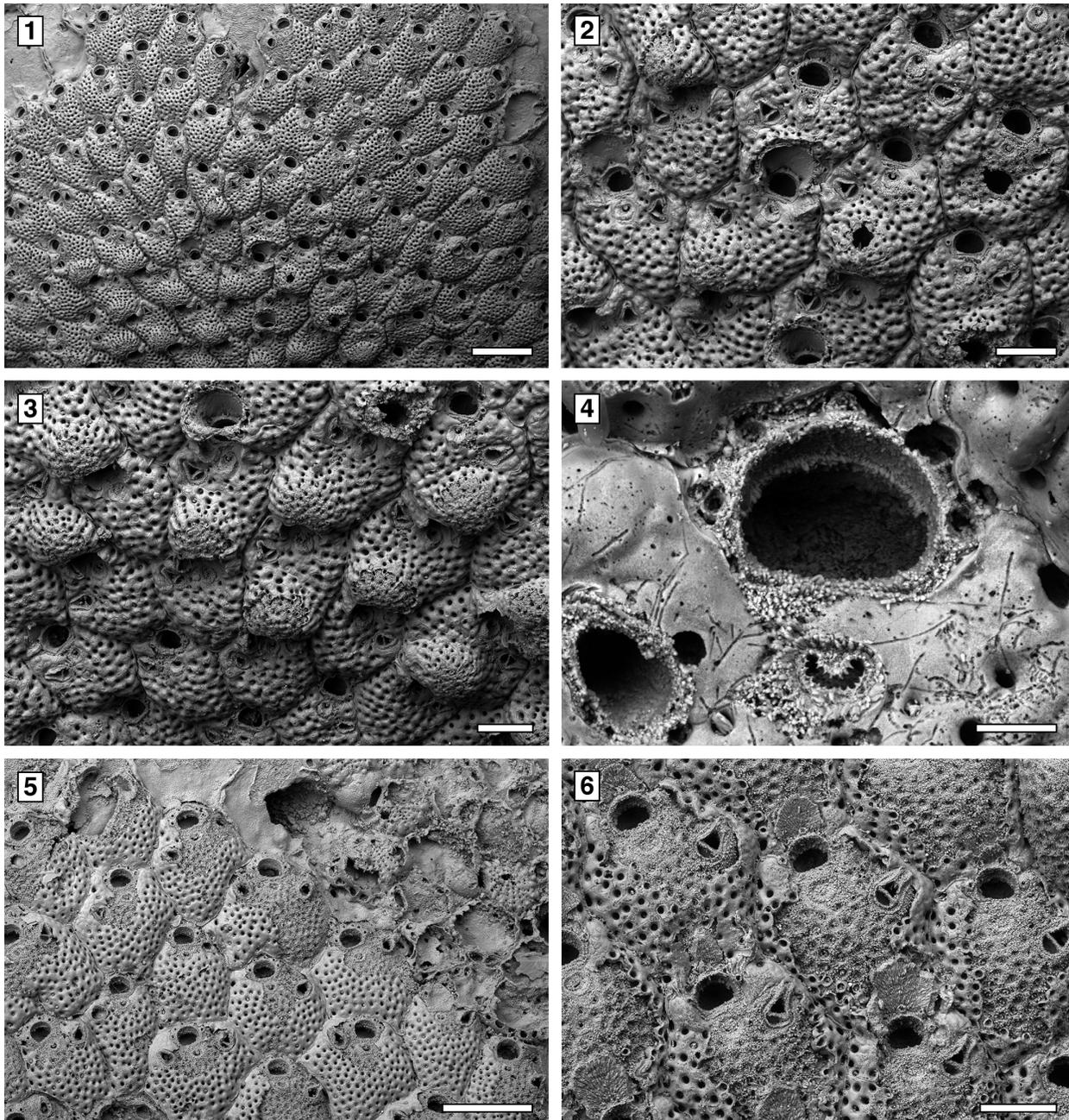


FIGURE 39. *Microporella cf. pontifica* Osburn, 1952, Pliocene, lower Tamiami Formation Units 10/11. **1–4**, UF 305814 (Shell 15). **1**, general view of part of the colony (500 μm). **2**, group of ovicellate and non-ovicellate zooids (200 μm). **3**, group of ovicellate zooids, one on the top left with a peristomial bridge (200 μm). **4**, close-up of the orifice (with cor-midial cusps and lobes in the position of oral spine bases) and denticulate ascopore (40 μm). **5**, UF 305815 (Shell 30), group of autozooids, some with paired avicularia (400 μm). **6**, UF 305816 (Shell 12), dissolution or abrasion of some zooids (200 μm).

ovicell) (Figure 39.3). Ancestrula similar to later autozooids but smaller, about 325 μm long by 220 μm wide, lacking an avicularium, budding one distal and two distolateral autozooids (335–390 μm long by 255–270 μm wide), surrounded by six autozooids in total.

Measurements (μm). ZL 476 \pm 28, 426–533 (2, 20); ZW 402 \pm 57, 304–505 (2, 20); OL 92 \pm 5, 83–100 (2, 15); OW 112 \pm 6, 103–121 (2, 15); AvL 108 \pm 8, 92–127 (2, 15); AvW 79 \pm 7, 66–94 (2, 15); OvL 211 \pm 20, 183–255 (2, 14); OvW 280 \pm 22, 250–321 (2, 14).

Remarks. This species is very similar in its general appearance to *Microporella pontifica* Osburn, 1952, a recent species found off the Pacific coast of Mexico and in the Gulf of California at 36–110 m depth. *Microporella pontifica* is similar to the fossil species in having up to six oral spines, orifice, and avicularium similar in size, and the peristome extended proximally across the orifice dividing the orifice from the ascopore. The latter feature seems constant in *M. pontifica*, while it has been observed only once in the Tamiami specimens, in which most often the peristome is limited to the lateral flaps, although in some instances it appears to have been broken off. *Microporella protea* Winston, 2005 found from Cape Hatteras to Florida also shares some similarities. Both species have a nodular frontal shield, a denticulate ascopore adjacent to the orifice, distal margin of the orifice with lobes and cusps formed by the next distal zooid (cormidial secondary orifice), and ovicellate zooids with projecting lateral flaps, which may or may not coalesce into a proximal bridge. However, the two species differ in several respects: the fossil species has a greater number of spines (5–7 vs 3–4), a larger orifice, and smaller avicularium.

Microporella cf. *pontifica* differs from its congeners from the Tamiami Formation in having a greater number of oral spines, ascopore placed closer to the orifice, and nodular frontal shield. All three *Microporella* species found in the Tamiami Formation assemblage show the same type of preservation, with the external layer of the frontal surface dissolved or abraded (Figures 37.7, 38.6, 39.5–6).

Superfamily Celleporoidea Johnston, 1838

Family Celleporidae Johnston, 1838

Genus *Pourtalesella* Winston, 2005

Pourtalesella chiarae sp. nov.

Figure 40

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Type material. Holotype UF 305817 (Shell 3); paratypes UF 305818 (Shell 25), UF 305819 (Shell 26), UF 305820 (Shell 119). Pliocene, lower Tamiami Formation.

Etymology. Named after Dr. Chiara Lombardi (ENEA, Marine Environment Research Centre, La Spezia, Italy) for her significant contributions in the field of bryozoan biomineralization.

Diagnosis. Colony encrusting, often multiserial; autozooids rounded polygonal. Pore-chamber windows present. Frontal shield pseudoporous proximally and centrally, averaging 30 pseudopores per zooid. Orifice with robust rounded condyles, semi-

circular anter and rounded V-shaped or broadly U-shaped sinus; oral spines absent. Avicularia adventitious, commonly paired, located proximolaterally to the orifice, directed distolaterally; crossbar complete. Ovicells globular; ectooecium incompletely calcified exposing an imperforate endooecium. Ancestrula with a well-developed, imperforate, smooth gymnocyst and a transversely D-shaped orifice.

Description. Colony encrusting, multiserial, uni- or multilaminar, extensive (>200 zooids) (Figure 40.1). Pore-chamber windows visible at colony growing edge (Figure 40.1). Autozooids distinct only laterally with shallow depressions, becoming indistinct through ontogeny, quincuncially arranged, rounded polygonal, longer than wide (mean L/W 1.45). Frontal shield slightly convex, granular, pseudoporous for about two-thirds of its length starting from the proximal margin, imperforate for the remaining one-third corresponding to the area surrounding the avicularia and orifice (Figure 40.2); pseudopores circular, 10–15 µm in diameter, numbering 24–36 per zooid, on average 30. Orifice almost as long as wide, two robust rounded condyles separating a semicircular anter from a slightly narrower rounded V-shaped or broadly U-shaped sinus (Figure 40.3). Oral spine bases absent. Avicularia adventitious, small, commonly paired, rarely single, the swollen cystid lateral to the orifice with the tip of the rostrum at about the same level as the proximal margin of the orifice, elliptical with complete crossbar, distolaterally directed, sloping proximally and convergent inwardly (Figure 40.2). Ovicells globular, initially prominent, recumbent on and later during ontogeny immersed on the frontal shield of the next distal zooid (Figure 40.4–7); ectooecium incompletely calcified forming a narrow band; endooecium largely exposed, imperforate, with an uncertain number of radial ribs. Ancestrula small, very different in appearance from the budded zooids, oval, 215 µm long by 180 µm wide, with a well-developed, imperforate, smooth gymnocyst and a transversely D-shaped orifice, 70 µm long by 110 µm wide, surrounded by a slightly raised, striated rim about 15 µm wide (Figure 40.8–9); presence of spines likely but uncertain owing to recrystallization; one distal and two distolateral zooids budded directly from the ancestrula.

Measurements (µm). ZL 438±34, 374–487 (2, 20); ZW 302±29, 261–361 (2, 20); OL 116±6, 106–124 (2, 10); OW 111±7, 104–125 (2, 10); AvL 54±5, 48–64 (2, 10); AvW 45±5, 38–55 (2, 10); OvL 186±8, 174–198 (2, 8); OvW 221±19, 192–246 (2, 8).

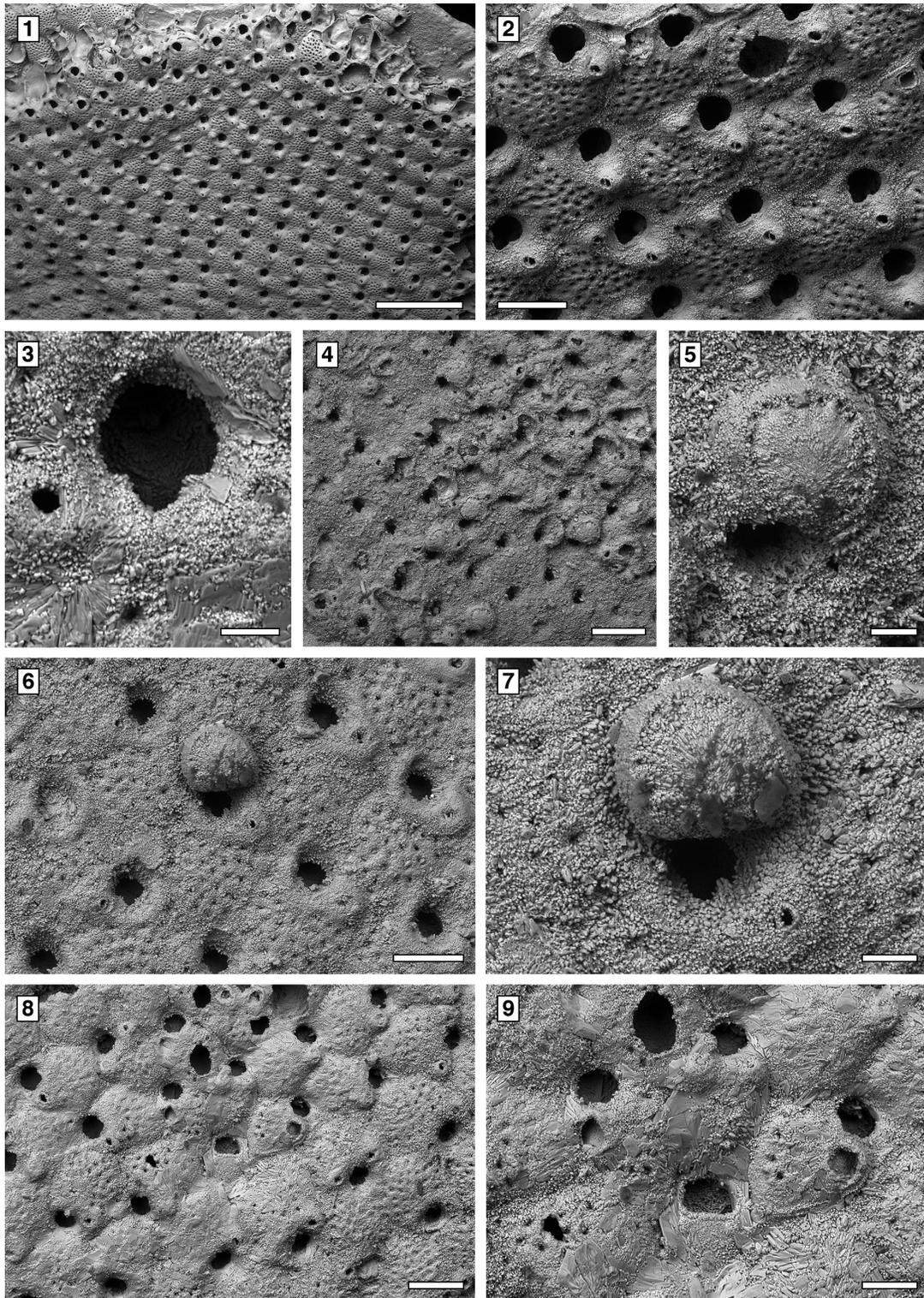


FIGURE 40. *Pourtalesella chiarae* sp. nov., Pliocene, lower Tamiami Formation Units 10/11. **1–2**, holotype UF 305817 (Shell 3). **1**, general view of the multilaminar colony (1 mm). **2**, group of autozooids (200 μ m). **3–5**, paratype UF 305818 (Shell 25). **3**, close-up of an orifice (40 μ m). **4**, group of ovicellate and non-ovicellate zooids (400 μ m). **5**, close-up of an ovicell (60 μ m). **6–7**, paratype UF 305819 (Shell 26). **6**, group of zooids, one ovicellate and one with a closure plate (200 μ m). **7**, close-up of an ovicell (60 μ m). **8–9**, paratype UF 305820 (Shell 119). **8**, ancestrula and early astogeny (200 μ m). **9**, close-up of the ancestrula and first budded autozooids (100 μ m).

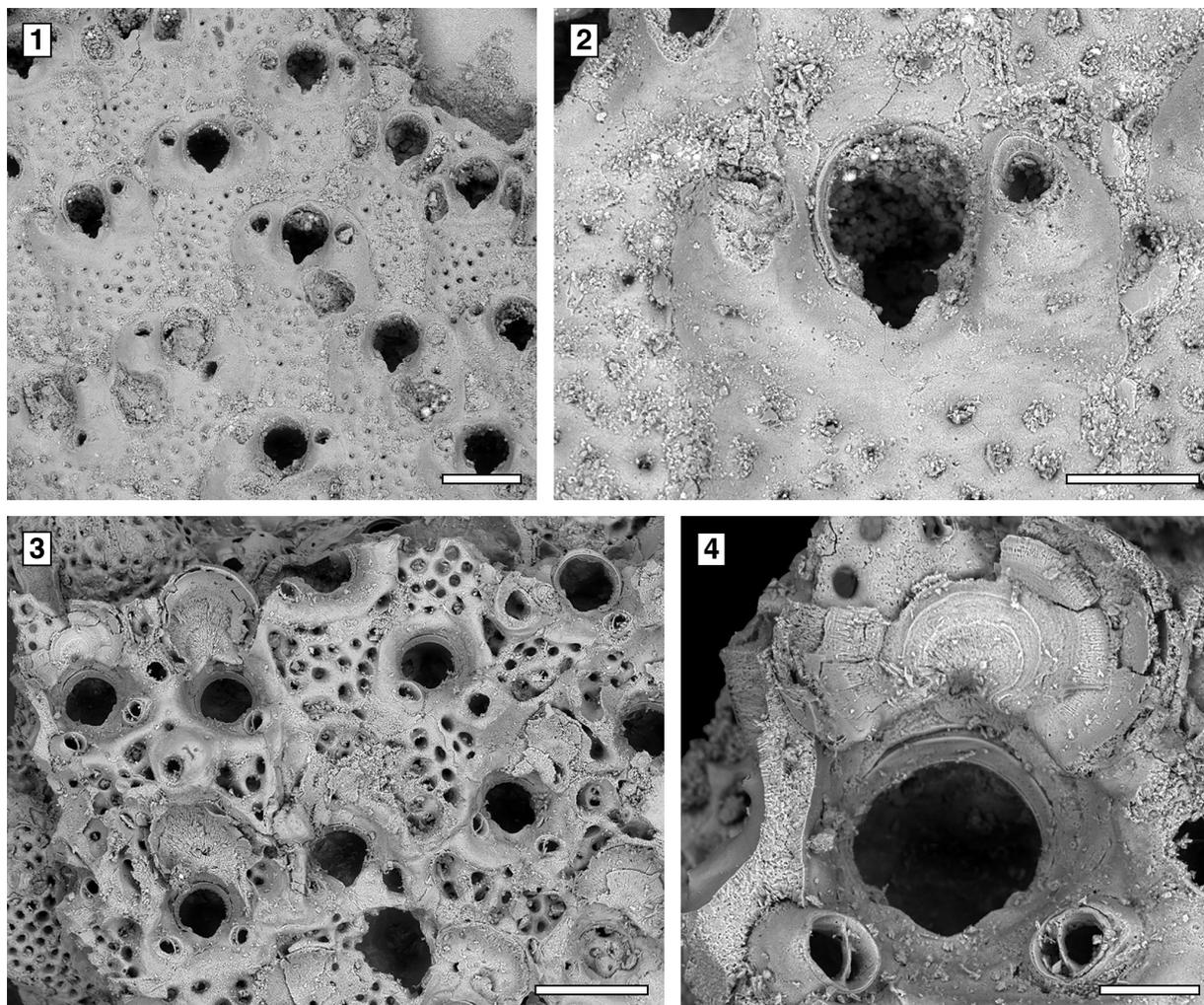


FIGURE 41. *Stephanosella biaperta* Michelin, 1842 sensu Canu and Bassler (1923). **1–2**, USNM 68544. **1**, group of autozooids (200 μ m). **2**, close-up of the orifice (100 μ m). **3–4**, USNM 68546. **3**, group of zooids, some with incomplete ovicells (200 μ m). **4**, close-up of an orifice, lateral avicularia and incomplete ovicell (50 μ m).

Remarks. *Pourtalesella* was introduced by Winston (2005) for species very closely related to *Bufonellaria*, but with a pseudoporous frontal shield. Four species are currently attributed to *Pourtalesella*, all recent and all from the western Atlantic. *Pourtalesella incrassata* (Canu and Bassler, 1928a) from the Gulf of Mexico differs from the Tamiami Formation species in having fewer (about 20), larger pseudopores and avicularia with serrated rostra; *P. rugosa* (Osburn, 1940) from Puerto Rico differs also in having fewer pseudopores, adventitious avicularia directed proximolaterally and calcification that during ontogeny covers the frontal shield with high irregular ridges leaving only a central area through which the pseudopores may be seen. *Pourtalesella carvalhoi* (Marcus, 1937) from Bahia (Brazil) differs in having a small avicularium embedded within the peristome and ridged

endoecium. Finally, *P. alipioi* (Marcus, 1955) from Espirito Santo (Brazil) is likely to have been incorrectly placed in this genus as it has oral spines and a perforate endoecium. Canu and Bassler's (1923, pl. 16, figures 4–9) *Stephanosella biaperta* Michelin, 1842 from the Neogene of North America and Jamaica may seem similar in appearance to *Pourtalesella chiarae* sp. nov. However, after SEM examination, it appears that the specimens figured by Canu and Bassler (1923) (USNM 68544 and USNM 68546) belong to two different species, both differing from *Pourtalesella chiarae* sp. nov. USNM 68544 (Figure 41.1–2) seems to be an undescribed species of *Pourtalesella*, which we defer formally naming because it lacks ovicells; it differs from the Tamiami Formation species in having adventitious avicularium placed much more distally and an orifice with a much narrower sinus. USNM

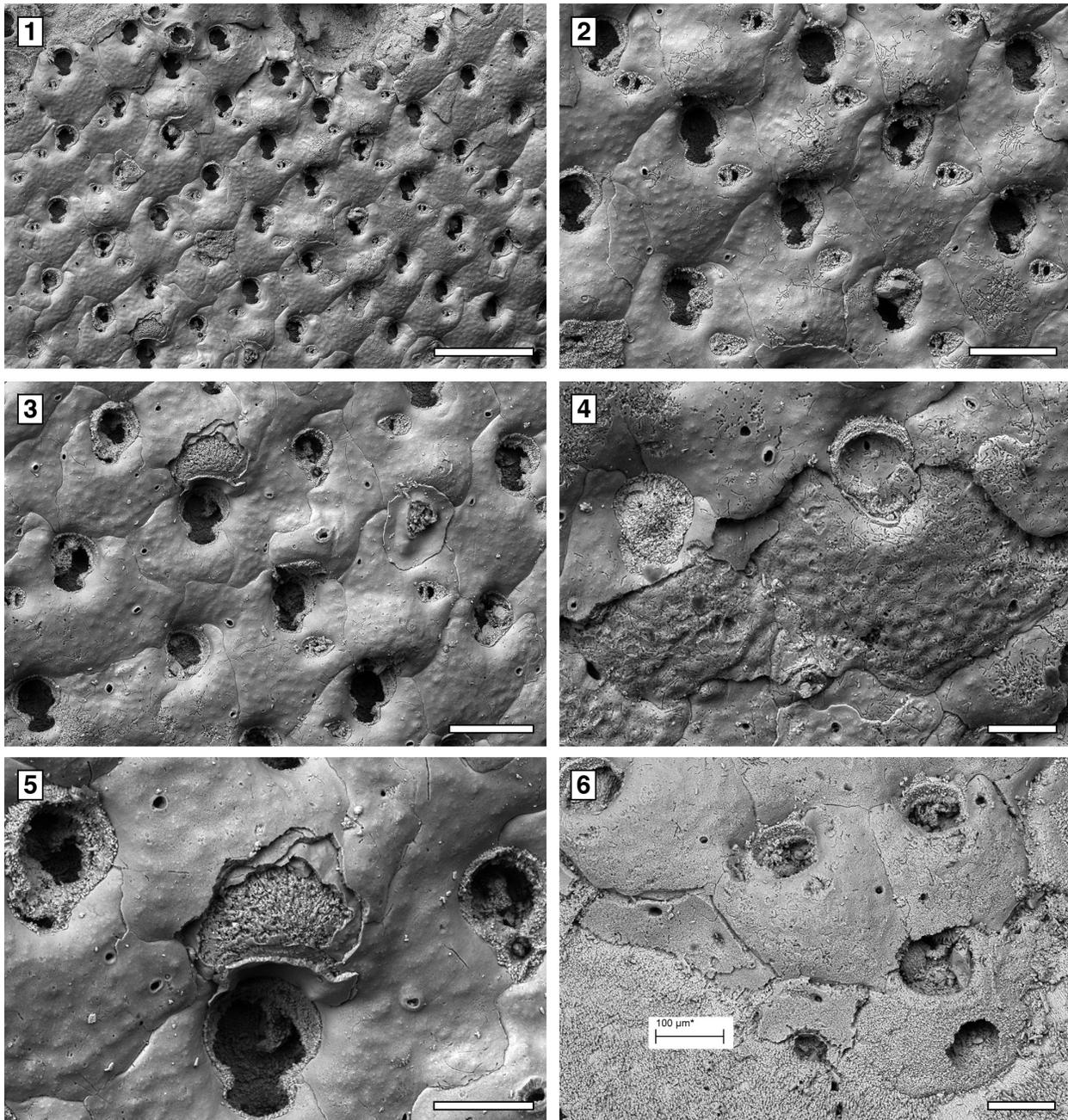


FIGURE 42. *Plesioleidochasma* cf. *vestitum* (Canu and Bassler, 1923), Pliocene, lower Tamiami Formation Units 10/11. **1–5**, UF 305821 (Shell 14). **1**, group of ovicellate and non-ovicellate zooids (500 μm). **2**, close-up of three ovicellate zooids (200 μm). **3**, group of zooids with secondary calcification spreading from marginal areolar pores (200 μm). **4**, autozooids with closure plates (100 μm). **5**, damaged ovicell showing the superposition of calcareous layers (100 μm). **6**, UF 305822 (Shell 11), tatiform ancestrula and first budded zooids (100 μm).

68546 (Figure 41.3–4) is likely to be a specimen of *P. rugosa* because of the characteristic ridges on the frontal shield developed through secondary calcification during ontogeny.

Family Phidoloporidae Gabb and Horn, 1862
Genus *Plesioleidochasma* Soule, Soule, and

Chaney, 1991
Plesioleidochasma cf. *vestitum* (Canu and
Bassler, 1923)
Figure 42

cf. 1923 *Hippoporina* (?) *vestita* Canu and Bassler, p.
130, figures 7–205.

Figured material. UF 305821 (Shell 14); UF 305822 (Shell 11). Pliocene, lower Tamiami Formation.

Description. Colony encrusting, multiserial, unilaminar (Figure 42.1). Autozooids distinct, with shallow interzooidal furrows, becoming indistinct later in ontogeny as a result of secondary calcification spreading from the marginal areolar pores, quincuncially arranged, hexagonal, almost as long as wide (mean L/W 1.08). Frontal shield slightly convex distally just below the orifice, rarely produced into a rounded, raised umbo, nodular, imperforate apart from one or two, rarely four, very small, circular areolar pores, 10–15 μm in diameter, located at lateral zooidal corners, almost symmetrically placed when paired (Figure 42.2–3). Orifice longer than wide, cleithridiate with two small, rounded condyles separating a deeply arched horseshoe-shaped anter from a shallow, narrower poster; oral spine bases observed only in early astogenetic zooids. Avicularia adventitious, teardrop-shaped, small, usually single but absent in some autozooids, budded from one of the two lateral marginal pores, placed approximately horizontally and directed laterally outwards; rostrum acutely triangular, rounded and crossbar complete. Ovicells globular, imperforate, becoming quickly immersed in the frontal shield of the next distal zooid (Figure 42.2, 5). Closure plates common, smooth with a small, circular opening distally (Figure 42.4). Secondary calcification spreading copiously from the marginal areolar pores obliterating the zooidal boundaries (Figure 42.3–5). Ancestrula tatiform with an uncertain number of spine bases and extensive proximal gymnocrystal walls sloping gently outwards, budding one distal and one (possibly two) distolateral zooids (Figure 42.6). First budded zooids smaller than later autozooids, lacking avicularia and with three, small, oral spine bases.

Measurements (μm). ZL 353 \pm 25, 311–403 (1, 20); ZW 326 \pm 37, 235–400 (1, 20); OL 133 \pm 4, 127–142 (1, 15); OW 110 \pm 5, 100–118 (1, 15); AvL 80 \pm 6, 64–87 (1, 15); AvW 56 \pm 4, 50–61 (1, 15); OvL 157 \pm 10, 143–172 (1, 8); OvW 194 \pm 18, 169–213 (1, 8).

Remarks. Canu and Bassler (1923) tentatively attributed this species to the genus *Hippoporina*. From the images they provided (pl. 18, figures 7–9), it would now be placed in *Plesiocleidochasma*. The specific name ‘*vestitum*’ derives from the main character attributed by the authors to the species (p. 130) “The zooecia [...] covered by two or three superposed calcareous *pellicles*”. The same character is present in our species, in which copious

secondary layers of calcification spread from the marginal areolar pores. In addition, these superimposed ‘calcareous pellicles’ are clearly visible in the broken ovicell depicted in Figure 42.5. Thus, we refer the Tamiami Formation species to *Plesiocleidochasma* cf. *vestitum*. The holotype of *P. vestitum* was described from the Choctawhatchee Marl (now assigned to the upper Pliocene Jackson Bluff Formation) of Florida. Unfortunately, the type specimen lacks avicularia and complete ovicells.

DISCUSSION

The bryozoan assemblage found encrusting *Anomia simplex* shells in Units 10/11 of the upper Pliocene lower Tamiami Formation in Florida is described here for the first time. The fauna is relatively low in diversity, comprising 29 species in 26 genera and 22 families, and including a single cyclostome and 28 cheilostomes (nine anascan- and 19 ascophoran-grade). *Acanthodesia* and *Microporella* are the only two genera represented by more than one species, specifically two and three species, respectively. Two species of *Microporella* are new to science. In addition to *Microporella*, the fauna includes new species of *Micropora*, *Spiniflabellum*, *Trypostega*, and *Pourtalesella*, making a total of six new species. Only two families are represented by more than a single genus, i.e., Calloporidae with four genera and Cribriliinidae with two genera. Eighteen of the bryozoans are established taxa identifiable to the species level, although some only tentatively. Among these, six species are extant and have western Atlantic distributions, while 12 species are known only as fossils, with the earliest record dating back to the early Miocene. Five species can be identified only to genus level owing to deficient preservation and a scarcity of available material.

The bulk of the assemblage is accounted for by five species that are very abundant (>500 colonies), while nine species are very rare (<10 colonies) (see Table 1). Most species form extensive, sheet-like colonies consisting of hundreds of zooids and occasionally completely enveloping the shell substrate. Six species have multilayered, flat colonies, and a single species forms multilaminar, mounded colonies.

Echols (1960) identified 26 species of bryozoans (one cyclostome and 25 cheilostomes) from other exposures of the Tamiami Formation. No images are available in his unpublished Masters dissertation, making any comparisons with the fauna described here difficult. Only two of the species mentioned by Echols (1960) – *Aplousina gran-*

dis and *Cyclocolpota perforata* – were also found encrusting the *Anomia* shells. However, it is likely that the two assemblages share more than just these two species. For instance, Echols (1960) described three species of *Microporella*, two of which he identified as *M. ciliata* (Pallas) and *M. marsupiata* (Busk), species that, before the advent of the SEM, were recorded worldwide, while the third species, *M. pontifica* Osburn, might correspond to our *M. cf. pontifica* (see Remarks).

Outer surfaces of the *Anomia* shells are encrusted in 96% of the shells analyzed, while inner surfaces are encrusted in 58% of the shells analyzed; however, 25% of the shells have adherent sediment that prevent observation of the entire inner surface. The high proportion of shells encrusted on their inner surfaces is consistent with post-mortem colonization of the shells.

Although the most abundant epibionts, bryozoans are not the only taxonomic group encrusting the *Anomia* shells, which host lesser numbers of the coral *Astrangia*, barnacles, foraminifera, and spirorbid polychaetes. A small number of interactions are present between bryozoan colonies and these groups of organisms, while a much larger number of interactions can be observed between the bryozoans, both intra- and interspecific. This taxonomic work establishes the necessary baseline for ranking the different bryozoan species in a

competitive hierarchy, which will be the subject of a future palaeoecological study.

ACKNOWLEDGMENTS

We thank L.J. Cotton (University of Bristol) who helped collecting the specimens; M. Spencer-Jones (Department of Life Sciences, Natural History Museum, London) for arranging the loan of the type specimens in her care; J. Sanner (Department of Paleobiology, National Museum of Natural History, Smithsonian Institution, Washington, DC, USA) for providing SEM images of Canu and Bassler's material in her care. T. Perkins (Volunteer Program, Natural History Museum London) helped processing the samples. E. Henshaw, Jr. (President) of Schroeder-Manatee Ranch (SMR) Aggregates, Inc. kindly allowed access to the quarry. S. Roberts (Florida Museum of Natural History, University of Florida) provided assistance with Figures 1, 2, and 3.

This research was funded by the Leverhulme Trust, Research Project Grants 'Origin of high tropical diversity: a test using bryozoans', award no. RGP-2015-036 and the Florida Museum of Natural History, Vokes and Florida Fossil Hunters scholarships. This is University of Florida Contribution to Paleobiology 856. Four anonymous reviewers provided very useful comments that greatly improved the originally submitted manuscript.

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